Angiotensin II Regulation of AT₁ and D₃ Dopamine Receptors in Renal Proximal Tubule Cells of SHR

Chunyu Zeng, Laureano D. Asico, Xiaoli Wang, Ulrich Hopfer, Gilbert M. Eisner, Robin A. Felder, Pedro A. Jose

Abstract—Dopamine and angiotensin II negatively interact to regulate sodium excretion and blood pressure. D₃ dopamine receptors downregulate angiotensin type 1 (AT₁) receptors in renal proximal tubule cells from normotensive Wistar-Kyoto rats. We determined whether AT₁ receptors regulate D₃ receptors and whether the regulation is different in cultured renal proximal tubule cells from normotensive and spontaneously hypertensive rats. Angiotensin II (10⁻⁸M/24 hours) decreased D₃ receptors in both normotensive (control, 36 ± 3; angiotensin II, 24 ± 3 U) and hypertensive (control, 30 ± 3; angiotensin II, 11 ± 3 U; n = 9 per group) rats; effects that were blocked by the AT₁ receptor antagonist, losartan (10⁻⁷M/24 hours). However, the reduction in D₃ expression was greater in hypertensive (60 ± 10%) than in normotensive rats (32 ± 9%). In normotensive rats, angiotensin II (10⁻⁸M/24 hr) also decreased AT₁ receptors. In contrast, in cells from hypertensive rats, angiotensin II increased AT₁ receptors. AT₁ and D₃ receptors co-immunoprecipitated in renal proximal tubule cells from both strains. Angiotensin II decreased D₃/AT₁ receptor co-immunoprecipitation similarly in both rat strains, but basal D₃/AT₁ co-immunoprecipitation was 6 times higher in normotensive than in hypertensive rats. Therefore, AT₁ and D₃ receptor interaction is qualitatively and quantitatively different between normotensive and hypertensive rats; angiotensin II decreases AT₁ expression in normotensive but increases it in hypertensive rats. In addition, angiotensin II decreases D₃ expression to a greater extent in hypertensive than in normotensive rats. Aberrant interactions between D₃ and AT₁ receptors may play a role in the pathogenesis of hypertension. (Hypertension. 2003;41[part 2]:724-729.)

Key Words: receptors, dopamine • receptors, angiotensin II • rats, spontaneously hypertensive • kidney

The proximal tubule is a major site of salt and water reabsorption in the mammalian nephron, with up to 60% of the filtrate reabsorbed in this segment.¹,² Renal proximal tubule (RPT) function is under hormonal control, with angiotensin II stimulating sodium reabsorption via activation of apical Na⁺/H⁺ exchanger and basolateral [Na⁺]/[K⁺]ATPase, and with dopamine inhibiting reabsorption by inhibiting these proteins.³–⁵

The paracrine regulation of sodium reabsorption in the proximal tubule by the renin-angiotensin system occurs via several angiotensin receptor subtypes (AT₁, AT₂, and AT₄).³–⁸ The activation of AT₁ receptors by angiotensin II increases sodium transport, whereas the activation of AT₂ and AT₄ receptors decreases sodium reabsorption in this nephron segment.⁵–⁸ However, under physiological conditions, the major effect of angiotensin II on sodium transport is stimulatory, via AT₁ receptors.⁵–⁸

The dopaminergic system also exerts a paracrine regulatory role on renal sodium transport in the proximal tubule via several receptor subtypes. Dopamine receptors, pharmacologically grouped into D₁-like (D₁ and D₅) and D₂-like (D₂, D₃, and D₄) receptors, as with the angiotensin II receptors, are expressed in brush border and basolateral membranes of RPTs. In contrast to the stimulatory effect of angiotensin II on sodium transport in RPTs, the major consequence of the activation of dopamine receptors is an inhibition of sodium transport via D₁-like and D₃ receptors.³,⁴,⁹–¹¹

Previous studies have indicated that dopamine and angiotensin II subserve counteracting functions in the proximal tubule. Inhibition of RPT angiotensin II production or blockade of AT₁ receptors increases the natriuretic effect of the D₁-like agonist fenoldopam.⁹ D₁-like and D₂-like receptor agonists also antagonize the stimulatory effect of angiotensin II, acting via AT₁ receptors, on RPT luminal sodium transport.¹²,¹³ The renal vasoconstrictor effect of angiotensin II can also be antagonized by D₁-like receptor agonists.¹⁴ Alterations in the balance of the responsiveness of the proximal tubule to angiotensin II and dopamine have important impli-
cations on net sodium reabsorption. Indeed, an altered interaction between angiotensin II and dopamine has been implicated in genetic hypertension.4,14

In a previous study, we showed that D3 and AT1 receptors co-localized in immortalized RPT cells from normotensive Wistar-Kyoto rats (WKY) and that D3 dopamine receptors decreased AT1 receptor protein expression in these cells.15 We have found that these immortalized RPT cells have characteristics similar to freshly obtained RPT brush border membranes and RPTs, at least with regard to D3 receptors and responses to G-protein stimulation.16–18 The present studies were designed to determine whether angiotensin II regulates AT1 and D3 receptor expression and whether the regulation is different in RPT cells from WKY and spontaneously hypertensive rats (SHR).

Methods

Cell Culture.

All studies have been approved by the Georgetown University Animal Care and Use Committee. Immortalized RPT cells obtained from 4- to 8-week-old WKY and SHR (Charles River Laboratory, Wilmington, Mass) were cultured at 37°C in 95% air/5% CO2 atmosphere in Dulbecco modified Eagle’s medium/F-12 culture media, as previously described.17,19 The cells (80% confluence) were extracted in ice-cold lysis buffer (PBS with 1% NP40, 0.5% sodium deoxycholate, 0.1% SDS, 1 mmol/L EDTA, 1 mmol/L EGTA, 1 mmol/L PMSF, 10 μg/mL aprotinin, and 10 μg/mL leupeptin), sonicated, kept on ice for 1 hour, and centrifuged at 16 000 g for 30 minutes. The supernatants were stored at −70°C until use for immunoblotting and/or immunoprecipitation.

Immunoblotting

The antibodies are polyclonal purified antipeptides. The amino acid sequence for the immunogenic peptide (rabbit anti-human AT1 receptor antibody) is QDDCPKAGRHC, amino acids 15–24 of the AT1 receptor. The specificity of this antibody to the AT1 receptor has been reported.20 The amino acid sequence of the peptide for the rabbit anti-rat D3 receptor antibody corresponds to the third intracellular loop of the D3 receptor.11,21 We have reported the specificity of this D3 receptor antibody.11 Rat RPT cells were treated with vehicle (dH2O), angiotensin II, or an AT1 receptor antagonist (losartan) at the indicated concentrations and times.22 Immunoblotting was performed as previously reported, except that the transblots were probed with the D3 (1:250) or the AT1 receptor antibody (1:400).23 The amount of protein transferred onto the membranes was determined by Ponceau-S staining and immunoblotting for α-actin.

Immunoprecipitation

RPT cells were incubated with vehicle or angiotensin II (10^{-8} M) for 24 hours, as described above. The cells were lysed with ice-cold lysis buffer for 1 hour and centrifuged at 16 000 g for 30 minutes. Equal amounts of lysates, except as indicated (300 μg protein/mL supernatant for RPT cells from WKY and 900 μg protein/mL supernatant for RPT cells from SHR) were incubated with affinity-purified anti-D3 receptor antibody (1 μg/mL) for 1 hour and G-protein agarose at 4°C for 12 hours. The immunoprecipitates were pelleted and washed 4 times with lysis buffer. The pellets were suspended in sample buffer, boiled for 10 minutes, and subjected to immunoblotting with the AT1 receptor antibody. To determine the specificity of the bands, normal rabbit IgG (negative control) and AT1 receptor antibody (positive control) were used as immunoprecipitants, instead of the D3 receptor antibody. The density of the bands were quantified by densitometry, using Quantiscan as previously reported.17,19,24

Materials

Rabbit anti-rat D3 and anti-human AT1 receptor antibodies were purchased from Alpha Diagnostic International (D3R12A) and Santa Cruz Biotechnology Inc (sc-1173), respectively. Angiotensin II (H-17057002) was purchased from Peninsula Laboratory Inc. Losartan was a gift from Merck & Co (Philadelphia, Penn). Other chemicals for various buffers were of the highest purity available and purchased either from Sigma or Gibco.

Statistical Analysis.

The data are expressed as mean±SEM. Comparison within groups was made by ANOVA for repeated measures (or paired t test when only 2 groups were compared), and comparison among groups (or t test when only 2 groups were compared) was made by ANOVA with the Duncan test. A value of P<0.05 was considered significant.

Results

Specificity of Receptor Antibodies.

The bands for the AT1 receptor (~45 kDa) and D3 receptor (~45 kDa) were found in RPT cells, as previously reported.11,15 Another band of ~40 kDa was noted for the D3 receptor and probably represents a degradation product of the 45-kDa D3 receptor.21 The bands were specific because they were completely blocked when the antibodies were preabsorbed by the immunizing peptide (Figures 1A and 1B, lanes 1 and 2). HEK293 cells, transfected with rat D3 receptor cDNA, expressed bands similar to those noted in RPT cells and were also blocked by immunizing peptide (Figure 1B, lane 1). The pattern of D3 receptor immunoblotting is similar to our previous report.11,15

AT1 Receptors Decrease D3 Receptor Expression in Rat RPT Cells from SHR and WKY

Angiotensin II decreased D3 receptor expression in a concentration- and time-dependent manner in RPT cells from WKY. The inhibitory effect was evident at 10^{-8} M, with a 50% decrease (EC50) in D3 receptor expression at (4.1×10^{-10} M) (Figure 2A). The inhibitory effect of angiotensin II (10^{-8} M) was noted as early as 8 hours and maintained for at least 30 hours (Figure 2B). In RPT cells from SHR, angiotensin II (10^{-8} M/24 hours) also decreased D3 expression (SHR: control, 30±3; angiotensin II, 11±3 density units [DU]; WKY: control, 36±3; angiotensin II, 24±3 DU; n=9 per group), but the reduction was greater in SHR (60±10%) than in WKY (32±9%) (Figure 2C). Basal D3 receptor (45 KDa) levels were also lower in SHR compared with WKY, similar to our previous report.11,15

The specificity of angiotensin II as an AT1 receptor agonist was also determined by studying the effect of the AT1 receptor antagonist losartan. Consistent with the study shown in Figures 2A and 2B, angiotensin II (10^{-8} M/24 hours), decreased D3 receptor expression (control, 25±2 DU; angiotensin II, 19±2 DU; n=8, P<0.05). The AT1 receptor antagonist, losartan (10^{-8} M), by itself, had no effect on D3 receptor expression (29±3 DU) but reversed the inhibitory effect of angiotensin II on D3 receptor expression (28±2 DU) (Figure 2D).

Angiotensin II Decreases AT1 Receptor Expression in Rat RPT Cells From WKY But Increases It From SHR

To investigate the effect of angiotensin II on its own receptor, RPT cells were incubated with angiotensin II (10^{-8} M) for 24
hours. Immunoblots showed that angiotensin II decreased its own receptor expression in RPT cells from WKY (control, 26±2 DU; angiotensin II, 17±3 DU; P<0.05, n=5) but increased it in RPT cells from SHR (control, 24±2 DU; angiotensin II, 38±5 DU; n=5 per group) (Figure 3).

**AT**$_1$ Receptor Co-Immunoprecipitates With the D$_3$ Receptor in Rat RPT Cells

To determine whether there is a physical interaction between the D$_3$ and the AT$_1$ receptor, additional experiments were performed. D$_3$ receptors were first immunoprecipitated with anti-D$_3$ receptor antibodies and then probed with anti-AT$_1$ receptor antibodies. As shown in Figure 4, the band of 45 kDa, representing the co-immunoprecipitated D$_3$ and AT$_1$ receptors, was decreased by a 24-hour treatment of angiotensin II (10$^{-8}$M) in both RPT cells from WKY and SHR (WKY: control, 45±10 DU; angiotensin II, 16±5 DU; n=7, P<0.05; SHR: control, 7±2 DU; angiotensin II, 3±1 DU; n=7, P<0.05). However, basal co-immunoprecipitation was

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**Figure 1.** Specificity of AT$_1$ and D$_3$ antibodies. A, Immunoblot of AT$_1$ receptors in RPT cells from WKY. Cell lysate protein (100 μg) was subjected to immunoblotting with anti-AT$_1$ receptor antibody (1:400). The 45-kDa band was completely blocked when the antibody was preadsorbed with immunizing peptide (1:10 w/w). B, Immunoblot of D$_3$ receptors. Cell lysate protein (100 μg) from HEK293 cells transfected with rat D$_3$ receptor cDNA (lane 1) and WKY RPT cells (lane 2) had similar bands (45 kDa). Another band of ~40 kDa, probably representing a degradation product of the D$_3$ receptor, was also noted. The bands were no longer visualized when the antibody was preadsorbed with immunizing peptide (1:10 w/w).

**Figure 2.** Effect of angiotensin II on D$_3$ receptor expression in RPT cells from SHR and WKY. A, Concentration-response of D$_3$ receptor expression in RPT cells from WKY treated with angiotensin II. Immunoreactive D$_3$ receptor expression was determined after 24-hour incubation with the indicated concentrations of angiotensin II. Results are expressed as relative density units (n=5; *P<0.05 vs control [C], ANOVA, Duncan test). B, Time-course of D$_3$ receptor expression in RPT cells from WKY treated with angiotensin II. The cells were incubated for the indicated times with 10$^{-8}$M angiotensin II. Results are expressed as relative density units (n=7; *P<0.05 vs control [0 time], ANOVA,
greater in RPT cells from WKY than from SHR, similar to our previous report.\textsuperscript{15}

**Discussion**

There are several novel observations in the present study. First, we show that angiotensin II, via AT\textsubscript{1} receptors, decreases D\textsubscript{3} receptor expression in rat RPT cells from WKY. This effect is exerted at the AT\textsubscript{1} receptor because an AT\textsubscript{1} antagonist, losartan, completely blocks the effect of angiotensin II. Second, angiotensin II also decreases D\textsubscript{3} receptors in SHR, but the reduction in D\textsubscript{3} expression is greater in SHR than in WKY. Third, angiotensin II decreases AT\textsubscript{1} receptor expression in RPT cells from WKY but increases it in cells from SHR. Fourth, AT\textsubscript{1} receptors co-immunoprecipitate with D\textsubscript{3} receptors in rat RPT cells. Angiotensin II stimulation decreases the physical interaction between AT\textsubscript{1} and D\textsubscript{3} receptors in rat RPT cells from both SHR and WKY to the same degree. However, the basal level of AT\textsubscript{1}/D\textsubscript{3} co-immunoprecipitation is much greater in WKY compared with SHR.

The effect of the D\textsubscript{3}-like receptor, by itself, on renal sodium transport is controversial; stimulation, no effect, and even inhibition have been reported.\textsuperscript{4} The reason for the apparent discrepancies may be related to the different experimental conditions and the specific D\textsubscript{3}-like receptor involved.\textsuperscript{3,4} However, under certain circumstances, such as during moderate sodium loading, D\textsubscript{1} and D\textsubscript{2}-like receptors can synergistically inhibit sodium transport.\textsuperscript{11} All of D\textsubscript{2}-like receptors exist in kidney.\textsuperscript{3,4} The D\textsubscript{2\textsubscript{long}} receptor heterologously expressed in fibroblasts (LTK\textsuperscript{-} cells) increases Na\textsuperscript{+}K\textsuperscript{+} ATPase activity,\textsuperscript{4} but the D\textsubscript{3} receptors expressed in the kidney are probably in prejunctional nerves because D\textsubscript{2} mRNA was not detected in microdissected rat RPTs or rat RPT cells (M.J. Bek, I. Yamaguchi, Z. Yang, and P.A. Jose, unpublished data, 2000). The D\textsubscript{1} receptor is expressed mainly in collecting ducts.\textsuperscript{4,25} In contrast, D\textsubscript{2} receptors, like the D\textsubscript{1} and D\textsubscript{3} receptors, are expressed in the proximal tubule.\textsuperscript{11,26} Thus, the D\textsubscript{1} receptor may be the major D\textsubscript{2}-like subtype receptor expressed in RPTs. Quinpirole, a D\textsubscript{2}-like receptor agonist with a greater selectivity for the D\textsubscript{3} and D\textsubscript{2} receptor over the D\textsubscript{1} receptor, decreases sodium excretion in dogs, whereas 7-OH-DPAT, a D\textsubscript{2}-like receptor with a greater selectivity for D\textsubscript{1} than D\textsubscript{2} or D\textsubscript{3} receptors, increases both glomerular filtration rate and sodium and water excretion in rats.\textsuperscript{27,28} PD128907,\textsuperscript{27} which has a greater selectivity for D\textsubscript{3} than for D\textsubscript{2} or D\textsubscript{3} receptors also increased sodium excretion in salt-loaded but not salt-depleted WKY. These studies suggest that D\textsubscript{3} receptors, under certain circumstances, mediate a natriuresis, an effect that seems to be lost in SHR (C. Zeng, L.D. Asico, and P.A. Jose, unpublished data, 2002). Whether the defective effect of D\textsubscript{3} receptors on sodium excretion in SHR is primary or secondary to interactions with other receptors is not known. However, preliminary studies from our laboratory indicate that the D\textsubscript{3} agonist, PD128907, increases D\textsubscript{3} receptor in RPT cells from WKY but has no effect in RPT cells from SHR.\textsuperscript{15,29}

We have reported that mice lacking both D\textsubscript{3} receptor alleles developed systemic hypertension and a decreased ability to excrete a systemic sodium load.\textsuperscript{10} In addition, renal renin activity and AT\textsubscript{1} receptor expression are much higher in the homozygous than in wild-type mice.\textsuperscript{10,15} Z1046, a dopaminergic agonist with a greater selectivity for D\textsubscript{1} and D\textsubscript{3} receptors over the other dopamine receptors, also increases sodium and water excretion in anesthetized WKY, but this effect is abrogated in SHR.\textsuperscript{11,30} These data suggest that a D\textsubscript{3} receptor in rat RPT cells from both SHR and WKY. The cells were incubated at the indicated times and concentrations. Results are expressed as relative density units (n = 5 or 7; \textit{P}<0.05 vs control; ANOVA, Duncan test). C, Differential effects of angiotensin (10\textsuperscript{-8} M/24 hours) on D\textsubscript{3} receptor expression in RPT cells from both SHR and WKY. The cells were incubated at the indicated times and concentrations. Results are expressed as relative density units (n = 5 or 7; \textit{P}<0.05 vs control; ANOVA, Duncan test). D, Effect of angiotensin II (Ang II) and an AT\textsubscript{1} receptor antagonist (losartan) on D\textsubscript{3} receptor expression in WKY RPT cells. The cells were incubated with the indicated reagents (10\textsuperscript{-8} M angiotensin II and 10\textsuperscript{-6} M losartan) for 24 hours. Results are expressed as relative density units (n = 8; \textit{P}<0.05 vs others, ANOVA, Duncan test).
receptor defect may play a role in the pathogenesis of some forms of hypertension, most likely via an impaired renal excretory capacity for sodium or via an impaired inhibition/interaction with renin-angiotensin system.

The current study shows that AT₁ receptor stimulation decreases D₃ receptor expression in RPT cells from both SHR and WKY, but the reduction in D₃ expression is greater in SHR than in WKY. The greater reduction in D₃ receptor expression in RPT cells caused by angiotensin II in SHR than in WKY may be caused by an increased activity of the AT₁ receptor in this rat strain. Indeed young SHR (5 to 7 weeks), the same age group as the rats in which the RPT cells are obtained for immortalization, RPT sodium transport is greater in SHR than in WKY. AT₁ receptor stimulation results in a decreased renal proximal tubular sodium reabsorption to a greater extent in SHR than in WKY.

We also found that angiotensin II decreases AT₁ receptor in cells from WKY. There are a few studies on the effect of angiotensin II on AT₁ receptor expression in RPT cells. In rabbit RPT cells, a 16-hour incubation with angiotensin II dose-dependently increases AT₁ receptor expression, assessed by radioligand binding. However, the systemic administration of angiotensin II (10⁻⁶M) increases AT₁ receptor mRNAs in rat and rabbit RPT cells are also increased by angiotensin II (10⁻⁶M). However, the systemic administration of angiotensin II that increases systemic blood pressure does not alter total renal AT₁ receptor expression, determined by immunoblotting, but decreases AT₁ receptor expression (radioligand autoradiography) in glomeruli and inner stripe of the outer medulla. Three days after the infusion of angiotensin II, there is a tendency for a decrease in AT₁ expression in the whole kidney and in RPTs, but the changes are small and do not reach statistical significance, results that could be taken to support our data. The differences in the results of these studies cannot be explained at this time.

In contrast to the inhibitory effect of angiotensin II on AT₁ expression in RPT cells from WKY, angiotensin II produces the opposite effect in RPT cells from SHR. The different effects of AT₁ agonist on D₃ receptor expression, and its own receptor, could be involved in the pathogenesis of hypertension in SHR. Decreased renal D₃ receptor expression and function have been reported in the Dahl salt-sensitive rat. However, it is not known whether the D₃ receptor dysfunction in these rodent models of genetic hypertension is primary or secondary to a dysfunction of renal D₁ receptors. An uncoupling of the D₃ receptor from its G-protein/effecter complex in the kidney has been shown to be important in the pathogenesis of genetic hypertension. The mechanism for the decrease in D₃ receptor expression in the 45-kDa species caused by AT₁ receptors was not studied. However, the D₃ receptor, a member of the D₂-like receptor, has been reported to decrease gene expression by translational and/or posttranslational protein modification. It is possible that AT₁ receptors regulate D₃ receptor expression by similar mechanisms.

We now report that AT₁ and D₃ receptors can directly interact with each other. In the current studies, the basal level of AT₁/D₃ co-immunoprecipitation is much lower in RPT cells from SHR than from WKY. AT₁ receptor stimulation results in a decreased interaction between AT₁ and D₃ receptors. The paucity of D₃ receptors on the RPT cell membranes could have been responsible for the decreased basal amount of D₃ and AT₁ receptor co-immunoprecipitation in SHR. Therefore, we tripled the amount of protein used for the immunoprecipitation studies in SHR. In a previous report, we found that doubling the amount of loaded protein from renal cortical membranes from SHR approximate D₃ expression in SHR to those observed in WKY. Moreover, reversing the antibodies used for immunoprecipitation and immunoblotting, by using the AT₁ antibody for immunoprecipitation and the D₃ antibody for immunoblotting, gives similar results (data not shown). Therefore, it is unlikely that the low abundance of D₃ receptors in SHR could have caused the decreased amounts of D₃ and AT₁ receptor co-immunoprecipitation in this rat strain. We suggest that the strain differences could be caused by a differential expression in adaptor or interacting proteins. The ability of angiotensin II to decrease the amount of D₃/AT₁ co-immunoprecipitation to the same degree in WKY and SHR suggests that the putative adaptor or interacting protein normally responds to angiotensin II. Further studies are needed to determine whether the decreased interaction between these 2 receptors is a direct or an indirect mechanism, possibly by the alteration of adaptor proteins. Thus, an adaptor protein for D₃ and D₃ receptors (protein 4.1N) has been recently identified that is important in their localization in plasma membranes. Others have also shown that several G protein–coupled receptors interact directly with each other resulting in homo- or hetero-oligomerization.

In summary, we have demonstrated that AT₁ receptors negatively regulate the expression of D₃ receptors in rat RPT cells. Furthermore, AT₁ and D₃ receptors interact in RPT cells, but this interaction is impaired in SHR.

Perspectives

The dopaminergic and renin-angiotensin systems are 2 important systems that regulate sodium excretion and blood pressure. D₁-like and D₂-like dopamine receptors synergistically inhibit sodium reabsorption in RPTs. The receptor subtypes mediating these synergistic effects are, most probably, D₁ and D₃ receptors because D₁ and D₃ receptors are the major dopamine receptors in the RPT cells. We have preliminary data showing an inhibition of Na⁺-K⁺ ATPase activity by the stimulation of both D₁ and D₃ receptors (data not shown). In SHR, the synergistic interaction between D₁-like and D₂-like receptors is impaired. D₁ receptor expression (45 kDa), in renal brush border membranes and RPT cells, is lower in rodent hypertensive models relative to normotensive controls. Despite similar abundance of D₁ receptors in RPTs in human essential and rodent genetic hypertension, D₁ receptor function is impaired because of increased G protein–coupled receptor kinase (GRK) 4 and decreased phospholipase 2A (PP2A) activities. It is possible that GRK₁ and PP2A have a similar effect on D₁ receptors, causing the decrease in D₁ receptor expression and function. Activation of D₁ receptor or D₁ receptor decreases AT₁ receptor expression in RPT cells from WKY, and those effects are impaired in SHR. AT₁ receptor activity is higher in SHR; however, it has not been determined whether the increased AT₁ receptor expression or function is primary or secondary to derangements of other systems. We
hypothesize the increased AT1 receptor activity in SHR is secondary to an impaired dopaminergic system, related to GRK4, D1, and D3 receptors. This hypothesis will be tested in future studies.

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References


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