Pressure Independence of Renin Release by Isolated Kidneys of Lyon Hypertensive Rats

Isac A. Medeiros, Madeleine Vincent, Daniel Benzoni, and Jean Sassard

In the present work the influence of perfusion pressure on renal functions and renin release was studied before and after the blockade of thromboxane A<sub>2</sub>/prostaglandin H<sub>2</sub> (TXA<sub>2</sub>/PGH<sub>2</sub>) receptors using isolated kidneys from 7-week-old genetically hypertensive (LH), normotensive (LN), and low blood pressure (LL) rats of the Lyon strain. Kidneys were single pass perfused with Krebs-Henseleit solution with a gelatine derivative (Polygeline) added as an oncostic agent. A servocontrolled system stabilized the renal perfusion pressure (RPP) at any chosen (±1 mm Hg) level. In baseline conditions (RPP, 90 mm Hg), LH (n=7) kidneys differed from LN (n=6) and LL (n=8) controls by increased vascular resistance, decreased glomerular filtration rate, and natriuresis. The LH kidney responses to stepwise changes in RPP (between 60 and 170 mm Hg) differed from those of LN and LL rats by a significantly lower perfusion flow, glomerular filtration rate, and natriuresis. Above all, the reduction in RPP, which induced a marked and highly reproducible renin release in LN and LL kidneys, was devoid of effects in LH kidneys. The blockade of TXA<sub>2</sub>/PGH<sub>2</sub> receptors by AH23848 (4 x 10<sup>-8</sup> M) did not change the baseline (RPP, 90 mm Hg) functions of kidneys of the three strains. During changes in RPP, the responses of LN and LL kidneys were not modified, whereas LH kidneys exhibited significant increases in both glomerular filtration rate and natriuresis. Finally, AH23848 significantly decreased the renin release by kidneys of the three strains. It is concluded that renin release of LH isolated kidneys is independent of the perfusion pressure and that renal TXA<sub>2</sub>/PGH<sub>2</sub> receptor activation participates in the renin release and in the altered functions exhibited by kidneys of LH rats.

KEY WORDS • kidney • renin • natriuresis • thromboxane A<sub>2</sub> • renal function • Lyon rat • genetic hypertension

Several recent experiments using kidney transplantation<sup>1,2</sup> have emphasized the primary role played by the kidney in the pathogenesis of genetic hypertension. Obviously, among the pressor factors originating in the kidney, the most important is the renin-angiotensin system. In Lyon genetically hypertensive (LH) rats, the plasma renin activity is normal in young animals and low in adult ones.<sup>3</sup> Similar findings have been reported in Japanese spontaneously hypertensive rats (SHR).<sup>4</sup> Despite these low levels of plasma renin, both models are highly sensitive to the blockade of the renin-angiotensin system.<sup>5,6</sup> This demonstrates that, in genetically hypertensive rats, the low renin levels remain inappropriate and actively contribute to blood pressure elevation.

Such an inappropriate renin secretion may reflect a polymorphism of the renin gene, as found in Dahl salt-sensitive rats,<sup>7</sup> or an abnormal control by its physiological regulators. Because the renin gene polymorphism in LH rats does not appear directly related to the blood pressure level,<sup>8</sup> we thought it of interest to assess the control of renin secretion by one of its major regulatory factors, the renal perfusion pressure (RPP). In addition, since in previous experiments we have shown that kidneys of LH rats synthesized an excess of thromboxane A<sub>2</sub> (TXA<sub>2</sub>),<sup>9</sup> a vasoconstrictor prostanoid that may be involved in renin secretion,<sup>10</sup> the baroreceptor control of renin secretion was studied before and after blockade of TXA<sub>2</sub>/prostaglandin H<sub>2</sub> (PGH<sub>2</sub>) receptors. All these experiments were conducted in isolated perfused kidneys from LH rats and its two control strains.<sup>11</sup>

Methods

Animals

Seven-week-old male LH, normotensive (LN), and low blood pressure (LL) rats of the Lyon strain were used. Animals were housed under constant conditions of temperature (21 ± 1°C), lighting (8 AM to 8 PM), and humidity (60±10%). They were fed a standard diet (Eleveage UAR, Villemoisson s/ Orge, France) and had free access to tap water. Systolic blood pressure (SBP) was measured on 2 consecutive days before the experiments by a plethysmographic method (Narco BioSystems, Austin, Tex.) in the prewarmed (37°C for 10 minutes), unrestrained conscious rats.

Kidney Preparations

In sodium pentobarbital (45 mg/kg i.p.)-anesthetized rats, the right kidney was isolated according to Schmidt
FIGURE 1. Line plots show renin release (RR) from Lyon normotensive isolated kidneys: Influence of the duration and the pressure of perfusion (RPP). Al, angiotensin I. *p<0.05 vs. values obtained at RPP of 120 mm Hg.

Kidney Perfusion

The perfusion solution was a blood-free modified Krebs-Henseleit solution containing 35 g/l of a gelatine derivative (Polygeline, Behring, Marburg, FRG) as an oncotic agent. The composition of the perfusion medium has been described elsewhere. Just before use, the perfusate was filtered through 2 millipore filters (1.2 μm and 0.8 μm, successively) and 0.5 g/l polyfructosan was added (Inutest, Laevosan, Linz, Austria). This solution, maintained at 37°C, was continuously bubbled with a 95% O₂-5% CO₂ mixture and single pass perfused using a peristaltic pump (Minipuls 2, Gilson, Paris). The RPP (mm Hg), monitored through a pressure transducer (model P231D, Statham Instrument Division, Gould Inc., Oxnard, Calif.), fed the pump through a specially designed device that stabilized the pressure at any chosen level (± 1 mm Hg) by servocontrolling the perfusion flow.

Renal Function Parameters

Renal perfusion flow (RPF) (ml/min/g) and urinary flow rate were measured by weighting. Renal vascular resistances (mm Hg/ml/min/g) were calculated as the ratio RPP/RPF. Glomerular filtration rate (GFR) (ml/min/g) was measured by polyfructosan clearance (Technicon) and sodium concentration by flame photometry (IL meter 243). Renin concentration in the venous effluent was measured by the radioimmunoassay of angiotensin I (Ang I) generated during incubation at pH 6.5 with binephrectomized rat plasma used as substrate according to Menard and Catt.14 For each venous sample, Ang I generation was measured after three incubation times to ensure the excess of substrate. Renin release (ng Ang I/hr/min) was calculated by multiplying renin concentration by the flow rate.

Protocols

Influence of renal perfusion pressure. Preliminary studies using LN kidneys demonstrated (Figure 1) that renin release from isolated kidneys perfused at 80 mm Hg remained stable for 2 hours and reductions in RPP from 120 to 50 mm Hg induced a rapid and reproducible renin release, which returned to baseline within 15 minutes after reestablishing RPP at 120 mm Hg. According to these data, the influence of RPP on the renin release was studied as follows: after a 30-minute stabilization period during which the kidneys were perfused at 90 mm Hg, venous effluent and urine samples were obtained to determine the baseline values. Then RPP was set at 170 mm Hg and was progressively decreased to 150, 130, 110, 85, 80, 75, and 60 mm Hg. Each level of RPP was maintained for 5 minutes. Venous samples were drawn after 4 minutes, and urine samples were collected during the last 4 minutes. After weighting, samples were kept at −20°C until assay.

Effects of a TXA₂/PGG₂ Receptor Blockade on Renal Response to Changes in Perfusion Pressure. At the end of the previous protocol, the kidneys were perfused with AH23848 (a gift from Glaxo, Greenford, England), a TXA₂/PGG₂ antagonist,15 at a final concentration of 4×10⁻⁶ M according to Liu et al.13 After a 15-minute stabilization period (RPP, 90 mm Hg), RPP was step-wise changed from 170 to 60 mm Hg according to the protocol described above. Venous effluent and urine...
samples were obtained and processed similarly. At the end of the study, the servocontrol of RPP was stopped, and the efficiency of the receptor blockade was demonstrated by the virtual disappearance of renin release from kidneys of Lyon hypertensive rats.

### Results

#### Baseline Characteristics

Seven-week-old LH rats exhibited a significantly higher indirect SBP (Table 1) than the LN and LL controls, both of which, as usual, did not differ in that respect. Since the body and kidney weights were higher in LH rats than in LN and LL controls, all the parameters were corrected for kidney weight. It must be emphasized that the interstrain differences in kidney weight were too small to account for the marked changes in renal functions described below. After 30 minutes of equilibration, the isolated kidneys perfused at a pressure of 90 mm Hg exhibited marked interstrain differences. LL kidneys differed from LN by a significantly lower urinary sodium excretion ($U_{UN,V}$). LH kidneys, compared with LN and LL controls, exhibited increased vascular resistance and lower renin release and GFR. This latter accounted for most of their low natriuresis since the sodium reabsorption rate (% of filtered sodium) did not differ among strains.

#### Influence of Perfusion Pressure on Renal Function and Renin Release

As shown by Figure 2, any increase in RPP was associated with increases in the perfusate flow and decreases in sodium reabsorption rate, which were identical in LN and LL kidneys. GFR and natriuresis increased also but more markedly in LH kidneys. Since the sodium reabsorption rate did not differ between these two strains, the more marked pressure–natriuresis observed in LN rats is related to an elevated filtration of sodium. Although the trend of the pressure effects was similar in LH kidneys, these latter strikingly differed from both LN and LL controls by a much lower elevation in flow, GFR, and natriuresis. At any perfusion pressure level above 130 mm Hg the

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**Table 1. Baseline Values Obtained in 7-Week-Old Rats of the Lyon Strains Before and After Thromboxane A2/Prostaglandin H2 Receptor Blockade With AH23848**

<table>
<thead>
<tr>
<th>Parameters</th>
<th>LL (n=8)</th>
<th>LN (n=6)</th>
<th>LH (n=7)</th>
</tr>
</thead>
<tbody>
<tr>
<td>SBP (mm Hg)</td>
<td>119±3</td>
<td>119±3</td>
<td>150±5*†</td>
</tr>
<tr>
<td>BW (g)</td>
<td>155±5</td>
<td>138±7</td>
<td>191±8*†</td>
</tr>
<tr>
<td>LKW (g)</td>
<td>0.73±0.02</td>
<td>0.77±0.03</td>
<td>0.89±0.05*†</td>
</tr>
<tr>
<td>LKW/BW (%)</td>
<td>0.47±0.02*</td>
<td>0.56±0.03</td>
<td>0.47±0.03</td>
</tr>
<tr>
<td>Control</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>RVR (mm Hg/ml/min/g)</td>
<td>8.2±0.3</td>
<td>8.3±0.4</td>
<td>14.9±0.9*†</td>
</tr>
<tr>
<td>GFR (ml/min/g)</td>
<td>0.47±0.02</td>
<td>0.54±0.03</td>
<td>0.40±0.01*†</td>
</tr>
<tr>
<td>$U_{UN,V}$ (µeq/min/g)</td>
<td>7.7±0.5*</td>
<td>12.7±2.1</td>
<td>5.9±0.5*†</td>
</tr>
<tr>
<td>$R_{Na}$ (%)</td>
<td>89±1</td>
<td>85±2</td>
<td>89±1</td>
</tr>
<tr>
<td>RR (ng Ang I/hr/min)</td>
<td>534±44</td>
<td>561±37</td>
<td>176±19*†</td>
</tr>
<tr>
<td>AH23848 (4×10⁻⁸ M)</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>RVR (mm Hg/ml/min/g)</td>
<td>7.1±0.2</td>
<td>7.6±0.4</td>
<td>12.1±0.7*†</td>
</tr>
<tr>
<td>GFR (ml/min/g)</td>
<td>0.49±0.03</td>
<td>0.60±0.07</td>
<td>0.37±0.02*†</td>
</tr>
<tr>
<td>$U_{UN,V}$ (µeq/min/g)</td>
<td>10.4±1.3*</td>
<td>16.2±1.1</td>
<td>7.5±0.5*†</td>
</tr>
<tr>
<td>$R_{Na}$ (%)</td>
<td>85±2</td>
<td>80±3</td>
<td>86±1</td>
</tr>
<tr>
<td>RR (ng Ang I/hr/min)</td>
<td>445±42</td>
<td>523±32</td>
<td>178±14*†</td>
</tr>
</tbody>
</table>

Values are mean±SEM. LL, Lyon low blood pressure rats; LN, Lyon normotensive rats; LH, Lyon hypertensive rats; SBP, systolic blood pressure; BW, body weight; LKW, left kidney weight; RVR, renal vascular resistance; GFR, glomerular filtration rate; $U_{UN,V}$, urinary sodium excretion; $R_{Na}$, sodium reabsorption rate; RR, renin release; Ang I, angiotensin I.

*p<0.05 vs. LL.

†p<0.05 vs. LN.

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**Statistical Analysis**

Values are mean±SEM. One-way analysis of variance (ANOVA) was used to determine the influence of the length of the perfusion. The Mann-Whitney test was used to assess the interstrain differences and the Wilcoxon test to evaluate the effects of AH23848 infusion.
Glomerular Filtration rate

Natriuresis

Perfusion Pressure (mmHg)

**FIGURE 2.** Line plots show influence of perfusion pressure on the function of kidneys from Lyon low blood pressure (LL) (○, n=8), Lyon normotensive (LN) (○, n=6), and Lyon hypertensive (LH) (△, n=7) 7-week-old rats. *p<0.05 vs. both LL and LN; †p<0.05 vs. LN. RPF, renal perfusion flow; RNa, sodium reabsorption rate; GFR, glomerular filtration rate; UNaV, urinary sodium excretion.

sodium reabsorption of LH kidneys was higher than that of LL kidneys.

The relation between RPP and renin release is shown in Figure 3. In LN and LL kidneys the maximum release was observed at RPP of 85 mm Hg and then progressively decreased when RPP was elevated up to 170 mm Hg. Surprisingly, there was no evidence of this well-known baroreceptor control of renin release in LH kidneys. The renin release of LH kidneys remained stable at values significantly lower than those of LN and LL kidneys up to 110 mm Hg and reached higher values at a RPP level of 170 mm Hg.

**Effects of TXA2/PGH2 Receptor Blockade**

Table 1 indicates that, in baseline conditions, a 15-minute infusion of AH23848 (4×10^-6 M) did not significantly alter the characteristics of the kidneys of the three strains. However in the three strains, vascular resistances, renin release, and sodium reabsorption rate tended to decrease while the urinary excretion of sodium slightly increased. As illustrated in Figure 4, the overall effects of AH23848 were to increase GFR and natriuresis and to decrease renin release in response to changes in RPP. In LN kidneys, only the renin release was significantly decreased by AH23848. In LL kidneys this decrease in renin release was more marked and was associated with an enhanced natriuresis at the perfusion pressure of 170 mm Hg. On the other hand, LH kidneys infused with AH23848 exhibited significant increases in GFR and natriuresis. The increase in natriuresis was more marked than that of GFR and therefore was partly accounted for by a decreased sodium reabsorption. Finally, AH23848 significantly lowered the renin release of LH kidneys, this latter remaining independent of the pressure level.

**Influence of Isoproterenol on Renin Release From Kidneys of Lyon Hypertensive Rats**

While the perfusion pressure was maintained at 90 mm Hg, the infusion of IPNA (2×10^-5, 2×10^-4, and
FIGURE 4. Line plots show influence of thromboxane A2 /prostaglandin H2 receptor blockade on the perfusion pressure effects on the function of kidneys isolated from Lyon low blood pressure (LL), Lyon normotensive (LN), and Lyon hypertensive (LH) 7-week-old rats. Control (open symbols); blockade (closed symbols). GFR, glomerular filtration rate; UNaV, urinary sodium excretion; RR, renin release; AI, angiotensin I. *p<0.05 vs. controls.

2×10⁻⁷ M) in kidneys from 7-week-old LH rats (n=4) induced a concentration-dependent renin release: from 147±29 before to 535±110, 1,086±145, and 1,260±218 ng Ang I/hr/min after the three concentrations of IPNA, respectively. The experiment was repeated with a perfusion solution free of calcium containing EGTA. A similar concentration-dependent renin release was observed but at higher values than in the presence of calcium: from 437±60 before to 761±99, 1,147±183, and 1,475±322 ng Ang I/hr/min after the three same concentrations of IPNA, respectively.

Discussion

The major finding of the present work is that the renin release of isolated perfused kidneys from young genetically hypertensive rats of the Lyon strain is not influenced by perfusion pressure as it is in kidneys of their normotensive controls. The present study was conducted to determine whether the baroreceptor control of renal renin could be abnormal in LH rats and thus lead to an inappropriate secretion that could contribute to the renin dependence of high blood pressure in that model.

Thus, we chose the isolated single pass perfused kidney preparation. Such an isolated organ is suitable to study the specific effects of perfusion pressure on renin release since it is free from sympathetic innervation and the perfusion medium is of known constant composition, and does not contain any stimulus or any component of the renin-angiotensin system. We designed a servocontrolled perfusion system to reach stable (±1 mm Hg) levels of RPP. With the perfused solution used, the sodium reabsorption rate was lower and the renal flow higher than in vivo. These findings are in accordance with those of others using similar preparations16 and are due to the relatively low oncotic pressure17 and viscosity18 of the perfusion solution compared with blood.

Kidneys were obtained from 7-week-old rats of the three Lyon strains, i.e., at an age preceding the full development of hypertension and major renal lesions in LH rats.19 Because we did not use a pulsatile perfusion system, in baseline conditions all the kidneys were...
perfused at the level of 90 mm Hg, which is approximately the mean arterial pressure in LN rats. In these baseline conditions, LH kidneys, compared with both LN and LL, exhibited significantly increased vascular resistances and decreased GFR and urinary sodium excretion. Such low GFR and urinary sodium excretion are not observed in vivo. This may reflect the fact that, in vivo, LH kidneys are perfused at higher levels than LN or LL kidneys. If one considers that, as indicated by the indirect SBP values recorded in the animals used, the perfusion pressure of LH kidneys should establish 30 mm Hg above that of controls, the present work (see Figure 2) demonstrates that the GFR and urinary sodium excretion of LH kidneys perfused at 120 mm Hg become identical to those of LN and LL kidneys perfused at 90 mm Hg. Therefore, according to Guyton’s hypothesis, these intrinsic renal alterations will allow LH rats to maintain almost normal body fluids in spite of an elevated blood pressure level.

According to Hofbauer et al., the study of perfusion pressure involved stepwise reductions from 170 to 60 mm Hg, each level being maintained for 5 minutes since our preliminary experiments showed that this duration permitted the maximum renin response. Almost no GFR and flow autoregulation could be observed in our conditions. This reflects the large vasodilation of the vessels of isolated kidneys perfused with artificial cell-free solutions. Indeed, the infusion of hydralazine or the use of calcium-free perfusion solutions did not significantly increase the flow through isolated kidneys (Liu, personal communication). When pressure was elevated the fractional sodium reabsorption decreased and the sodium excretion increased. LH kidneys differed from both LN and LL controls by lower perfusion flow, GFR, and natriuresis. The blunted pressure–natriuresis exhibited by LH kidneys appeared to be mainly due to a lower filtered load but also to a slightly higher sodium reabsorption rate.

Great care was taken to measure the renin release in strictly controlled conditions involving the use of rat angiotensinogen (biphenethylomat rat plasma) and the assessment of its excretion by the use of the kinetics of Ang I production for each sample. This latter point is important since renin release exhibited sevenfold changes during the experiments. Using LN and LL kidneys, a classic and reproducible pressure-dependent renin release was observed that reached a plateau at 85 mm Hg. Contrary to dog kidneys, renin release increased as soon as the perfusion pressure decreased, and no threshold pressure level was found. Surprisingly, the renin release of LH kidneys appeared to be pressure-independent. This data is in accordance with those of Tobian et al., who used isolated kidneys of SHR or Dahl salt-sensitive rats. In the present work, we could rule out that the absence of renin release after decreases in perfusion pressure was due to a lack of renin storage in LH kidneys since these latter responded dose-dependently to a β-adrenergic receptor stimulation, a reduced extracellular calcium concentration, or both. Because the sodium reabsorption rate remained largely below that observed in vivo, an alteration in the chloride-sensitive mechanisms of the macula densa is probable in isolated kidney preparations. However, it is likely to occur similarly in the kidneys of the three strains. In addition, because the macula densa stimulates renin release when it senses a decrease in chloride delivery by the proximal tubule, any alteration in this mechanism should have favored the renin release by the LH kidney since its proximal tubular reabsorption of NaCl is slightly higher than that of LN and LL controls. Other probable explanations may involve structural alterations (i.e., stiffness) of the renal vasculature of LH rats, which are suggested by the lower responses of perfusion flow and GFR to increases in pressure, or an enhanced control of renin release by locally produced angiotensin II since single pass–perfused isolated kidneys may be able to synthesize angiotensin II. A more likely hypothesis is that the pressure independence of renin release by LH kidneys could be a consequence of a chronic sodium retention induced in these rats by their increased synthesis of mineralocorticoids. De Rouffignac et al. showed that sodium loading depleted a promptly releasable store of renal renin and Fray demonstrated that the renin release of sodium-loaded kidneys was unresponsive to changes in pressure. Such a lack of pressure influence on renin release in states of sodium retention appears logical since it will contribute to limiting the excess of body fluids. Obviously, further experiments are needed to check this hypothesis and determine the mechanisms involved.

Infusion of AH23848, a specific TXA2/PGH2 receptor antagonist, at a concentration of 4x10⁻⁶ M, which blocks the effects of U46619, did not significantly change the baseline characteristics of the kidneys of the three strains. It also did not markedly alter the responses of LN and LL kidneys to changes in perfusion pressure. On the contrary, it significantly improved the pressure-induced increases in GFR and natriuresis in LH kidneys. This data confirms those of Liu et al., who increased the perfusion pressure of LH kidneys by means of norepinephrine infusions. Interestingly, AH23848 significantly decreased the renin release by the kidneys of the three strains. This result is in accordance with those of Jackson et al., demonstrating that, in dogs, two different TXA2 synthase inhibitors lowered the renin release induced by decreases in RPP. Conversely, Welch et al. reported that TXA2 synthase inhibition elevated the plasma renin activity in anesthetized rats. However, it must be emphasized that in their experiment, the doses of TXA2 synthase inhibitors that enhanced plasma renin were threefold higher than those that blocked TXA2 synthesis. Therefore, our data taken together with those of Jackson et al. favor a stimulating role on renin secretion for TXA2/PGH2.

In conclusion, the present work demonstrated in servocontrolled single pass–perfused isolated kidneys that the renin secretion of young LH rats is not stimulated by decreases in perfusion pressure. Whatever could be the underline mechanisms, this finding suggests that such a pressure independence of renin release could represent an attempt to limit any further sodium retention and blood pressure increase. In addition, it was shown that the activation of intrarenal TXA2/PGH2 receptors enhances the renin release response to changes in perfusion pressure. This latter observation strengthens the potential pathogenetic role of the increased renal synthesis of TXA2 exhibited by kidneys of genetically hypertensive rats of the Lyon strain.
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References

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