Effects of Treppe and Calcium on Intracellular Calcium and Function in the Failing Heart From the Spontaneously Hypertensive Rat

Wesley W. Brooks, Oscar H.L. Bing, Sheldon E. Litwin, Chester H. Conrad, James P. Morgan

Abstract We studied functional and intracellular calcium responses to treppe and extracellular calcium in spontaneously hypertensive rat (SHR) hearts during the transition from compensated pressure overload to failure. Intracellular calcium was measured using aequorin, a bioluminescent Ca\(^{2+}\) indicator. Experiments were performed with intact, isovolumically contracting, buffer-perfused hearts from three rat groups: (1) aging SHR with evidence of heart failure (SHR-F), (2) age-matched SHR with no evidence of heart failure (SHR-NF), and (3) age-matched normotensive Wistar-Kyoto (WKY) rats. In each experiment, left ventricular pressure and intracellular calcium transients were simultaneously recorded. Hearts were studied at 30°C and paced at a rate of 1.6 Hz while being perfused with oxygenated Krebs-Henseleit solution (95% O\(_2\)/5% CO\(_2\)) at 100 mm Hg. At the baseline state, peak systolic pressure was greatest in the SHR-NF group and lowest in the SHR-F group. Peak and resting [Ca\(^{2+}\)], were not significantly different among groups; however, the calcium transient was prolonged in the SHR-NF and SHR-F groups. With increasing perfusate [Ca\(^{2+}\)], from 0.5 to 3.0 mmol/L, the relative increases in peak [Ca\(^{2+}\)], and peak systolic pressure were similar among groups. When stimulation rate was increased from 1.6 to 2.0, 2.4, 2.8, and 3.2 Hz, peak [Ca\(^{2+}\)], peak systolic pressure, and ±dP/dt fell in SHR-F hearts. Peak systolic pressure decreased significantly in the WKY group at rates above 2.4 Hz but did not decline in the WKY group. Peak [Ca\(^{2+}\)], increased in the WKY and SHR-NF groups with increasing heart rates. Peak systolic pressure did not fall significantly in the WKY group at any heart rate. Elevation of diastolic [Ca\(^{2+}\)], and/or calcium transient and pressure alternans were present in 8 of 13 SHR-F hearts at the highest stimulation rate, findings that were absent in both the WKY and SHR-NF hearts. We conclude the following: (1) Under baseline conditions, depressed contractile function of failing myocardium cannot be explained by decreased peak [Ca\(^{2+}\)]; (2) relative increases in [Ca\(^{2+}\)], and inotropy with increasing [Ca\(^{2+}\)], are proportional among groups; and (3) although peak systolic [Ca\(^{2+}\)], and inotropy are maintained with increasing stimulation rate in the WKY and SHR-NF groups, peak systolic [Ca\(^{2+}\)], and pressure decrease in parallel in the SHR-F heart with increasing stimulation rate, suggesting that impaired calcium cycling may contribute to compromised pump function in the SHR-F heart. (Hypertension. 1994;24:347-356.)

Key Words • heart hypertrophy • heart failure, congestive • calcium • muscle, cardiac

Relatively few models of cardiac pressure overload have stable hypertrophy persisting over most of the animal's life followed by a state in which chamber and muscle function are impaired. The spontaneously hypertensive rat (SHR) is a model that in many respects parallels the course of the hypertensive heart disease. Studies of both failing human\(^1,3\) and hypertrophied and failing SHR\(^4,5\) myocardia reveal altered calcium transients and impaired force development; however, the possible role of altered intracellular calcium handling in the development of depressed contractile function during the transition from compensated hypertrophy to failure remains unclear. Recent isolated muscle studies have demonstrated differences in the force-frequency response between failing and nonfailing human myocardia and a reduced ability of the failing heart to increase force at increasing stimulation rates.\(^6\) Studies of isolated ventricular strips from failing human ventricles demonstrate that increased stimulation rates enhance disturbances in calcium handling\(^1,2\) and force development.\(^6\) It is well recognized that treppe and increasing [Ca\(^{2+}\)], lead to an increase in [Ca\(^{2+}\)], and the number of occupied crossbridges per unit time, which in turn results in an increase in myocardial energy demands. In addition, both of these interventions lead to increases in [Ca\(^{2+}\)], and a greater load on intracellular calcium control systems. However, one problem with isolated muscle studies is the uncertainty regarding diffusion, inherent in isolated muscle studies, and the applicability to in vivo physiology. To bridge the gap between isolated muscle studies and in vivo physiology, we have carried out studies in the isolated perfused heart preparation. Studies using a variety of calcium indicators have been applied to studies of perfused heart preparations.\(^7,10\) One of us (J.P.M.) has developed a system in which the bioluminescent calcium indicator aequorin can be used to measure [Ca\(^{2+}\)], in isolated perfused hearts.\(^11,13\) This system permits a study of beat-to-beat changes in [Ca\(^{2+}\)], and ventricular global function in coronary perfused...
preparations, thus avoiding potentially confounding variables such as core hypoxia that may be present in isolated papillary muscle preparations at higher stimulation frequencies. We considered that by comparing two interventions which increase both energy demands and the demands on intracellular calcium control systems, we may be able to obtain additional insights into possible mechanisms of cardiac decompensation in the SHR. Therefore, in the present study we investigated the effects of perfusate calcium concentration and stimulation rate on intracellular calcium handling and left ventricular function in the SHR model of chronic left ventricular pressure overload and failure.

Methods

Animal Model

Male SHR and normotensive Wistar-Kyoto (WKY) control rats, 6 to 9 months of age, were purchased as retired breeders from Taconic Farms, Germantown, NY, and boarder in the animal facility at the Boston VA Medical Center until the time of study (18 to 24 months of age). We have previously found that a number of SHR demonstrate evidence of heart failure beginning at the age of 18 months. Therefore, at the age of 18 months the rats were observed on a daily basis. When SHR were observed to develop tachypnea and labored respiration, they were studied within 1 to 2 weeks. No WKY rats developed respiratory difficulties. A few SHR with respiratory findings died before they could be studied and at autopsy had pathological findings consistent with heart failure, as described below. Isolated intact perfused heart preparations were examined from three rat groups: (1) aging SHR with physical and pathological findings consistent with heart failure, as described below; (2) age-matched normotensive WKY rats without evidence of heart failure (WKY-NF), and (3) age-matched normotensive SHR without evidence of heart failure (SHR-NF).

Preparation of Isolated Perfused Heart

The isolated isovolumically contracting rat heart preparation has been previously described. Briefly, at the time of study the rats were decapitated. Their hearts were quickly removed and placed in oxygenated bicarbonate-buffered physiological salt solution containing heparin in 23° to 25°C. The ascending aorta was cannulated, the pulmonary artery was ligated, and hearts were perfused with oxygenated Krebs-Henseleit solution at 30°C with a constant flow set to achieve coronary flow rate and temperature pressures16 were obtained in all rats on the day before the study. Cardiac catheterization was performed on a number of animals from each group on the day of study as previously described. Briefly, animals were anesthetized with ether, and the right carotid artery was isolated. A catheter-tipped micromanometer (model SPR-249, Millar Instruments) was advanced into the left ventricle where left ventricular pressures were recorded at high and low gain in order to obtain baseline hemodynamic data.

Hemodynamic Parameters

Baseline measurements were obtained when the preparation achieved a steady state after instrumentation (approximately 15 minutes). After this equilibration period, a baseline pressure-volume relation was determined; left ventricular peak systolic and end-diastolic pressures were measured in the oxygenated control state 20 to 30 seconds after 0.02-ml increments in balloon volume. Volumes were increased until an end-diastolic pressure of approximately 20 mm Hg was achieved. The balloon volume was then adjusted to an end-diastolic pressure of 2 to 4 mm Hg and kept isovolumic throughout the experiment.

Loading Procedure for Aequorin

After equilibration and baseline measurements, a localized region of the left ventricular apex was microinjected with aequorin as previously described. In brief, hearts were cooled to 24°C and perfused with a medium containing 0.2 mmol/L calcium. Within 2 minutes systolic pressure fell from 100 to less than 20 mm Hg. After 5 minutes of 0.2 mmol/L calcium perfusion, coronary perfusion was reduced to achieve a perfusion pressure of 10 to 20 mm Hg in preparation for aequorin loading. Subsequently, the heart was temporarily raised out of the organ bath, and aequorin solution (3 to 5 /lL) was injected with a low-resistance glass micropipette into the interstitium of the epicardium of the apex of the left ventricle just beneath the epicardium. The heart was then repositioned in the bottom of the organ bath. Hearts were enclosed in a light-tight box designed for aequorin studies by Blinks and modified for the Langendorff-perfused heart as previously described by Kihara et al. In brief, this system consists of an ellipsoid mirror with the aequorin-loaded region of the heart positioned at one focal point and the photocathode of a photomultiplier tube (9635QA, Thorn-EMI, Gencom, Inc) at the other. This arrangement maximized the collection of light emitted from the aequorin-loaded site.

Protocol

Baseline isovolumic contraction parameters were recorded simultaneously with the aequorin light signal from all heart preparations after a 10- to 15-minute equilibration period under baseline conditions. In preparation for determination of the concentration-response relation to calcium, the perfusate was replaced with a zero phosphate solution to avoid precipitation of calcium salts. Increasing concentrations of calcium were then added to the perfusate to obtain a dose-response
relation. Concentration-response relations to \( \left[Ca^{2+}\right] \), (0.5 to 3.0 mmol/L) were studied at a constant heart rate of 1.6 Hz in all hearts. Baseline measurements were obtained at 1.0 mmol/L calcium and then reduced to 0.5 mmol/L before the calcium concentration was raised to 0.75, 1.0, 2.0, and 3.0 mmol/L calcium. Calcium concentrations higher than 3 mmol/L frequently caused electrical irritability, particularly in the SHR-F group, and therefore were not used. The responses to increasing stimulation rates were examined by increasing the paced frequency in 0.4-Hz increments from a baseline state of 1.6 Hz up to a maximum of 4.0 Hz. At each rate examined, the hearts were paced at the test frequency for 1 minute and then allowed to return to the baseline state (1.6 Hz) before the hearts were paced at a higher rate. In these experiments the perfusate calcium concentration was maintained constant at 1.0 mmol/L. Calcium was measured by the method of fractional luminescence. At the conclusion of the experiment, hearts were perfused with 8 mmol/L calcium and 5% Triton X-100 to lyse the cells and expose all the remaining aequorin to calcium. This results in a nearly instantaneous burst of light that declines over a period of minutes to seconds. The area under the resulting curve was then integrated to obtain a value for \( L_{\text{max}} \), the total amount of light emitted from the aequorin in the heart. Intracellular calcium was derived from the ratio of instantaneous light to \( L_{\text{max}} \) using a calibration curve. Intracellular magnesium was assumed to be 0.85 mmol/L, the value reported in isolated rat heart under baseline (zero) conditions. The area under the resulting curve was then compared using one-way ANOVA with replication. A one-way ANOVA was used to examine group effects. The Newman-Keuls multiple-sample comparison test was used to localize differences when appropriate. Data are expressed as mean±SD.

### Results

#### Cardiac Catheterization

Studies were conducted with 24 SHR and 16 WKY rats, 18 to 24 months of age. Fourteen SHR had findings of heart failure (SHR-F group), including tachypnea (n=13), pleural and/or pericardial effusions (n=14), left atrial thrombi (n=14), and right ventricular hypertrophy (as defined by right ventricular weight–body weight ratio >0.8 mg/g, n=14).

Cardiac catheterization was performed on 13 animals in the SHR-F group, 7 SHR-NF, and 10 WKY rats (Table 1). Tail-cuff blood pressures obtained in the conscious animal were elevated in both SHR groups (SHR-NF, 177±22 mm Hg; SHR-F, 150±13 mm Hg) relative to the WKY group (116±4 mm Hg, \( P<.01 \)) but were lower in the SHR-F group compared with the SHR-NF group (\( P<.05 \), Table 2). In all groups, peak systolic measurements were lower in the anesthetized animals compared with tail-cuff measurements in the conscious animals. Peak systolic pressure was greater in the SHR-NF group compared with both the SHR-F and WKY groups. Diastolic pressure was elevated in the SHR-F group relative to the WKY and SHR-NF groups. Mean aortic (perfusion) pressure for the WKY group was 86±16 mm Hg; SHR-F, 86±20 mm Hg; and SHR-NF, 118±17 mm Hg.

#### Clinicopathological Data

Fibrosis was noted by gross inspection in the hearts from both the SHR-F and SHR-NF groups; fibrosis was not evident in any of the WKY hearts. Table 2 presents data on body weight, cardiac chamber weight, and chamber weight normalized for body weight. Both SHR groups demonstrated left ventricular hypertrophy, as indicated by an increase in the ratio of left ventricular weight to body weight; this ratio was greater in the SHR-F relative to the SHR-NF group. Right ventricular hypertrophy was one of the criteria for selection of the SHR-F group; the ratio of right ventricular weight to body weight increased in the SHR-F group compared with both the SHR-NF and WKY groups.

### Table 1. In Vivo Heart Rate and Hemodynamic Data

<table>
<thead>
<tr>
<th>Group</th>
<th>HR, bpm</th>
<th>SP, mm Hg</th>
<th>DP, mm Hg</th>
<th>+dP/dt, mm Hg/s</th>
<th>-dP/dt, mm Hg/s</th>
</tr>
</thead>
<tbody>
<tr>
<td>WKY (n=10)</td>
<td>404±48*</td>
<td>105±11†</td>
<td>3±1*</td>
<td>544±67†</td>
<td>3760±883</td>
</tr>
<tr>
<td>SHR-F (n=13)</td>
<td>305±35f</td>
<td>101±11†</td>
<td>10±5§</td>
<td>2907±347§</td>
<td>2582±374§</td>
</tr>
<tr>
<td>SHR-NF (n=7)</td>
<td>407±43</td>
<td>138±17</td>
<td>6±3</td>
<td>6216±167</td>
<td>4589±933</td>
</tr>
</tbody>
</table>

HR indicates heart rate; SP, systolic pressure; DP, diastolic pressure; +dP/dt, maximal rate of rise of systolic pressure; -dP/dt, maximal rate of fall of systolic pressure; WKY, normotensive Wistar-Kyoto rats; SHR-NF, spontaneously hypertensive rats without heart failure; and SHR-F, spontaneously hypertensive rats with heart failure. Values are mean±SD.

*\( P<.01 \), t\( P<.05 \) vs SHR-F.

### Table 2. Chamber Weight, Body Weight, and Blood Pressure Data

<table>
<thead>
<tr>
<th>Group</th>
<th>LV, g</th>
<th>RV, g</th>
<th>BW, g</th>
<th>LV/BW, mg/g</th>
<th>RV/BW, mg/g</th>
<th>BP, mm Hg</th>
</tr>
</thead>
<tbody>
<tr>
<td>WKY (n=16)</td>
<td>1.4±0.2</td>
<td>0.3±0.1</td>
<td>560±92‡</td>
<td>2.6±0.4†‡</td>
<td>0.6±0.3*</td>
<td>116±4‡</td>
</tr>
<tr>
<td>SHR-F (n=16)</td>
<td>1.9±0.2</td>
<td>0.5±0.1</td>
<td>398±32</td>
<td>4.9±0.6†</td>
<td>1.2±0.1‡</td>
<td>150±13</td>
</tr>
<tr>
<td>SHR-NF (n=12)</td>
<td>1.5±0.3</td>
<td>0.2±0.03</td>
<td>415±56</td>
<td>3.6±0.8</td>
<td>0.6±0.1</td>
<td>177±22</td>
</tr>
</tbody>
</table>

LV indicates left ventricular wet weight; RV, right ventricular wet weight; BW, body weight; BP, systolic arterial pressure determined via indirect tail-cuff method; WKY, normotensive Wistar-Kyoto rats; SHR-NF, spontaneously hypertensive rats without heart failure; and SHR-F, spontaneously hypertensive rats with heart failure. Values are mean±SD.

*\( P<.01 \), t\( P<.05 \) vs SHR-F.

\( P<.05 \) vs SHR-NF.
Isovolumic left ventricular pressure (LVP) (bottom) in normotensive, hypertrophied, and failing hearts. The percent of maximal developed pressure was reduced (P<.01) in the SHR-F group relative to both the SHR-NF and WKY groups.

Table 4 presents light transient and mechanical temporal data. The duration of the light transient (TPL, TPL, and TPL+L90) was increased in the SHR-NF group relative to the WKY group. Time to peak pressure (TPP) was increased in both SHR groups relative to the WKY group. The time needed for pressure to fall from peak to 50% of peak (P50) was reduced in the SHR-F group compared with the WKY group.

Resting and Peak Intracellular Calcium Concentrations

Resting and peak intracellular calcium concentrations were calculated from signals in WKY, SHR-NF, and SHR-F hearts using the method of fractional luminescence. Table 5 summarizes the fractional luminescence values for each group. Data on fractional luminescence and estimated intracellular calcium with 1.0 mmol/L perfusate calcium were obtained under baseline conditions (stimulation rate, 1.6 Hz at 30°C). Diastolic intracellular calcium and the peak of the Ca2+ transient were not significantly different among groups.

Changing Perfusate Calcium Concentration

Fig 3 demonstrates the effect of increasing perfusate calcium concentration on left ventricular function and the aequorin light signal. When perfusate calcium was increased from 0.5 to 3.0 mmol/L, peak systolic pressure increased in all groups (WKY, from 55±17 to 101±27 mm Hg; SHR-NF, from 75±25 to 133±30 mm Hg; SHR-F, from 41±10 to 77±21 mm Hg; all P<.001). Significant increases in systolic pressure and the aequorin light signal were seen in all groups in response to increasing calcium (Figs 3 and 4). The light signal response to increasing calcium in the SHR-NF and SHR-F groups appeared greater than in the WKY group. To test the calcium responsiveness of the normotensive, hypertrophied, and failing hearts, we determined the calcium concentration-response relation. However, the percent of maximal developed pressure and light at increasing calcium concentrations were comparable among the groups at each calcium concentration studied (Fig 5). The duration of both the pressure and light signals (TPP and TPL) were largely unchanged at the various calcium concentrations. The relaxation parameters of the pressure and light signals tended to decrease slightly with increasing perfusate calcium. There were no significant differences in the response of these parameters among the groups at each calcium concentration examined.

Changing Stimulation Rate

Fig 6 demonstrates the effect of increasing stimulation rate on the aequorin light signal and left ventricular function. The left panel demonstrates the usual pattern observed in the unpaced (intrinsic heart rate) preparation under baseline conditions in the WKY, SHR-NF, and SHR-F hearts. Table 6 compares the changes in baseline aequorin light signal and left ventricular functional parameters for the three groups as the stimulation rate increased from 1.6 Hz. As noted above, the intrinsic heart rate of the SHR-F hearts was significantly slower than both SHR-NF and WKY hearts (P<.01). At a constant paced rate of 1.6 Hz, peak systolic pressure
was 94±23, 92±30, and 55±21 mm Hg in the WKY, SHR-NF, and SHR-F groups, respectively. Increasing the frequency of stimulation from a paced rate of 1.6 to 2.0 Hz resulted in a decrease in peak light and peak systolic pressure in the SHR-F hearts, whereas these parameters increased in both SHR-NF and WKY hearts. When the frequency was increased to 2.4 Hz, peak light further decreased in the SHR-F hearts and increased in the WKY and SHR-NF hearts. Peak systolic pressure was decreased in the SHR-NF hearts, markedly decreased in the SHR-F hearts, and only slightly decreased from control levels in the WKY hearts at this increased stimulation rate of 3.2 Hz. Eight of the 13 SHR-F hearts examined exhibited either light and pressure alternans or a marked increase in resting diastolic calcium and diastolic pressure values at the higher stimulation frequency. These phenomena were not in evidence in hearts from WKY or SHR-NF animals. The bottom right panel of Fig 6 demonstrates the presence of well-defined electrical alternans in the SHR-F heart at a simulation rate of 3.2 Hz. Fig 7 is another example of the effect of increasing stimulation rate from 1.6 to 3.2 Hz in WKY and SHR-F hearts. Note the presence of distinct electrical and mechanical alternans in the SHR-F hearts and a marked increase in diastolic pressure. At the higher stimulation rate (3.2 Hz), the amplitude of the calcium transient was decreased, whereas the calcium signal appears to be prolonged in the SHR-F heart. In contrast, in the WKY heart the amplitude of the calcium signal is increased and its duration decreased.

The percent of maximal rate of developed pressure (+dP/dt) and relaxation (−dP/dt) were compared at increasing stimulation rates (Fig 8). The values for +dP/dt at 1.6 Hz were 1520±400, 1109±250, and 732±353 mm Hg/s for the WKY, SHR-NF, and SHR-F groups, respectively. At relatively low stimulation rates the SHR-F curve begins to diverge from both the SHR-NF and WKY groups. At the frequency of 2.0 Hz, peak light and +dP/dt markedly decreased in the SHR-F hearts and increased in the WKY and SHR-NF hearts (P<.01). The value for +dP/dt was first depressed relative to control values in the SHR-NF hearts at 2.4 Hz and decreased to a greater extent at a higher stimulation frequency. In comparison, in WKY hearts +dP/dt remained at control levels up to a frequency of 3.2 Hz. Peak light was increased above control values in both the WKY and SHR-NF groups at all stimulation rates examined.

The frequency dependence of the peak rates of pressure fall (−dP/dt) was generally similar to that pattern observed for +dP/dt. The values for −dP/dt at 1.6 Hz were 622±173, 660±221, and 416±177 mm Hg/s for the WKY, SHR-NF, and SHR-F groups, respectively. In the WKY hearts −dP/dt increased as the frequency of stimulation was increased up to 3.2 Hz. In the SHR-NF hearts −dP/dt increased at 2.0 Hz and began to decrease at 2.4 Hz. In the SHR-F hearts −dP/dt decreased as the stimulation rate was increased from 1.6 to 2.0, 2.8, and 3.2 Hz.

Increasing the stimulation rate resulted in an abbreviation of the pressure response and to a lesser extent the duration of the intracellular calcium transient in all groups. Time to peak pressure at 1.6 Hz was 141±15, 149±24, and 152±14 milliseconds for the WKY, SHR-F, and SHR-NF groups, respectively. At 3.2 Hz these values decreased to 125±9, 120±8, and 122±15 milliseconds, respectively. In general, the duration of the light signal (TPL+L*) decreased (range, 4% to 9% among groups) in all groups when the stimulation rate was increased from 1.6 to 3.2 Hz.

### Table 3. Comparison of Hemodynamic Parameters at Steady State

<table>
<thead>
<tr>
<th>Group</th>
<th>LV Systolic Pressure, mm Hg</th>
<th>Diastolic Pressure, mm Hg</th>
<th>LV Wall Thickness, mm</th>
<th>LV Chamber Radius, mm</th>
<th>Systolic Stress, kdyne/cm²</th>
</tr>
</thead>
<tbody>
<tr>
<td>WKY (n=10)</td>
<td>93±22*</td>
<td>3±2</td>
<td>0.46±0.04</td>
<td>0.23±0.01</td>
<td>16.0±5.6</td>
</tr>
<tr>
<td>SHR-F (n=13)</td>
<td>65±12†</td>
<td>3±2</td>
<td>0.51±0.04</td>
<td>0.26±0.04</td>
<td>11.2±3.9†</td>
</tr>
<tr>
<td>SHR-NF (n=10)</td>
<td>120±22†</td>
<td>4±3</td>
<td>0.48±0.04</td>
<td>0.24±0.04</td>
<td>20.2±6.6</td>
</tr>
</tbody>
</table>

*P<.05 vs SHR-F.
†P<.05 vs WKY.

### Table 4. Temporal Characteristics of Calcium Transients and Developed Pressure Under Baseline Conditions

<table>
<thead>
<tr>
<th>Group</th>
<th>TPL</th>
<th>L*</th>
<th>Total</th>
<th>TPP</th>
<th>P100</th>
</tr>
</thead>
<tbody>
<tr>
<td>WKY (n=11)</td>
<td>73±11</td>
<td>84±18</td>
<td>176±19</td>
<td>137±16*</td>
<td>138±14*</td>
</tr>
<tr>
<td>SHR-F (n=13)</td>
<td>78±16</td>
<td>95±20</td>
<td>198±25</td>
<td>156±20</td>
<td>115±15</td>
</tr>
<tr>
<td>SHR-NF (n=11)</td>
<td>77±10</td>
<td>103±19†</td>
<td>211±20†</td>
<td>166±18†</td>
<td>137±25</td>
</tr>
</tbody>
</table>

TPL Indicates time to peak light; L*, time to 90% decline from peak light; Total, total duration of light transient; TPP, time to peak systolic pressure; P100, time for pressure to fall from peak to 50% of peak; WKY, normotensive Wistar-Kyoto rats; SHR-NF, spontaneously hypertensive rats without heart failure; and SHR-F, spontaneously hypertensive rats with heart failure. Values are mean±SD expressed as milliseconds.

*P<.05 vs SHR-F.
†P<.05 vs WKY.

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TABLE 5. Resting and Peak [Ca$^{2+}$]

<table>
<thead>
<tr>
<th>Group</th>
<th>Resting Calcium</th>
<th>Peak Calcium</th>
</tr>
</thead>
<tbody>
<tr>
<td>WKY (n=9)</td>
<td>0.33±0.20</td>
<td>0.67±0.21</td>
</tr>
<tr>
<td>SHR-F (n=12)</td>
<td>0.24±0.13</td>
<td>0.93±0.17</td>
</tr>
<tr>
<td>SHR-NF (n=10)</td>
<td>0.26±0.17</td>
<td>0.89±0.20</td>
</tr>
</tbody>
</table>

WKY indicates normotensive Wistar-Kyoto rats; SHR-NF, spontaneously hypertensive rats without heart failure; and SHR-F, spontaneously hypertensive rats with heart failure. Values are mean±SD and expressed as micromoles per liter.

Discussion

The present study examines the relation between left ventricular pump function and intracellular calcium transients in isolated perfused hearts from SHR with compensated hypertrophy and failure relative to a non-hypertensive age-matched group of WKY hearts. Left ventricular developed pressure in the SHR-F group was significantly less than in both the SHR-NF and WKY groups. Abnormal calcium handling has been implicated as a possible cause for myocardial dysfunction in a number of settings.\textsuperscript{1,3,13,24,25} In the baseline state, significant differences in peak or resting [Ca$^{2+}$] were not present among the three animal groups studied. However, the calcium signals of the SHR-NF and SHR-F were prolonged, consistent with previous findings from isolated muscle studies of hypertrophy and failure in which depressed contractile function\textsuperscript{1,2,4,5} has been associated with prolonged calcium transients.\textsuperscript{1,2,4,24} Since peak [Ca$^{2+}$] was not significantly different among the groups, decreased peak [Ca$^{2+}$] cannot account for the depressed left ventricular function seen in the decompensated SHR hearts. However, findings of the present study in perfused hearts show that increasing stimulation rate markedly reduced both peak light and peak systolic pressure in the SHR-F group. In the normotensive WKY group, peak light remained above control values at all stimulation rates studied, and peak systolic pressure was maintained. The response of the SHR-NF hearts was intermediate between the WKY and the SHR-F hearts. In response to increasing rates of stimulation, the maximal rate of pressure fall (−dP/dt) was most depressed in the SHR-F and to a lesser extent the SHR-NF heart relative to WKY hearts. It appears that as the heart rate is increased, the left ventricle of SHR-F hearts is unable to maintain calcium homeostasis, further exacerbating contractile dysfunction. Reduced sarcoplasmic reticulum (SR) calcium accumulation and subsequent release is thought to lead to impaired mechanical relaxation and reduced pressure development in subsequent contractions, a problem that may be enhanced at increased stimulation rates. This formulation could explain impaired SR calcium uptake and depressed function in the failing heart at rapid stimulation rates while adequate calcium handling appears to be present in the baseline state.

As noted above, the finding of similar peak [Ca$^{2+}$] among normotensive, hypertrophied, and failing groups

Fig 3. Tracings show effect of increasing perfusate calcium on aequorin light signal and left ventricular pressure (LVP) in Wistar-Kyoto (WKY) heart (top) and hypertrophied (SHR-NF) (middle) and failing (SHR-F) (bottom) spontaneously hypertensive rat heart. Note the parallel increase in light and pressure as calcium is increased in normotensive, hypertrophied, and failing hearts.

Fig 4. Bar graphs show concentration response of developed pressure and peak light from Wistar-Kyoto (open bars) hearts and hypertrophied (shaded bars) and failing (black bars) spontaneously hypertensive rat hearts to increasing calcium concentrations (0.5 to 3.0 mmol/L). Changes in developed pressure (left) and peak light amplitude (right) are expressed as percent change from 1.0 mmol/L calcium. Data are mean±SD.
Fig 5. Line graphs show concentration response to changes in extracellular calcium in aequorin-loaded whole hearts from normotensive Wistar-Kyoto (○) rats and spontaneously hypertensive rats without (●) and with (★) evidence of failure. Left, isovolumic pressure response expressed as percent of pressure at 3.0 mmo/L calcium concentration; right, peak light response represented as percent of light at 3.0 mmo/L calcium concentration. Data are mean±SD.

(Table 5) indicates that the availability of intracellular calcium for activation of the myofilaments is not diminished. Therefore, decreased peak [Ca²⁺], cannot account for the depressed developed pressure of the SHR-F group in the baseline state. Peak intracellular calcium concentrations from isolated cell, papillary muscle, and perfused heart preparations have been reported to approximate 1.0 mol/L [Ca²⁺] in normotensive animals under similar baseline experimental conditions. Similar to the present study, values reported from isolated rat papillary muscles of SHR-NF and SHR-F hearts and experimentally infarcted WKY hearts also indicate no significant differences in peak [Ca²⁺]. Resting [Ca²⁺], was also similar for the three groups and fell within the range of 0.1 to 0.3 μmol/L [Ca²⁺], reported for rat ventricular myocytes using aequorin and fura as well as in rat papillary muscles using aequorin. Findings are also comparable to resting values reported for normal and hypertrophied ferret hearts.

The possibility that the responsiveness of the myofilaments to calcium is altered by hypertrophy and failure must also be considered. Although baseline left ventricular pressure was significantly reduced in the SHR-F hearts, the relative responses of light and pressure to changes in [Ca²⁺], were similar in normotensive, hypertrophied, and failing hearts. The similarity of these relations among groups suggests a similar responsiveness to calcium. In a study of saponin-skinned myocardial fibers from a similar group of animals, myofilament calcium sensitivity and cooperativity were not different among groups; however, maximal calcium-activated force was increased in the SHR-NF group. Maximal calcium-activated force was found to fall to control (WKY) levels with the development of failure. This finding in skinned myocardial fibers supports the finding of the present study that tension development is depressed with heart failure in the isolated heart. The decrease in maximal force development by the failing heart may be caused by some other factor not directly examined, such as the presence of connective tissue. Overall, depressed contractile function does not appear to be explained by a decrease in [Ca²⁺], or a decrease in the responsiveness of the myofilaments to calcium.

It may be considered that the increased demands associated with an increase in stimulation rate may have shifted energy supply/demand relations in favor of hypoxia or ischemia in the failing heart. However, the increased demands associated with increasing bath (and intracellular) calcium did not result in a reduced response to [Ca²⁺], and pressure development in the
TABLE 6. Effect of Increasing Stimulation Rate on Light and Pressure Parameters

<table>
<thead>
<tr>
<th>Stimulation Rate, Hz</th>
<th>PL (mmHg)</th>
<th>PSP (mmHg)</th>
<th>PL (mmHg)</th>
<th>PSP (mmHg)</th>
<th>PL (mmHg)</th>
<th>PSP (mmHg)</th>
<th>PL (mmHg)</th>
<th>PSP (mmHg)</th>
</tr>
</thead>
<tbody>
<tr>
<td>2.0</td>
<td>108±20</td>
<td>102±15</td>
<td>113±21</td>
<td>90±8*</td>
<td>104±21†</td>
<td>95±8†</td>
<td>118±31†</td>
<td>89±16†‡</td>
</tr>
<tr>
<td>2.4</td>
<td>95±11</td>
<td>86±10</td>
<td>87±13†</td>
<td>71±14</td>
<td>77±19</td>
<td>64±14</td>
<td>66±13§</td>
<td>55±15</td>
</tr>
<tr>
<td>2.8</td>
<td>149±46</td>
<td>104±21</td>
<td>114±21</td>
<td>83±7</td>
<td>107±17</td>
<td>71±7</td>
<td>110±19</td>
<td>62±8</td>
</tr>
<tr>
<td>3.2</td>
<td>104±21†</td>
<td>77±19§</td>
<td>113±21</td>
<td>87±13*</td>
<td>114±21</td>
<td>114±21</td>
<td>110±19</td>
<td>62±8</td>
</tr>
</tbody>
</table>

PL indicates peak light; PSP, peak systolic pressure; WKY, normotensive Wistar-Kyoto rats; SHR-NF, spontaneously hypertensive rats without heart failure; and SHR-F, spontaneously hypertensive rats with heart failure. Values are mean±SD and expressed as percent control at 1.6 Hz.

*P<.05, †P<.01 vs SHR-F.
‡P<.05, §P<.01 vs SHR-NF.

SHR-F group. The changes in the calcium signal in the SHR-F group with increasing heart rate are not consistent with those changes observed with either brief periods of ischemia or hypoxia. In this case, the calcium transient is relatively preserved while force falls precipitously. Smith and Allen reported no change in [Ca^{2+}], with simulated ischemia while force declined. Findings of proportional decreases in [Ca^{2+}] and pressure may be more consistent with primary impairment of SR calcium cycling with increasing heart rate as opposed to energy supply/demand relations. Additional evidence of impaired SR function found in the present study is the appearance of alternans of both calcium transients and mechanical function found only in the SHR-F group at increased stimulation rates (Fig 7). In fact, nearly 62% (8 of 13) of the hearts in the SHR-F group demonstrated an elevation of diastolic [Ca^{2+}] and/or calcium transients and pressure alternans. Although the mechanism underlying alternans is not completely understood, alternans in contractile force are thought to result from variations in intracellular calcium release. Additional evidence of impaired SR function at increasing stimulation rates with heart failure reported by other investigators are fusion of light transients and contraction with increases in both end-diastolic intracellular calcium concentration and end-diastolic tone and a decrease in peak active tension development. Under the same conditions, control muscle preparations did not exhibit fusion or a decline in the force of contraction.

Myothermal data from human hearts indicate a decrease in tension-independent heat thought to be caused in part by reductions in calcium release and the rate of calcium removal. Mulieri et al have studied the effects of heart rate in isolated muscle preparations from normal and failing human hearts and found that in contrast to normal myocardium, force falls at increasing heart rates in failing myocardium; they propose that impairment of function at increasing heart rates reflects an impairment of calcium cycling.

In studies of SR function in partially skinned fibers (40 μg/mL saponin concentration for 20 minutes) from hypertrophied hearts caused by constriction of the abdominal aorta for 6 to 8 weeks, quantitative differences in responses to caffeine and inositol 1,4,5-trisphosphate and in the ability of the SR to accumulate and release calcium and contract have led investigators to conclude that depressed calcium accumulation by the SR may play a role in the modulation of contractile performance in this model of pressure-overload hypertrophy. A similar conclusion was reached in more recent studies of SR function from chemically skinned myocardial fibers obtained from patients with idiopathic dilated cardiomyopathy and...
congestive heart failure due to myocardial infarction in rat. 37

In summary, the present study carried out in isolated perfused hearts demonstrates depressed left ventricular function in failing SHR hearts, whereas peak intracellular calcium levels do not differ from those in control hearts. In response to an increase in perfusate calcium, peak left ventricular pressure increases in proportion to the increase in [Ca 2+], in all groups. In contrast, at increasing heart rates, [Ca 2+], and pressure are maintained or increased in WKY hearts, and [Ca 2+] and pressure decline at higher heart rates in the SHR-NF group. In the SHR-F group, both [Ca 2+], and pressure markedly fall at all increases in heart rate, with the appearance of light and mechanical alternans in many hearts at the higher rates. Thus, the failing heart appears unable to increase peak intracellular calcium in response to increasing rate. Although these data may be consistent with a decreased ability of the SR to take up and release calcium at increased stimulation rates, the role of changes in calcium handling in the progressive development of mechanical dysfunction during the transition from compensated hypertrophy to heart failure remains to be further elucidated.

Acknowledgments

This work was supported in part by US Public Health Service grants HL-31117 and HL-01611 (J.P.M.) and Medical Research Funds from the Department of Veterans Affairs. This work was done during the tenure of a Clinical-Scientist Award from the American Heart Association (C.H.C.).

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Hypertension. 1994;24:347-356
doi: 10.1161/01.HYP.24.3.347

Hypertension is published by the American Heart Association, 7272 Greenville Avenue, Dallas, TX 75231
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Print ISSN: 0194-911X. Online ISSN: 1524-4563

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