Role of Endothelium in Endothelin-Evoked Contractions in the Rat Aorta

Stefano Taddei and Paul M. Vanhoutte

We designed experiments to determine the role of endothelium-derived contracting factor or factors in the response to endothelin-1 and endothelin-3 in the aorta of normotensive and hypertensive rats. Rings of thoracic aortas, with and without endothelium, from normotensive and spontaneously hypertensive rats were suspended in organ chambers for recording of isometric tension in the presence of nitro-L-arginine, an inhibitor of nitric oxide synthase. The removal of endothelium decreased the contractions evoked by both endothelins in the aorta of spontaneously hypertensive but not of normotensive rats. Indomethacin (inhibitor of cyclooxygenase), dazoxiben (inhibitor of thromboxane synthase), and SQ-29,548 (agonist of thromboxane A2 receptors) reduced, in aortic rings of spontaneously hypertensive rats, the contractions to endothelins in rings with but not in those without endothelium, whereas their effect was not endothelium-dependent in tissues of normotensive rats. BQ-123, a selective endothelin-A receptor antagonist, shifted the concentration–response curve to endothelin-1 to the right in a concentration-dependent manner and abolished the endothelium-dependent component of the contractions evoked by the peptide. The presence of the endothelium increased the basal and endothelin-stimulated release of thromboxane B2, the stable metabolite of thromboxane A2, in aortas of spontaneously hypertensive rats but not in those of normotensive rats. These data suggest that endothelium-derived thromboxane A2 contributes to contractions evoked by endothelin-1 and endothelin-3 in the aorta of the spontaneous hypertensive rat but not in that of the normotensive rat. Both the receptors on the endothelial cells (mediating the release of thromboxane A2) and those on vascular smooth muscle belong to the endothelin-A subtype. (Hypertension 1993;21:9–15)

KEY WORDS • endothelins • thromboxane A2 • indomethacin • rats, inbred SHR • rats, inbred WKY • endothelium • aorta

In most isolated arteries, acetylcholine affects the tone of vascular smooth muscle through the release of endothelium-derived relaxing factor.1-3 However, in the aorta of spontaneously hypertensive rat (SHR), the endothelium-dependent relaxations to acetylcholine are blunted by the concomitant release of a cyclooxygenase-dependent endothelium-derived contracting factor, which most likely is an endoperoxide.4,5 In the same preparation, with endothelium from both the SHR and normotensive Wistar-Kyoto (WKY) rat, the direct constrictor effect of endothelin-1 (ET-1) and endothelin-3 (ET-3) on the vascular smooth muscle is blunted by the release of endothelium-derived relaxing factor (most likely nitric oxide).6,7 In addition, in aorta with endothelium from the hypertensive but not from the normotensive strain, contractions to ET-1 are reduced by the inhibitor of cyclooxygenase indomethacin, suggesting that the peptide causes the release of a cyclooxygenase-dependent endothelium-derived contracting factor.8 We designed the present experiments to determine the nature of the cyclooxygenase-dependent endothelin-derived contracting factor released by endothelin in SHR aorta and, because two different endothelin receptor subtypes exist (endothelin-A and endothelin-B), to identify the endothelin receptor subtype involved.

Methods

Contractile Responses

Experiments were performed on the thoracic aortas from male, normotensive WKY rats and age- and weight-matched SHRs (28–32 weeks old; weight: WKY, 394±9.3 g; SHR, 403±10.2 g; n=24 for each group; Harlan Sprague Dawley, Inc., Indianapolis, Ind.). All procedures followed were in accordance with the guidelines of the Animal Protocol Review Committee of Baylor College of Medicine. Systolic blood pressure was determined by an indirect tail-cuff method in the unanesthetized animal before the experiment (WKY, 128±9 mm Hg; SHR, 212±11 mm Hg; p<0.001). The thoracic aorta was excised with rats under sodium pentobarbital anesthesia (50 mg/kg i.p.) and was placed into ice-cold modified Krebs-Ringer bicarbonate solution of the following composition (mM): NaCl 118.3, KCl 4.7, CaCl2 2.5, MgSO4 1.2, KH2PO4 1.2, NaHCO3 25.0, EDTA 0.026, and glucose 11.1 (control solution). Up to eight rings (3–4-mm length) were obtained from

From the Center for Experimental Therapeutics, Baylor College of Medicine, Houston, Tex.

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Address for correspondence: Paul M. Vanhoutte, MD, PhD, Center for Experimental Therapeutics, Baylor College of Medicine, One Baylor Plaza, Room 826E, Houston, TX 77030.

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each animal. In some rings, the endothelium was removed by gently rubbing the intimal surface with the tip of a small forceps. The rings were suspended in organ chambers between a clip and a force transducer (UTC-2, Gould Inc., Cleveland, Ohio) by two stainless-steel stirrups inserted into their lumen. The organ chambers were filled with 25 ml control solution, kept at 37°C, and aerated with a 95% O2–5% CO2 gas mixture. Changes in isometric force were recorded. The preparations were set individually at the optimal point of their length–tension relation as determined by repeated exposure to phenylephrine (10^-6 M) at different levels of stretch. The presence of the endothelium was confirmed by the occurrence of relaxations to acetylcholine (10^-6 M) in rings contracted with phenylephrine (10^-6 M). After that, all experiments were performed in the presence of nitro-L-arginine (10^-4 M) to prevent the production of nitric oxide, which reduces endothelium-dependent contractions. Rings with and without endothelium of the same aorta were studied in parallel.

After 1 hour of equilibration, concentration–response curves (10^-11 to 10^-7 M) to ET-1 or ET-3 were obtained by cumulative addition of the peptides. Responses to ET-1 and ET-3 were obtained both under control conditions and after incubation (45 minutes) with indomethacin (10^-4 M; inhibitor of cyclooxygenase), dazoxiben (10^-4 M; inhibitor of thromboxane synthase), and SQ-29,548 (10^-7 M; antagonist of thromboxane A2/prostaglandin H2 receptors). In another series of experiments, the effect of BQ-123 (3x10^-4, 3x10^-3, 3x10^-6 M; 20 minutes of incubation); selective antagonist of endothelin-A receptors was evaluated.

**Radioimmunoassay of Thromboxane B2**

Rings, with and without endothelium, of aorta were placed in a siliconized glass tube containing 1 ml control solution, oxygenated with 95% O2–5% CO2, and incubated for 60 minutes at 37°C in a shaking water bath. The incubation buffer was then replaced with 1 ml fresh buffer containing nitro-L-arginine (10^-4 M) and, where indicated, dazoxiben (10^-4 M). After 15 minutes, the buffer was replaced once more with fresh solution containing the drugs, and the preparations were allowed to equilibrate for 30 minutes before an aliquot was collected for the determination of the basal generation of thromboxane B2. At that time, the buffer was changed, and ET-1 (or ET-3) was added. A second aliquot was removed after 30 minutes of incubation. Rings were then homogenized for the determination of dry weight. The generation of thromboxane B2 was quantitated by radioimmunoassay using a commercially available kit (Du Pont, Boston). The average intra-assay coefficient of variation was 6.2%, and the interassay coefficient of variation was 5.9%. The sensitivity of thromboxane B2 in this radioimmunoassay was 4 pg per assay tube. The cross-reactivity of the assay was 0.28% for prostaglandin D2, 0.003% for prostaglandin E2, 0.04% for prostaglandin F2α, 0.002% for prostaglandin E1, 0.01% for 6-ketoprostaglandin F1α, 0.001% for 15-dehydroketo prostaglandin E2, 0.002 for prostaglandin A1, and 0.001% for prostaglandin B2.

**Drugs**

Acetylcholine hydrochloride, indomethacin, phenylephrine, and nitro-L-arginine were purchased from Sigma Chemical Co., St. Louis, Mo.; ET-1 and ET-3 were purchased from Peninsula Laboratories, Inc., Belmont, Calif.; SQ-29,548 was generously supplied by Bristol-Myers Squibb, Princeton, Pa.; BQ-123 was supplied by Banyu Pharmaceuticals, Tokyo, and dazoxiben by Pfizer, Groton, Conn. Drugs were prepared daily in distilled water, except for indomethacin, which was dissolved by sonication in an equimolar concentration of Na2CO3 (final bath concentration, 10^-5 M).

**Calculations and Statistical Analysis**

Increases in tension are expressed as percent of the maximal response to phenylephrine (10^-6 M). Results are presented as mean±SEM; n represents the number of rats studied. For experiments with BQ-123, Schild plots were constructed; the dose ratios of concentrations of ET-1 eliciting 50% of the contraction to phenylephrine in the presence and absence of BQ-123 were used. The pA2 values were calculated if the slope of the plot did not differ significantly from unity (determined by linear regression analysis). Subtraction of the data obtained in paired rings with and without endothelium of the same arteries was performed to evaluate the endothelium-dependent component of the response to endothelins.

Statistical evaluation of the data was performed by Student’s t test for paired and unpaired observations, and analysis of variance was used to check the difference between different treatments. Differences were considered to be statistically significant at a value of p<0.05.

**Results**

**Contractile Responses**

Experiments were performed in the presence of nitro-L-arginine to eliminate the production of nitric oxide. In some rings, this agent caused contractions. When the contraction exceeded 1 g, the experiment was discarded to maintain comparable basal tension levels among experimental groups. For the preparations reported, the increases in basal tension induced by nitro-L-arginine averaged 0.24±0.09 g.

In SHR and WKY aortas, with and without endothelium, ET-1 (10^-10 to 10^-7 M) caused concentration-dependent increases in tension that were slow in onset and reached a stable plateau after approximately 15 minutes. At 10^-7 M (the highest concentration tested), ET-3 evoked significantly smaller contractions than ET-1; the threshold concentration of ET-3 was significantly larger than that of ET-1. In SHR aorta, removal of the endothelium significantly reduced the contraction to the highest concentrations tested of both ET-1 and ET-3, by 31.2% and 33.3%, respectively (Figure 1 and Table 1). When the endothelial component of the response to endothelins in SHR aorta was compared, the concentration–response curve to ET-3 was significantly to the right of that to ET-1 (Figure 2). In WKY aorta, the contractions evoked by ET-1 and ET-3 were minimally affected by the removal of endothelium (Figure 1 and Table 1).

In SHR rings with endothelium, indomethacin, dazoxiben, or SQ-29,548 caused a rightward shift of the concentration–response curve and a reduction of the response to ET-3 (Figure 2). This shift was significant at concentrations of approximately 10^-9 M ET-3 (Figure 3).
Figure 1. Line graphs show contractions evoked by endothelin-1 in aorta from spontaneously hypertensive rat (SHR) (left panel) and Wistar-Kyoto (WKY) rat (right panel) with and without endothelium. All experiments were performed in the presence of nitro-L-arginine \(10^{-4}\) M. Results are mean±SEM (n=6 for each experiment) and expressed as percent of maximal contractions to phenylephrine. *Significant difference between rings with and without endothelium (p<0.05).

**Table 1.** Effect of Indomethacin, Dazoxiben, and SQ-29,548 on Contractions Evoked by Endothelin-3 in Rat Aorta With and Without Endothelium

<table>
<thead>
<tr>
<th>Treatment</th>
<th>10^{-9} M</th>
<th>10^{-8} M</th>
<th>10^{-7} M</th>
<th>10^{-8} M</th>
<th>10^{-7} M</th>
</tr>
</thead>
<tbody>
<tr>
<td>SHR aortas</td>
<td></td>
<td></td>
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<td></td>
<td></td>
</tr>
<tr>
<td>Rings with endothelium</td>
<td></td>
<td></td>
<td></td>
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<td></td>
</tr>
<tr>
<td>Control</td>
<td>3.47±1.18</td>
<td>9.77±3.46</td>
<td>20.71±3.42</td>
<td>49.91±5.10</td>
<td>86.97±5.79</td>
</tr>
<tr>
<td>Indomethacin, 10^{-5} M</td>
<td>1.52±0.96</td>
<td>5.55±1.97</td>
<td>14.39±3.49</td>
<td>24.04±5.49</td>
<td>39.49±7.03</td>
</tr>
<tr>
<td>Dazoxiben, 10^{-4} M</td>
<td>1.75±0.80</td>
<td>5.10±1.87</td>
<td>11.00±2.30</td>
<td>26.42±6.59</td>
<td>48.60±7.56</td>
</tr>
<tr>
<td>SQ-29,548, 10^{-7} M</td>
<td>2.19±1.38</td>
<td>5.51±2.60</td>
<td>12.82±3.65</td>
<td>24.76±6.00</td>
<td>45.15±6.76</td>
</tr>
<tr>
<td>Rings without endothelium</td>
<td></td>
<td></td>
<td></td>
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<td></td>
</tr>
<tr>
<td>Control</td>
<td>1.66±1.12</td>
<td>5.78±2.50</td>
<td>14.51±3.27</td>
<td>30.00±3.15</td>
<td>58.01±5.71</td>
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<tr>
<td>Indomethacin, 10^{-5} M</td>
<td>0.00±0.00</td>
<td>2.78±1.60</td>
<td>10.17±3.17</td>
<td>22.88±5.78</td>
<td>41.10±8.54</td>
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<tr>
<td>Dazoxiben, 10^{-4} M</td>
<td>2.03±0.95</td>
<td>4.93±2.57</td>
<td>9.74±2.81</td>
<td>19.55±3.68</td>
<td>45.17±7.89</td>
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<tr>
<td>SQ-29,548, 10^{-7} M</td>
<td>1.61±0.75</td>
<td>5.01±2.35</td>
<td>13.49±3.27</td>
<td>24.59±3.30</td>
<td>49.33±2.81</td>
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<tr>
<td>WKY aortas</td>
<td></td>
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<tr>
<td>Rings with endothelium</td>
<td></td>
<td></td>
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</tr>
<tr>
<td>Control</td>
<td>0.00±0.00</td>
<td>1.12±0.72</td>
<td>30.22±8.13</td>
<td>59.84±8.48</td>
<td>95.96±11.04</td>
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<td>Indomethacin, 10^{-5} M</td>
<td>0.00±0.00</td>
<td>0.95±0.95</td>
<td>8.41±3.13</td>
<td>26.35±7.73</td>
<td>79.85±4.87</td>
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<tr>
<td>Dazoxiben, 10^{-4} M</td>
<td>0.69±0.69</td>
<td>2.63±1.25</td>
<td>10.33±2.29</td>
<td>40.73±6.80</td>
<td>85.45±3.89</td>
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<tr>
<td>SQ-29,548, 10^{-7} M</td>
<td>0.69±0.69</td>
<td>1.97±1.25</td>
<td>6.20±2.28</td>
<td>25.06±8.97</td>
<td>73.63±11.05</td>
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<tr>
<td>Rings without endothelium</td>
<td></td>
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<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Control</td>
<td>0.00±0.00</td>
<td>1.65±1.05</td>
<td>8.09±1.75</td>
<td>44.26±8.63</td>
<td>77.37±7.11</td>
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<td>Indomethacin, 10^{-5} M</td>
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<td>0.00±0.00</td>
<td>7.88±2.82</td>
<td>21.76±6.25</td>
<td>69.48±5.29</td>
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<td>Dazoxiben, 10^{-4} M</td>
<td>0.40±0.40</td>
<td>1.19±1.19</td>
<td>9.06±3.16</td>
<td>41.23±10.57</td>
<td>76.00±7.86</td>
</tr>
<tr>
<td>SQ-29,548, 10^{-7} M</td>
<td>0.30±0.30</td>
<td>1.06±0.68</td>
<td>6.27±2.16</td>
<td>34.24±10.56</td>
<td>60.67±11.87</td>
</tr>
</tbody>
</table>

SHR, spontaneously hypertensive rats; WKY, Wistar-Kyoto rats. All experiments performed in the presence of nitro-L-arginine \(10^{-4}\) M. Results are mean±SEM (n=6) and are expressed as percent of maximal contraction to phenylephrine.

*Significant difference between control and treated rings.
†Significant difference between rings with and without endothelium.
ET-1, -log (M)

FIGURE 2. Line graph shows contractions evoked by endothelin-1 (ET-1) and endothelin-3 (ET-3) in aorta from spontaneously hypertensive rat. All experiments were performed in the presence of nitro-L-arginine (10^{-4} M). Data are presented as subtraction curves (response of ring without endothelium minus response of paired ring with endothelium). Results are mean±SEM (n=6 for each experiment) and expressed as percent of maximal contractions to phenylephrine. *Significant difference between rings with and without endothelium (p<0.05).

ben, or SQ-29,548 produced a comparable inhibition of the contractions evoked by ET-1 and ET-3, which was not significantly different between rings with or without endothelium (Figure 4 and Table 1).

In SHR aorta with endothelium, BQ-123 caused a parallel, concentration-dependent shift to the right of the concentration–response curve to ET-1, as well as a reduction in the response to the highest concentration of the peptide (Figure 5). The endothelin-A antagonist abolished the endothelium-dependent component of the response to ET-1 (Figure 6). In WKY aorta with endothelium, BQ-123 induced a shift to the right in the concentration–response curve to ET-1 without affecting the maximal response (Figure 5); removal of the endothelium did not significantly alter the effect of BQ-123 (data not shown). In all groups, the slope of the Schild plot for the endothelin-A antagonist was not significantly different from unity (Figure 7 and Table 2). BQ-123 did not affect the concentration–response curve to phenylephrine (from 10^{-11} to 10^{-4} M) in rings (with endothelium) from either SHR or WKY aortas (n=2; data not shown).

Release of Thromboxane B_{2}

In SHR aorta but not in WKY aorta, the presence of endothelium significantly increased the basal release of thromboxane B_{2}. At 10^{-7} M, ET-1 and ET-3 augmented the production of thromboxane B_{2}, which was increased further by the presence of endothelium in SHR aorta (Figure 8). In WKY aorta, the presence of endothelium did not affect the increased production of thromboxane B_{2} evoked by the peptides (Figure 8). Dazoxiben abolished both the basal and endothelin-induced release of thromboxane B_{2} (data not shown).

Discussion

In rat aorta, ET-1 and ET-3 can release endothelium-derived relaxing factor, and the removal of the endothelium or treatment with antagonists of nitric oxide synthase augment contractions evoked by endotheлина.6,7 Because this effect is more pronounced in arteries from hypertensive than those from normotensive animals, all the experiments of the present study were performed in the presence of nitro-L-arginine, a noncompetitive inhibitor of nitric oxide synthase.13 Inhibitors of this enzyme also potentiate endothelium-dependent contractions in SHR aorta, most likely by...
preventing a chemical interaction between nitric oxide and endothelium-derived contracting factor.9

In the present study, ET-1 and ET-3 evoked contractions in SHR aorta that were blunted by the removal of the endothelium. In confirmation of earlier findings with ET-1,8 the inhibitor of cyclooxygenase indomethacin inhibited the response to the peptides. This finding indicates that endotheilins induce the production of a cyclooxygenase-dependent endothelium-derived contracting factor or factors in the aorta of hypertensive rats.8 The observation that dazoxiben, an inhibitor of thromboxane synthase, and SQ-29,548, an antagonist of thromboxane A2 receptors, have an inhibitory effect comparable to that of indomethacin suggests that thromboxane A2 rather than endoperoxides5 is the cyclooxygenase-dependent endothelium-derived contracting substance that contributes to the response to endothelins. The greater inhibition of the response to the peptide induced by the treatment with indomethacin, SQ-29,548, and dazoxiben as compared with the removal of the endothelium indicates that a certain amount of thromboxane A2 must be produced in the smooth muscle cells. This interpretation is strengthened by the observation (in confirmation of previous reports14,15) that in the aortas of normotensive animals, the removal of the endothelium did not modify the response to endothelins, and that indomethacin, dazoxiben, and SQ-29,548 blunted the contractions evoked by

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**Figure 4.** Line graphs show effect of indomethacin, SQ 29548, and dazoxiben on contractions evoked by endothelin-1 in aorta from Wistar-Kyoto (WKY) rat with (left panel) and without (right panel) endothelium. All experiments were performed in the presence of nitro-L-arginine (10^{-4} M). Results are mean±SEM (n=6 for each experiment) and expressed as percent of maximal contractions to phenylephrine. *Significant difference between control and treated rings (p<0.05).

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**Figure 5.** Line graphs show effect of treatment with BQ-123 (from 3×10^{-8} to 3×10^{-6} M) on contractions evoked by endothelin-1 in aorta from spontaneously hypertensive rat (SHR) (left panel) and Wistar-Kyoto (WKY) rat (right panel) with endothelium. All experiments were performed in the presence of nitro-L-arginine (10^{-4} M). Results are mean±SEM (n=4 for each experiment) and expressed as percent of maximum to phenylephrine.
the peptides to the same extent in rings with and without endothelium. As reported previously, in rings without endothelium from SHR aorta, the contractions to endothelins were reduced as compared with WKY aorta. Treatment with indomethacin, SQ-29,548, and dazoxiben blunted the response to endothelins in WKY aorta without endothelium, whereas it was less effective in the same preparations from the SHR. This finding indicates that in aorta of normotensive rats endothelins cause the production of vasoconstrictor products of cyclooxygenase in the vascular smooth muscle cells.

Taken in conjunction, the present data indicate that a component of the contraction to endothelins in the rat aorta is mediated by the generation of a cyclooxygenase-dependent substance, presumably thromboxane A2, which in the hypertensive aorta is produced mainly by the endothelium, whereas in the normotensive blood vessel wall, it originates in another structure, most likely the vascular smooth muscle. This interpretation is supported by the finding that BQ-123, a selective antagonist of this subtype of endothelin receptors, abolishes the endothelial component of the response. An identical interpretation can be reached for the subtype mediating the direct effect or effects of endothelins on vascular smooth muscle. Indeed, in both SHR and WKY aorta without endothelium, BQ-123 caused a rightward shift of the concentration–response curves to ET-1 and ET-3 but only results in a decrease in response to the highest concentration of the peptides. The most likely explanation for these observations is that, at lower concentrations, the effect of the peptides on the smooth muscle is responsible for the contraction, whereas the release of endothelium-derived contracting factor only occurs when higher concentrations of endothelins are reached.

The fact that ET-1 is more potent than ET-3 in inducing endothelium-dependent contractions, and presumably the underlying release of thromboxane B2, suggests that the receptor involved at the endothelial cells belongs to the endothelin-A subtype. This interpretation is supported by the finding that BQ-123, a selective antagonist of this subtype of endothelin receptors, abolishes the endothelial component of the response. An identical interpretation can be reached for the subtype mediating the direct effect or effects of endothelins on vascular smooth muscle. Indeed, in both SHR and WKY aorta without endothelium, BQ-123 caused a rightward shift of the concentration–response.

### Table 2. \( pA_2 \) Values and Slopes for BQ-123 in Rat Aorta With and Without Endothelium

<table>
<thead>
<tr>
<th>Parameters</th>
<th>SHR</th>
<th>WKY</th>
</tr>
</thead>
<tbody>
<tr>
<td>With endothelium</td>
<td>Without endothelium</td>
<td>With endothelium</td>
</tr>
<tr>
<td>( pA_2 )</td>
<td>8.19±0.09</td>
<td>7.83±0.08</td>
</tr>
<tr>
<td>Slope</td>
<td>0.98</td>
<td>0.98</td>
</tr>
</tbody>
</table>

SHR, spontaneously hypertensive rats; WKY, Wistar-Kyoto rats.
curve to ET-1, whereby the slopes of the Schild plots were not different from unity, suggesting the competitive nature of the antagonism in both strains. The pA2 values were not significantly modified by removal of endothelium in both strains; however, there was a 10-fold difference between the values obtained in the SHR and WKY rat. This suggests that there may be different receptor subtypes involved in the response to ET-1 in the SHR and WKY rat.

In conclusion, the present observations indicate that in SHR aorta, a component of the contractions evoked by ET-1 and ET-3 is mediated by the thromboxane A2 that is produced by endothelial cells.

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References

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