C242T CYBA Polymorphism of the NADPH Oxidase Is Associated With Reduced Respiratory Burst in Human Neutrophils

Keith E. Wyche, Shaoshan S. Wang, Kathy K. Griendling, Sergey I. Dikalov, Harland Austin, Swapna Rao, Bruno Fink, David G. Harrison, A. Maziar Zafari

Abstract—Oxidative stress contributes to the pathogenesis of atherosclerosis. p22^{phox}-based NAD(P)H oxidases exist in the vessel wall, acting as important superoxide-generating systems in the vasculature. Some studies have identified reduced atherosclerosis in the presence of the C242T CYBA polymorphism, whereas others have not. Because vascular p22^{phox} is identical to neutrophil p22^{phox}, we studied the association between the C242T, A640G, and −930^{AG} CYBA polymorphisms and the quantity of superoxide produced from neutrophils isolated from healthy adults to determine if these polymorphisms had any functional impact on NADPH oxidase function. Neutrophils were isolated from 90 subjects by Percoll density gradient centrifugation. Genotypes were determined by polymerase chain reaction (PCR) and restriction mapping, as well as real-time PCR. The oxidative burst was stimulated with phorbol 12-myristate 13-acetate. Superoxide was quantified using the superoxide dismutase inhibitable oxidation of the spin probe hydroxylamine 1-hydroxy-3-carboxy-pyrrolidine, detected by electron paramagnetic resonance. Superoxide production was significantly affected by the C242T polymorphism, being 8.7±0.7, 7.9±0.6, and 5.9±1.2 μmol/L per minute per 10^6 neutrophils for the C242T CC, CT, and TT genotypes, respectively (P<0.05). In contrast, the A640G and the −930^{AG} polymorphisms did not alter the neutrophil respiratory burst. Phagocytic respiratory burst activity in homozygous individuals with the T allele of the C242T CYBA polymorphism is significantly lower than of wild-type carriers and heterozygous individuals. Because p22^{phox} exists in both the neutrophil and vessel wall, vascular oxidative stress is likely diminished in individuals with this polymorphism. (Hypertension. 2004;43:1246-1251.)

Key Words: atherosclerosis • neutrophils • polymorphism • risk factors

Oxidative stress plays a significant role in the pathogenesis of coronary artery disease (CAD) by altering vasomotor tone, enhancing atherosclerosis and contributing to hypertension. It is clear that vascular cells can produce reactive oxygen species (ROS), including superoxide, hydrogen peroxide, and nitric oxide (NO). Currently, attention is focused on p22^{phox}-based NAD(P)H oxidases as major sources of ROS in blood vessels. The molecular structure of vascular Na(D)H oxidases is similar to the phagocytic NADPH oxidase that is responsible for the respiratory burst. The neutrophil oxidase is a multicomponent enzyme system composed of membrane and cytosolic components. The membrane components, known as the cytochrome b558, comprise 2 subunits, p22^{phox} and gp91^{phox}. The cytoplasmic components include p47^{phox}, p67^{phox}, p40^{phox}, and rac-2. The p22^{phox} subunit is central to the normal functioning of the oxidase because it stabilizes the large subunit and serves a docking function for the cytosolic factors. Functionally, p22^{phox} is critical for the activity, because antisense inhibition of p22^{phox} expression in vascular smooth muscle cells decreases superoxide and hydrogen peroxide production in response to angiotensin II by these cells. In human coronary arteries, p22^{phox} is expressed in atherosclerotic areas more intensely than in nonatherosclerotic areas. At position 242 of CYBA, the gene encoding p22^{phox}, have been identified. The C242T polymorphism is located on chromosome 16q24, exon 4, with no frank amino acid substitution. The A640G gene polymorphism is located in the 3′ untranslated region of CYBA with no frank amino acid substitution. The −930^{AG} gene polymorphism is located in the p22^{phox} promoter, and is associated with higher promoter activity. Given the critical role of the p22^{phox}-based vascular NAD(P)H oxidases in ROS generation in the vasculature, it is conceiv-
able that altered activity of p22<sup>Hello</sup> may modify risk for atherosclerosis. Indeed, the association of the CYBA genotypes with CAD was reported previously in clinical association studies, and the results were contradictory. Inoue et al reported a significant protective effect of the C242T polymorphism on the presence of CAD in Japanese subjects. In contrast, Cahilly et al found the C242T polymorphism to be a marker of progression of CAD. We and others found no association between C242T CYBA polymorphism and the severity of CAD detected by coronary angiography, suggesting no functional role of the polymorphism in prevalence and severity of CAD. The conflicting data on whether this polymorphism contributes to CAD is likely because multiple long-standing risk factors and atherosclerotic burden confound the possible effect of this polymorphism on a polygenic disease such as atherosclerosis.

Importantly, to our knowledge to date, there has not been a precise assessment of the impact of these polymorphisms on NADPH oxidase function. This is noteworthy, because in the absence of an alteration in NADPH oxidase activity, it is highly unlikely that a polymorphism in p22<sup>Hello</sup> would alter vascular function or predispose to CAD. In view of this, we examined the functional consequences of CYBA polymorphisms using a cell type in which the NADPH oxidase subunit composition is well defined in a population of healthy adults. This approach was chosen to eliminate the contribution of other sources of superoxide present in the vasculature, confounding risk factors, and atherosclerotic burden. Because polymorphisms are shared across tissues and p22<sup>Hello</sup> is common to all known NAD(P)H oxidases, measurement of neutrophil respiratory burst activity permits assessment of the effect of a given polymorphism on enzyme function in multiple tissues. We therefore examined the effect of the C242T, A640G, and −93O<sup>−1G</sup> polymorphisms on neutrophil NADPH oxidase activity.

**Methods**

**Subjects**
The study group consisted of 90 healthy adults who gave informed consent for participation. Coronary risk factors in this study were defined as described previously. Subjects were excluded if there was a history for CAD, if >2 risk factors for CAD were present, if there was clinical evidence for an acute or chronic infection, and if they were older than 45 years. This study was approved by the Emory University Institutional Review Board and the procedures followed institutional guidelines.

**Blood Sampling and Genotyping**
DNA was extracted from venous blood samples, and the DNA fragment containing the C242T and A640G polymorphic sites of CYBA was amplified from genomic DNA by polymerase chain reaction (PCR) as previously described. The −93O<sup>−1G</sup> polymorphism in the promoter of the CYBA gene was determined using the TaqMan (Applied Biosystems) PCR method and the ABI PRISM 7900HT Sequence Detector (Applied Biosystems). Clear identification of each genotype was independently validated with sequencing by Emory DNA sequencing core facility as well as by DNA high-performance liquid chromatography using Transgenic WAVE DNA fragment analysis.

**Neutrophil Isolation**
Soon after venous blood sampling, neutrophils were isolated by Percoll density gradient centrifugation as previously described.

**Superoxide Measurement**
Superoxide production by human neutrophils was measured using electron spin resonance spectroscopy (ESR) with the superoxide dismutase (SOD) inhibitable oxidation of the spin probe CPH (Figure 1), which after reaction with superoxide yields a stable nitroxide radical with a half-life of several hours. Neutrophils (1×10<sup>6</sup>) were incubated with 5 mmol/L CPH and stimulated by the addition of 200 nmol/L of phorbol 12-myristate 13-acetate (PMA). All ESR samples were prepared using chelated phosphate-buffered saline, pH 7.4. ESR spectra were recorded in a 50-μL glass capillary tube at room temperature using a Bruker EMX spectrometer. ESR spectra were obtained using the following instrument settings: field sweep, 60 G; microwave frequency, 9.822 GHz; microwave power, 20 mW; modulation amplitude, 1 G; conversion time, 163.840 ms; time constant 327.68 ms; sweep time, 167.772 s; receiver gain, 1×10<sup>6</sup>. ESR kinetic scans were obtained over 10 minutes in unstimulated neutrophils and after PMA stimulation in the presence or absence of 100 units of SOD (Figure 1). The rate of superoxide formation was measured by monitoring the amplitude of the low-field component of the ESR spectrum as previously described. The concentration of CP-nitroxide was determined from a calibration curve for intensity of the ESR signal at various known concentrations of CP-nitroxide. The rate of superoxide production was calculated from the accumulation of the CP-nitroxide, obtained from the time scan of CPH oxidation (Figure 1). For this purpose, the
**Clinical and Demographic Characteristics of Study Subjects**

<table>
<thead>
<tr>
<th>Characteristic</th>
<th>Study Subjects</th>
</tr>
</thead>
<tbody>
<tr>
<td>Age, y</td>
<td>33 (20–45)</td>
</tr>
<tr>
<td>Gender, n (%)</td>
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</tr>
<tr>
<td>Male</td>
<td>55 (61)</td>
</tr>
<tr>
<td>Female</td>
<td>35 (39)</td>
</tr>
<tr>
<td>Risk factors, n (%)</td>
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</tr>
<tr>
<td>Family history of CAD</td>
<td>18 (20)</td>
</tr>
<tr>
<td>Hypercholesterolemia</td>
<td>2 (2)</td>
</tr>
<tr>
<td>Hypertension</td>
<td>1 (1)</td>
</tr>
<tr>
<td>Smoking</td>
<td>2 (2)</td>
</tr>
<tr>
<td>Ethnicity, n (%)</td>
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<tr>
<td>White</td>
<td>78 (86)</td>
</tr>
<tr>
<td>Black</td>
<td>6 (7)</td>
</tr>
<tr>
<td>Other</td>
<td>6 (7)</td>
</tr>
</tbody>
</table>

Values are mean or n (%).

The corresponding allele prevalences for blacks were 0.17 (0.02, 0.48), 0.58 (0.28, 0.85), and 0.58 (0.28, 0.85). However, these prevalences were based on only 6 persons and hence were imprecise.

### Results

**Western blotting** was performed as previously described.[3] Briefly, equal amount of protein from neutrophils were applied on 12% SDS-polyacrylamide gels. Polyclonal anti-p22^phox^ antibody and horseradish peroxidase-labeled donkey anti-rabbit immunoglobulin (Santa Cruz and Amersham) were used as primary and secondary antibodies. The signals were detected by the enhanced chemiluminescence method and quantified by densitometry.

**Statistical Analysis**

Exact procedures for the binomial distribution were used to obtain 95% confidence intervals for the prevalences of the T and G alleles of the CYBA polymorphisms. The observed distribution of the CYBA genotypes compared with that expected under Hardy-Weinberg equilibrium were evaluated by a 1-degree-of-freedom χ² test.[21] Differences in the means of neutrophil-generated respiratory burst activity according to the genotypes of the 3 CYBA polymorphisms were evaluated by standard ANOVA.[22] Finally, we also used the Wilcoxon rank-sum test, a nonparametric procedure when data were not normally distributed.[23] All statistical analysis was implemented using Stata.[24]

### Relationship Between Superoxide Generation and Genotype

The mean levels of superoxide generation in our entire cohort (n = 72) was 0.08±0.005 μmol/L O$_2^-$/min per 10$^6$ cells in the basal state, and 8.3±0.4 μmol/L O$_2^-$/min/10$^6$ cells after PMA stimulation (n = 90). Neutrophils from subjects with the CC and CT genotypes of the C242T CYBA polymorphism produced significantly more superoxide than those from subjects with the TT genotype (P = 0.038; Figure 2A). After correcting for basal production, the TT genotype neutrophils generated only 32% as much superoxide as CC genotype neutrophils. Superoxide production from unstimulated neutrophils of subjects with the CC, CT, and TT genotypes was not statistically significantly different (P > 0.20).

Neutrophils from subjects with AA, AG, and GG genotypes of the A640G CYBA polymorphism produced 8.1±1 μmol/L O$_2^-$/min per 10$^6$ cells, 7.8±0.7 μmol/L O$_2^-$/min per 10$^6$ cells, and 7.9±0.8 μmol/L O$_2^-$/min per 10$^6$ cells, respectively.

Superoxide production from stimulated neutrophils of subjects with the AA, AG, and GG genotypes were not statistically significantly different (P > 0.20, Figure 2B). There was lower superoxide production in unstimulated neutrophils of subjects with the GG genotype (0.08±0.01 μmol/L O$_2^-$/min per 10$^6$ cells) compared with AG and AA genotypes (0.12±0.01 μmol/L O$_2^-$/min per 10$^6$ cells) and 0.12±0.02 μmol/L O$_2^-$/min per 10$^6$ cells) (P < 0.02).

Neutrophils from subjects with AA, AG, and GG genotypes of the −930^AG^ polymorphism produced 8.3±1.1 μmol/L O$_2^-$/min per 10$^6$ cells, 8.5±0.5 μmol/L O$_2^-$/min per 10$^6$ cells and 8.4±1.1 μmol/L O$_2^-$/min per 10$^6$ cells, respectively. Superoxide production by either stimulated or unstimulated neutrophils of subjects with the AA, AG, and GG genotypes were not statistically significantly different (P > 0.20, Figure 2C).

### Effect of C242T Polymorphism on p22^phox^ Expression

One possible explanation for the observed decrease in NADPH oxidase activity is a decreased expression of p22^phox^.[25] However, p22^phox^ protein levels were equivalent in the CC, CT, and TT genotypes (Figure 3).

### Discussion

Our study demonstrates the effect of 3 CYBA polymorphisms on phagocytic superoxide production. Healthy adults who carry the TT genotype of the C242T CYBA polymorphism have a significant reduction in respiratory burst compared with wild-type carriers, suggesting decreased activity of the NADPH oxidase in these individuals. In contrast, neither the A640G nor the −930^AG^ CYBA polymorphism affects superoxide production.

During the past decade, p22^phox^-based NAD(P)H oxidases were identified as major sources of vascular oxidative stress.[2] Azumi et al demonstrated that p22^phox^ was expressed in human coronary arteries, and its expression was more intense in atherosclerotic than nonatherosclerotic arteries.[5] In athero-
sclerotic lesions, p22phox was present around lipid core and shoulder regions and was localized in T-lymphocytes, endothelial and smooth muscle cells, as well as fibroblasts. More recently, a significant association between superoxide generation, p22phox expression, and oxidized low-density lipoprotein cholesterol was shown. Previous observations suggested that gp91phox is the sole heme binding subunit of the NADPH oxidase; however, functional assembly of the active NADPH oxidase to mediate electron transfer for superoxide generation requires both subunits of flavocytochrome b558. Because the gp91phox, p22phox subunit stoichiometry is 1:1, and because the presence of p22phox is necessary to stabilize the native heme environment of the cytochrome b558, it is conceivable that polymorphisms in the CYBA gene reduce the ability of p22phox to anchor gp91phox and act as a scaffold, and subsequently alter NADPH oxidase activity.

Our observations regarding decreased respiratory burst activity in subjects with the TT genotype confirm a functional role for this polymorphism and support and extend the observations made by Guzik et al. They demonstrated that the 242T allele is associated with reduced vascular NAD(P)H oxidase activity in saphenous veins of CAD patients, independent of other clinical risk factors. In contrast to these observations, in our study population, the C242T CYBA polymorphism does not affect basal activity of the phagocytic enzyme, but leads to ~30% reduced activity on stimulation.

Because vascular tissue contains multiple enzymes with NAD(P)H oxidase activity, it is difficult to make an absolute correlation between p22phox-based oxidase activity and genotype in this system. For this reason we isolated neutrophils that express significant amounts of a well-defined NADPH oxidase of single molecular composition, this system permits us to directly investigate the relationship between CYBA polymorphisms and enzymatic activity. The close agreement between our study and Guzik’s indicates that the correlation found between the C242T CYBA polymorphism, p22phox-based oxidase activity and endothelial function might be a relevant clinical marker for CAD.

There are 2 limitations of studies of this type. First, CAD patients have multiple risk factors and significant atherosclerotic lesions, p22phox was present around lipid core and shoulder regions and was localized in T-lymphocytes, endothelial and smooth muscle cells, as well as fibroblasts. More recently, a significant association between superoxide generation, p22phox expression, and oxidized low-density lipoprotein cholesterol was shown.

We and others showed the critical role of p22phox in NAD(P)H oxidase activity in different cell lines and tissues. The p22phox subunit binds to gp91phox and nox1, and appears to be necessary for mutual stability (unpublished observations). Previous observations suggested that gp91phox is the sole heme binding subunit of the NADPH oxidase; however, functional assembly of the active NADPH oxidase to mediate electron transfer for superoxide generation requires both subunits of flavocytochrome b558. Because the gp91phox, p22phox subunit stoichiometry is 1:1, and because the presence of p22phox is necessary to stabilize the native heme environment of the cytochrome b558, it is conceivable that polymorphisms in the CYBA gene reduce the ability of p22phox to anchor gp91phox and act as a scaffold, and subsequently alter NADPH oxidase activity.

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oxidative burden, making it difficult to investigate the effect of a polymorphism free of other factors known to increase superoxide. This was not an issue here, because our study subjects were young, had no history of CAD, and very few had coronary risk factors. Second, methods used to measure superoxide are subject to many potential problems, such as redox cycling observed with higher doses of lucigenin, signals caused by direct reduction of the substrate, as with cytochrome C, and poor quantification. In this study, we used ESR to measure NADPH oxidase activity, which is extremely specific and not subject to many of the difficulties encountered with other methods. The use of the cyclic hydroxylamine is also an advantage, because it is not subject to reduction to a spin-inactive state by reductants in cells, which is a problem encountered with commonly used nitrore spin traps.

A reduced level of oxidative stress would either prevent or delay the development of or protect the vasculature against oxidant-mediated conditions such as atherosclerosis and endothelial dysfunction by maintaining a higher bioavailability of NO. Furthermore, atherosclerotic plaque rupture in areas of the shoulder region where NADPH oxidase-containing monocytes/macrophages and neutrophils are recruited is known to be promoted by superoxide. In addition, neutrophils from patients with acute myocardial infarction and peripheral obstructive atherosclerotic disease produce more superoxide and cytochrome b light chain (p22phox), gene structure, chromosomal localization and mutations in cytochrome-negative autosomal recessive chronic granulomatous disease. J Clin Invest. 1990;86:1729–1737.


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