Effect of Insulin and Angiotensin II on Cell Calcium in Human Skin Fibroblasts

Giulio Ceolotto, Roberto Valente, Elisabetta Baritono, Stefania Reato, Elisabetta Iori, Alessandra Monari, Roberto Trevisan, Andrea Semplicini

Abstract—We have recently shown that insulin attenuates angiotensin II–induced intracellular Ca\textsuperscript{2+} mobilization in human skin fibroblasts from normotensive subjects. This study was designed to investigate the effects of angiotensin II and the interactions between insulin and angiotensin II on intracellular Ca\textsuperscript{2+} mobilization in skin fibroblasts from patients with essential hypertension. Fibroblasts were obtained from 9 normotensives and 18 hypertensives. Spectrofluorophotometric measurement was performed in monolayers of 24-hour serum-deprived cells. Resting intracellular Ca\textsuperscript{2+} level and angiotensin II–stimulated intracellular Ca\textsuperscript{2+} peak were higher in fibroblasts from hypertensives compared with those from normotensives. The effect of acute insulin exposure was evaluated in fibroblasts from hypertensives subdivided on the basis of insulin sensitivity. In insulin-sensitive hypertensives, insulin significantly blunted the effects of angiotensin II on intracellular Ca\textsuperscript{2+} response, whereas in insulin-resistant patients, insulin did not modify intracellular Ca\textsuperscript{2+} response to angiotensin II. Pertussis toxin, a G\textsubscript{iα}-inhibitor, reduced angiotensin II–stimulated Ca\textsuperscript{2+} peak in insulin-sensitive but not in insulin-resistant hypertensives. In conclusion, the effects of angiotensin II on intracellular Ca\textsuperscript{2+} mobilization are more pronounced in fibroblasts from hypertensives compared with those from normotensives, and the inhibitory effect of insulin is blunted in insulin-resistant hypertensives by a G\textsubscript{iα} pertussis toxin–sensitive abnormality. (Hypertension. 2001;37:1486-1491.)

Key Words: angiotensin II ■ calcium ■ fibroblasts ■ G proteins ■ insulin ■ insulin resistance

Insulin resistance and hyperinsulinemia are often found in hypertensive patients. The relative role of hyperinsulinemia versus that of impaired insulin action in the pathogenesis of hypertension in these patients has not been fully elucidated. In fact, hypertention has been attributed to selective insulin resistance resulting in hyperinsulinemia, with renal sodium retention.\textsuperscript{2} However, it has also been shown that in vivo, insulin infusion induces vasodilation by an endothelium-dependent mechanism\textsuperscript{1} and that the presence of insulin resistance may lead to a partial loss of this insulin-mediated effect, resulting in an increase in pressure.\textsuperscript{4}

In vitro, acute insulin exposure attenuates intracellular calcium (Ca\textsuperscript{2+}) mobilization and contractile response to pressure agents in rat aorta,\textsuperscript{5} in cultured vascular smooth muscle cells,\textsuperscript{6} and in platelets.\textsuperscript{7} This may result from an action on Ca\textsuperscript{2+}-ATPase, voltage- and receptor-operated Ca\textsuperscript{2+}-channels, vasoconstrictor receptors, G-protein, phospholipase C, or inositoltrisphosphate (IP\textsubscript{3})-sensitive Ca\textsuperscript{2+} release channels,\textsuperscript{8} but its precise mechanism remains to be defined.

We have recently found that in human skin fibroblasts from normotensive subjects, insulin blunts the angiotensin (Ang) II–induced Ca\textsuperscript{2+} response by modulating the transmembrane signal transduction.\textsuperscript{9} Most of the known actions of Ang II are exerted through the AT\textsubscript{1} receptor, a G protein–coupled receptor, which stimulates phospholipase C, IP\textsubscript{3}, and Ca\textsuperscript{2+} release,\textsuperscript{10} but recently, it has been shown that Ang II exerts biological actions also through G\textsubscript{iα}-coupled AT\textsubscript{1} receptors.\textsuperscript{11} Insulin acts through its receptor, a protein tyrosine kinase that undergoes a rapid autophosphorylation inducing the phosphorylation of several intracellular protein substrates.\textsuperscript{12} Insulin receptors are also able to phosphorylate some G-protein subunits coupled to vasoconstrictor receptors inhibiting ADP ribosylation of G\textsubscript{iα}-subunit and therefore influencing their intracellular pathway.\textsuperscript{13}

The intracellular cross-talk between insulin and Ang II signaling could be pathophysiologically relevant for Ca\textsuperscript{2+} regulation, and an abnormal interaction may be involved in the association among high blood pressure, insulin resistance, and target organ damage in patients with essential hypertension and diabetes mellitus. The aim of this study was therefore to characterize the effects of insulin on Ang II–stimulated free Ca\textsuperscript{2+} in human skin fibroblasts from insulin-sensitive and insulin-resistant hypertensive patients. In particular, we compared Ca\textsuperscript{2+} responses induced by Ang II in normotensives and hypertensives and the effect of acute insulin exposure and of pertussis toxin on Ang II–induced Ca\textsuperscript{2+} in insulin-sensitive and insulin-resistant hypertensives.
We used cultured skin fibroblasts in vitro, which can be easily obtained through a skin biopsy. They express a variety of different receptors, including G protein–coupled AT₁ receptors, activating well-known pathways of intracellular signal transduction. They can be cultured for several passages in standardized conditions, offering a useful model for the investigation of intrinsic (possibly genetic) defects of cell function, independent of the environmental abnormality caused by hypertension and hyperinsulinemia in vivo. Furthermore, they are actively involved in the process of renal and cardiovascular fibrosis and in the development of target organ damage in hypertensive and diabetic patients.

**Methods**

**Subjects**

We have recruited 9 healthy normotensive volunteers, without a family history of hypertension and diabetes, and 18 patients with essential hypertension from the Hypertension Outpatient Clinic of the Department of Clinical and Experimental Medicine, University Hospital of Padua (Italy). All gave informed consent to the study, which had been approved by the local ethical committee. Arterial hypertension was diagnosed according to the ISH/WHO guidelines (systolic pressure >140 mm Hg and/or diastolic pressure >90 mm Hg). All the hypertensives but three underwent a 75 g oral glucose tolerance test (OGTT) and were classified according to their insulin sensitivity. This parameter was calculated by the glucose/insulin ratio during OGTT and, as previously described, is the mean value minus 2 SD of 20 healthy volunteers which is the mean value minus 2 SD of 20 healthy volunteers (0.53±0.13 mg/mL, mean±SD) (Figure 1). According to this criterion, 9 hypertensives were insulin sensitive and 6 insulin resistant. The clinical characteristics of the normotensives and of the hypertensives are reported in the Table. In comparison to normotensives, both insulin-sensitive and insulin-resistant hypertensives were heavier, but by no means were insulin-sensitive and insulin-resistant hypertensives different.

**Cell Culture**

Human fibroblasts were derived from a skin biopsy taken from the anterior surface of the left forearm by excision, under topical anesthesia with ethyl chloride, and cultured in Nutrient Mixture F-10 HAM medium supplemented with 10% fetal bovine serum, 100 U/mL penicillin, 100 µg/mL streptomycin, and 4 mmol/L glutamine. Cells were seeded onto a 25-cm² flask and incubated at 37°C, and the medium was changed every 2 to 3 days. Fibroblasts obtained from each subject and grown separately were used for the experiments at the 5th passage. They were identified morphologically. In particular, they were strictly diploid and, on morphological confluence, they appeared oriented with respect to one another, forming a typical parallel array of cells with no dividing nuclei visible by microscope. The cells were in the plateau phase of growth, in which there is a steady-state condition with an almost completely ceased cell proliferation, as confirmed by the evaluation of [3 H]-thymidine incorporation in fibroblasts was 10 to 15 times lower than that observed in cells during the log phase of growth and similar to those observed in cells cultured in serum-free medium (data not shown). Moreover, they make type I collagen and were not factor VIII positive. There were no morphological differences in fibroblasts from hypertensives, either insulin-sensitive or insulin-resistant, and normotensive and frequent observations with phase-contrast optic revealed no differences in granularity and vacuolation, which may have a bearing on the health of the culture.

**Measurement of Intracellular Calcium**

The cells (5×10⁴) were seeded onto coverslips (3 cm×1 cm) and allowed to grow to confluence. The medium was then changed to a quiescent medium without serum, and the cells were used after 24

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**Clinical Characteristics of Study Groups**

<table>
<thead>
<tr>
<th>Parameters</th>
<th>Normotensives</th>
<th>Insulin Sensitive</th>
<th>Insulin Resistant</th>
<th>P (NT vs HT)</th>
<th>P (IS vs IR)</th>
</tr>
</thead>
<tbody>
<tr>
<td>No. of subjects (male)</td>
<td>9 (5)</td>
<td>9 (6)</td>
<td>6 (2)</td>
<td>NS</td>
<td>NS</td>
</tr>
<tr>
<td>Age, y</td>
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<td>50±4</td>
<td>54±4</td>
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<td>NS</td>
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<tr>
<td>Systolic blood pressure, mm Hg</td>
<td>110±4</td>
<td>157±3</td>
<td>158±8</td>
<td>&lt;0.01</td>
<td>NS</td>
</tr>
<tr>
<td>Diastolic blood pressure, mm Hg</td>
<td>74±3</td>
<td>99±2</td>
<td>98±2</td>
<td>&lt;0.01</td>
<td>NS</td>
</tr>
<tr>
<td>Body mass index, kg/m²</td>
<td>22±1</td>
<td>29±1</td>
<td>30±1</td>
<td>&lt;0.01</td>
<td>NS</td>
</tr>
</tbody>
</table>

Values are mean±SEM. Insulin sensitivity was defined as reported in Methods. NT indicates normotensives; HT, hypertensives; IS, insulin sensitive; and IR, insulin resistant.
hours. Before starting the experiments, the cells were loaded with 3 μmol/L Fura-2 AM for 1 hour at room temperature. Then, Fura-2 was removed, a physiological medium containing (mmol/L) NaCl 129, KCl 2.8, KH 2 PO 4 0.8, CaCl 2 1, NaHCO 3 8.9, MgCl 2 0.8, glucose 5.6, HEPES 5.6, pH 7.4 was added, and cells were incubated for 30 minutes at room temperature. The coverslip was placed into a quartz cuvette inside a fluorescent spectrophotometer (Shimadzu RF-1501). The cuvette (3 mL), which was held in a thermostatted holder, was superfused with physiological medium at 37°C. A peristaltic perfusion pump delivered solutions at 3 mL/min at the bottom of the cuvette through a glass capillary tube that ran on the side of the coverslip that was opposite from the excitation light and pumped out the solution from the top of the cuvette at the same rate. The turnover half-time of the solution in the cuvette was 0.46 minute; the storage half-time of the solution in the cuvette was 0.46 minute; the storage

Experimental Protocol
Ang II was added to the perfusion solution when baseline fluorescence was stable and the fluorescence measurements continued until Ca 2+ recovered to basal level. To study the effects of insulin incubation, different coverslips seeded with fibroblasts from the same biopsy were used and treated in parallel without and with insulin (100 nmol/L for 20 minutes). Moreover, coverslips prepared in the same way were used to perform parallel experiments in the presence of pertussis toxin (20 ng/mL, overnight).

Chemicals
Ang II, insulin, pertussis toxin, thapsigargin, and Fura-2 AM were from Calbiochem. All other chemicals were from Sigma Aldrich.

Statistical Analysis
Values are expressed as mean±SEM except where otherwise indicated. Statistical comparisons between groups were performed with a Student’s t test for nonpaired variables and ANOVA. A value of P<0.05 was considered statistically significant.

Results
Effect of Ang II, Thapsigargin, and Calcium-Free Media on Cell Calcium
Ca 2+ was measured in quiescent cells before and after exposure to 100 nmol/L Ang II. This concentration has been previously shown to induce a significant and maximal Ca 2+ response. Mean resting cytosolic free calcium and Ca 2+ peak were markedly increased in hypertensives compared with normotensives (78±3 versus 62±3 nmol/L, P<0.01; 224±8 versus 160±12 nmol/L, P<0.01). Basal and Ang II–stimulated Ca 2+ levels in obese (body mass index [BMI]>25 kg/m²) and nonobese subjects did not differ (73±6 versus 64±3 nmol/L, NS, and 178±8 versus 165±9 nmol/L, NS, respectively). Moreover, Ang II–stimulated Ca 2+ increase was not correlated to BMI (r=0.029, NS).

The hypertensives were divided into 2 subgroups according to insulin sensitivity, but basal Ca 2+ and Ang II–induced Ca 2+ peak were not different in fibroblasts from insulin-sensitive and insulin-resistant patients (79±5 versus 75±7 nmol/L and 204±9 versus 212±14 nmol/L, respectively). To determine whether the different Ca 2+ mobilization induced by Ang II in fibroblasts from hypertensives was due to an abnormal Ca 2+ influx, we measured Ang II–induced Ca 2+ response in the absence of extracellular Ca 2+. The removal of extracellular Ca 2+ significantly reduced basal Ca 2+ levels (from 70±5 to 56±3 nmol/L, P<0.05) without affecting Ang II–induced Ca 2+ peak (195±9 versus 193±7 nmol/L, NS). Similar effects have been recently reported in normotensive subjects, thus ruling out a significant role of excessive Ca 2+ influx in inducing a greater Ca 2+ response in hypertensives in comparison to normotensives in these experimental conditions.

Thapsigargin, a Ca 2+-ATPase inhibitor acting on intracellular Ca 2+ pools including the one that is sensitive to IP 3 , has been used to investigate Ca 2+ release from intracellular stores. Ca 2+ release induced by thapsigargin was similar in normotensives and hypertensives (93±8 versus 106±8 nmol/L), suggesting that the different Ca 2+ mobilization is unrelated to thapsigargin-sensitive calcium stores.

Effect of Insulin on Ang II–Stimulated Cell Ca 2+
Acute insulin exposure (100 nmol/L, 20 minutes) markedly reduced Ang II–induced Ca 2+ peak in insulin sensitive but not in insulin-resistant hypertensives (Figure 2). Basal Ca 2+ levels were not modified by insulin in both groups (79±5 versus 78±5 nmol/L and 75±7 versus 79±5 nmol/L, respectively), whereas Ang II–induced Ca 2+ peak was significantly
therefore suggest that the impaired intracellular Ca$^{2+}$ sensitive but not in insulin-resistant hypertensives. We increase of Ca$^{2+}$ mobilization in filbroblasts from 1 insulin-sensitive hypertensive. Figure displays Ca$^{2+}$ response to Ang II either alone (100 nmol/L, thin line), in presence of pertussis tox in (20 ng/mL, 24 hours, thick line), or in presence of both pertussis tox in (20 ng/mL, 24 hours) and insulin (100 nmol/L, 20 minutes) (thick dotted line). Figure reports 1 of 2 identical experiments.

**Effect of Pertussis Toxin on Ang II–Stimulated Ca$^{2+}$**

Pertussis tox in (20 ng/mL, overnight) affected basal Ca$^{2+}$ levels neither in insulin-sensitive nor in insulin-resistant hypertensives. However, it significantly decreased Ang II–induced Ca$^{2+}$ peak in insulin-sensitive hypertensives (from 192±4 to 131±6 nmol/L, P<0.01). This effect was markedly reduced and no longer significant in insulin resistant patients (from 208±10 to 177±9 nmol/L, NS). Furthermore, in insulin-sensitive hypertensives, the blunting effect of insulin on Ang II–induced Ca$^{2+}$ mobilization was completely abolished by pretreatment with pertussis tox in (Figure 3).

**Discussion**

The present study demonstrates that (1) the stimulation of human skin fibroblasts with Ang II evokes a higher Ca$^{2+}$ transient in hypertensives than in normotensives; (2) insulin blunts the effects of Ang II on Ca$^{2+}$ response in insulin-sensitive but not in insulin-resistant hypertensives; (3) pertussis tox in reduces Ang II–stimulated Ca$^{2+}$ peak in insulin-sensitive but not in insulin-resistant hypertensives. We therefore suggest that the impaired intracellular Ca$^{2+}$ regulation in response to insulin in fibroblasts from insulin-resistant hypertensives results from an abnormal regulation or expression of pertussis tox in–sensitive G proteins.

Abnormalities of Ca$^{2+}$ homeostasis have been reported in a variety of cell models of experimental and human arterial hypertension. Resting unstimulated Ca$^{2+}$ and Ca$^{2+}$ transient after several vasoactive calcium-mobilizing hormones in vascular smooth muscle cells, cardiomyocytes, and fibroblasts are higher in spontaneously hypertensive rats than Wistar-Kyoto rats.20,21 In human essential hypertension, an increase in the basal Ca$^{2+}$ levels and a larger agonist-induced increase of Ca$^{2+}$ was also found in circulating blood cells.22–24 Our results confirm these observations because basal Ca$^{2+}$ levels and Ca$^{2+}$ transient induced by Ang II were higher in skin fibroblasts from hypertensives than from normotensives.

Removal of extracellular Ca$^{2+}$ significantly reduced basal Ca$^{2+}$ levels in fibroblasts from hypertensives and abolished the difference in basal Ca$^{2+}$ between cells from normotensives and hypertensives. This suggests that increased calcium influx may contribute to the abnormal regulation of basal Ca$^{2+}$ levels in fibroblasts from hypertensives.

Ang II mediates its effects on Ca$^{2+}$ by IP$_3$–induced mobilization from reticular stores, which induces a rapid and transient Ca$^{2+}$ response, and through entry of Ca$^{2+}$ through Ca$^{2+}$ channels, resulting in a prolonged and sustained Ca$^{2+}$ response.10 The present study shows that under Ca$^{2+}$–free conditions, Ang II–induced Ca$^{2+}$ peak is still higher in fibroblasts from hypertensives compared with normotensives, suggesting that an increased Ca$^{2+}$ influx is not the cause of this abnormal Ca$^{2+}$ response to Ang II. Moreover, the similarity in the thapsigargin-induced Ca$^{2+}$ increase in fibroblasts from normotensives and hypertensives indicates that thapsigargin-sensitive Ca$^{2+}$ stores do not differ. Therefore, our results are consistent with the hypothesis that in human fibroblasts from hypertensives, there is an abnormal Ang II–induced Ca$^{2+}$ mobilization that is independent from intracellular Ca$^{2+}$ pools and suggest that these cells have an increased responsiveness to Ang II. Although we did not determine Ang II receptor density, and therefore a receptor upregulation in hypertensives cannot be excluded, data on confluent cultured cells from spontaneously hypertensive rats and Wistar-Kyoto rats did not show differences in AT$_1$ receptor density.25 Therefore, we propose that the abnormalities of Ca$^{2+}$ response shown in the present work in hypertensives are accounted for an abnormal signal transduction of Ang II, probably located at postreceptor level. Similar alterations in Ca$^{2+}$ homeostasis were present in insulin-sensitive and in insulin-resistant hypertensive patients, suggesting that arterial hypertension rather than insulin resistance accounts for the abnormal calcium handling.

The second aim of our study was to compare the effects of acute insulin exposure on Ang II–induced Ca$^{2+}$ mobilization in fibroblasts from insulin-sensitive and insulin-resistant hypertensives.

Previous studies, in vitro, have shown that insulin attenuates Ca$^{2+}$ mobilization in different cell types.6,7,26 In vascular smooth muscle cells, insulin inhibition of agonist-induced Ca$^{2+}$ transient is due to inhibition of voltage- and receptor-operated Ca$^{2+}$ channels,27 inhibition of IP$_3$–induced Ca$^{2+}$ release from intracellular stores,28 and stimulation of plasma membrane Ca$^{2+}$–ATPase–mediated Ca$^{2+}$ efflux.29 Insulin may also inhibit transmembrane signal transduction of vasoconstrictor agents acting on receptor-coupled G proteins, phospholipase C, or IP$_3$–sensitive Ca$^{2+}$ release channels.13,30

The inhibitory effect of insulin on Ca$^{2+}$ responses to agonists is reduced in vascular smooth muscle cells from insulin-resistant Zucker obese rats compared with their lean controls, as the result of an impairment in the stimulation of both plasmalemma and sarcoplasmic reticulum Ca$^{2+}$–ATPase.31 Furthermore, the inhibitory effect of insulin on
Ca^{2+} mobilization induced by Ang II and endothelin-1 was impaired in platelets from patients with essential hypertension in proportion to the degree of hyperinsulinemia.32,33

In this study, insulin attenuated Ang II–induced Ca^{2+} mobilization in fibroblasts from insulin-sensitive but not from insulin-resistant hypertensives. This indicates that insulin resistance but not hypertension is responsible for the reduced effect of insulin on Ang II–induced Ca^{2+} transient in insulin-resistant hypertensives.

Such effect is unlikely to be caused by insulin receptor downregulation in cultured cells from insulin-resistant patients. In adipose tissue, in muscle, and in red blood cells, there is some evidence for downregulation of insulin receptors in insulin-resistant patients because of altered insulin-stimulated tyrosine kinase activity.34 However, these abnormalities could be reversed both in vivo and in vitro, thereby suggesting that they are secondary rather than intrinsic molecular alterations. Furthermore, it has been shown that in long-term cultured skin fibroblasts, as the case of our study, insulin receptors are not reduced in insulin-resistant patients.35 The majority of the studies, therefore, now agrees that insulin resistance is caused by a postreceptor defect.

We recently reported that insulin blunts agonist-induced Ca^{2+} responses in human skin fibroblasts, an observation that was consistent with a specific action of insulin on the agonist-sensitive Ca_{i}^{2+} release cascade, and proposed that insulin can influence the Ang II transmembrane signal transduction.9 This hypothesis was supported by the fact that tyrosine kinase signaling pathways may modulate Ang II–induced Ca_{i}^{2+} transients in rat vascular smooth muscle cells.36 We still do not know the exact site of action and nature of this inhibition, but the results of the present study suggest that insulin can act at the AT_{1} receptors coupled to G protein. In fact, it has been shown that Ang II negatively modulates L-type Ca^{2+} channels through a pertussis toxin–sensitive G protein37 and stimulates T-type Ca^{2+} channels through G protein in bovine adrenal glomerulosa cells.38 However, our previous study suggested that insulin modulates Ca_{i}^{2+} release and not Ca_{a}^{2+} channels in human fibroblasts, because the effect of insulin persisted in Ca_{a}^{2+}-free media.9

Recent studies have demonstrated an interaction between the insulin receptor and its signaling on one side and G protein–coupled receptors and their signaling pathways on the other.13,30 In a mouse harboring inducible expression of RNA antisense to the gene encoding the G protein G_{i,2a}, G_{i,2a} deficiency in the liver and in the adipose tissue led to hyperinsulinemia, impaired glucose tolerance, and resistance to insulin.39 Interestingly, G_{i,2a} deficiency also increased protein tyrosine phosphatase activity and thereby attenuated insulin-stimulated tyrosine phosphorylation of the insulin receptor substrate 1 (IRS-1). Moreover, mice made transgenic for G_{i,2a}, which exhibited overexpressed G protein, have a more efficient utilization of glucose than control animals. Finally, it has been also demonstrated that in adipocytes, the downregulation of G_{i,2a} subtype by A_{2a} adenosine receptors decreased both insulin-sensitive glucose transport and tyrosine kinase activity of the insulin receptor.40

We have shown that pertussis toxin blunts Ang II–induced Ca_{i}^{2+} mobilization in insulin-sensitive hypertensives, but it does not affect Ca^{2+} mobilization to a significant extent in insulin-resistant hypertensives. Pertussis toxin uncouples G_{i} from the receptor through ADP ribosylation of its carboxyl terminus, inhibiting the function of this G protein.41 The reduced effect of pertussis toxin paralleled that of insulin on Ca^{2+} mobilization in insulin-resistant hypertensives. Furthermore, in the presence of pertussis toxin, insulin was no longer able to attenuate the Ang II–dependent Ca^{2+} mobilization in insulin-sensitive hypertensives. The whole of these data suggests that (1) a defect in the regulation or in the expression of pertussis toxin–sensitive G_{i} is associated to insulin resistance, and (2) G_{i} is involved in insulin inhibition of Ang II–induced Ca^{2+} mobilization. Therefore, we suggest that a defective G_{i} signal transduction pathway may be a potential site for the link between insulin resistance and the inability of insulin to blunt Ang II calcium response in fibroblasts from insulin-resistant hypertensive patients.

Conclusions

We have demonstrated an increased responsiveness to Ang II in hypertensive patients. Insulin modulates Ca^{2+} mobilization induced by Ang II through a pertussis toxin–sensitive G_{i} mechanism. In fibroblasts from insulin-resistant patients, insulin failed to attenuate the increased responsiveness to Ang II. This alteration was seen after several passages in vitro. Therefore, these in vitro phenotypic characteristics of fibroblasts are likely to be genetically determined and independent of the in vivo metabolic and hemodynamic abnormalities. A disturbed balance between Ang II and insulin signaling might be relevant for the development target organ damage in insulin-resistant patients with hypertension and diabetes.

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