A New Transgenic Rat Model of Hepatic Steatosis and the Metabolic Syndrome

Nathan R. Qi, Jiaming Wang, Vaclav Zidek, Vladimir Landa, Petr Mlejnek, Ludmila Kazdová, Michal Pravenec, Theodore W. Kurtz

Abstract—Fatty liver is extremely common in insulin-resistant patients with either obesity or lipodystrophy and it has been proposed that hepatic steatosis be considered an additional feature of the metabolic syndrome. Although insulin resistance can promote fatty liver, excessive hepatic accumulation of fat can also promote insulin resistance and could contribute to the pathogenesis of the metabolic syndrome. We sought to create a new nonobese rat model with hypertension, hepatic steatosis, and the metabolic syndrome by transgenic overexpression of a sterol-regulatory element-binding protein (SREBP-1a) in the spontaneously hypertensive rat (SHR). SREBPs are transcription factors that activate the expression of multiple genes involved in the hepatic synthesis of cholesterol, triglycerides, and fatty acids. The new transgenic strain of SHR overexpressing a dominant-positive form of human SREBP-1a under control of the phosphoenolpyruvate carboxykinase (PEPCK) promoter exhibited marked hepatic steatosis with major biochemical features of the metabolic syndrome, including hyperglycemia, hyperinsulinemia, and hypertriglyceridemia. Both oxidative and nonoxidative skeletal muscle glucose metabolism were significantly impaired in the SHR transgenic strain and glucose tolerance deteriorated as the animals aged. The SHR transgenic strain also exhibited reduced body weight and reduced adipose tissue stores; however, the level of hypertension in the transgenic SHR was similar to that in the nontransgenic SHR control. The transgenic SHR overexpressing SREBP-1a represents a nonobese rat model of fatty liver, disordered glucose and lipid metabolism, and hypertension that may provide new opportunities for studying the pathogenesis and treatment of the metabolic syndrome associated with hepatic steatosis. (Hypertension. 2005; 45:1004-1011.)

Key Words: hypertension ■ insulin resistance ■ liver ■ metabolism ■ rats ■ transcription, genetic

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syndrome and its relationship to nonalcoholic fatty liver disease.

In the current study, we sought to create a nonobese rat model with hypertension, fatty liver, and the metabolic syndrome by transgenic overexpression of a sterol-regulatory element-binding protein (SREBP) in the spontaneously hypertensive rat (SHR). SREBPs are transcription factors involved in the regulation of fatty acid and lipid metabolism and can activate the expression of multiple genes involved in the hepatic synthesis of cholesterol, triglycerides, fatty acids, and phospholipids. Transgenic overexpression of SREBPs is also known to be an effective method for causing hepatic steatosis in rodents. In addition, genetic variants in SREBPs have been reported to be associated with increased risk for obesity, type 2 diabetes, and dyslipidemia. We now report that transgenic overexpression of human SREBP-1a in the SHR induces hepatic steatosis and multiple biochemical features of the metabolic syndrome, including hyperinsulinemia, hyperglycemia, and hypertriglyceridemia, in the absence of obesity. The transgenic SHR expressing human SREBP-1a may be a useful model for studying the relationship between hepatic steatosis and the metabolic syndrome and for investigating the pathogenesis and treatment of these very common and highly related clinical disorders.

Methods

Generation of Transgenic Rats

Transgenic rats overexpressing a dominant-positive form of human SREBP-1a were derived on the genetic background of the spontaneously hypertensive SHR/Ola strain that harbors a mutant form of the CD36 fatty acid transporter. The SHR/Ola strain descends directly from the widely used SHR/NIH strain and was one of the original strains used to map the metabolic phenotypes linked to CD36 deficiency. Transgenic SHR were produced by microinjection of zygotes with a cDNA construct encoding amino acids 1 to 460 of human SREBP-1a under control of the rat PEPCK promoter, which produces high levels of gene expression in the liver. These primers generated a truncated form of SREBP-1a that can enter the nucleus without proteolysis and is not subject to feedback regulation. The attendant increase in nuclear SREBP-1a stimulates the hepatic expression of multiple genes involved in cholesterol and triglyceride synthesis, thereby resulting in a fatty liver.

Transgenic SHR were detected by polymerase chain reaction (PCR) using primers specific for the human transgene: upstream, 5’-cag ccc tgt gtt agg cta c-3’; downstream, 5’-ttc atg gtc aga aga aga-3’. These primers generated a 29-basepair amplicon in transgene-positive rats but not in SHR transgene-negative controls. The presence of the transgene was further confirmed using a PCR primer set that co-amplified the human and rat genes encoding SREBP-1a. These primers produced a single set of primers that co-amplified the human and rat genes encoding SREBP-1a but that generated different sizes of amplicons for the 2 genes. This primer pair bridged an intron in the endogenous rat gene that was absent in the cDNA transgene, thereby generating a larger amplicon from the endogenous rat gene (~700 basepairs) and a smaller amplicon (484 basepairs) from the human transgene. The upstream primer for this PCR testing was 5’-cct ccc tgt cct att tgc-3’ and the downstream primer was 5’-ctc gga cct ggg tgt g-3’.

The first transgenic line obtained exhibited a fatty liver and abundant hepatic expression of the SREBP-1a transgene. This line was used in all studies and was compared with nontransgenic SHR controls (transgene-negative littermates or transgene-negative rats from the same SHR colony used to derive the transgenic strain). The rats were housed in an air-conditioned animal facility and allowed free access to food and water. All protocols were performed in agreement with the Animal Protection Law of the Czech Republic and were approved by the Ethics Committee of the Institute of Physiology, Czech Academy of Sciences, Prague or the Institutional Animal Care and Use Committee of the University of California, San Francisco.

Phenotyping Protocols

Oral glucose tolerance testing was performed in young (age 10 weeks) and old (age 16 months) male transgenic SHR and male SHR controls after 2 weeks on a high-fructose diet (60% fructose) (K4102.0 diet; Hope Farms, the Netherlands). The oral glucose tolerance testing protocol was performed as previously described. A high-fructose diet was used because previous studies have shown that high-fructose diets can accentuate features of the metabolic syndrome in genetically susceptible strains of SHR. Two days after the oral glucose tolerance testing, serum levels of insulin, fatty acid, and triglyceride levels were obtained in the nonfasting state. In the 10-week-old rats, tissue samples were also obtained at the end of the study for measurements of soleus muscle glucose oxidation, soleus muscle glucose intake into glycogen, triglyceride levels in liver and skeletal muscle, and organ weights.

In a separate experiment, radiotelemetry measurements of arterial pressure were obtained from 17- to 21-week-old male transgenic SHR (n=9) and transgene-negative controls (n=9) fed normal rat chow. Subsets of these animals were euthanized in the nonfasting state at 22 to 25 weeks of age and serum samples obtained for measurements of leptin and adiponectin levels and tissue samples obtained for measurements of adipocyte size in epididymal fat according to the method of Di Girolamo et al. Liver samples were also obtained for histology in these rats and in a group of older (16 months) rats.

Expression Analysis of Transgenic SHR Rats

Real-time PCR analysis was used to detect tissue patterns of expression of the human SREBP-1a transgene and to quantify expression levels in fat and liver of several target genes of SREBP-1a that encode key enzymes involved in lipid metabolism, namely fatty acid synthase (FAS), stearoyl coenzyme A desaturase 1 (SCD-1), and acetyl coenzyme A carboxylase 1. The gene expression levels were assessed in 22- to 25-week-old rats that had been fed a high-fructose diet for 2 weeks. For real-time PCR, total RNA was isolated using standard methods and cDNA was prepared and analyzed by real-time PCR testing using Quantifect SYBR Green reagents (Qiagen, Inc., Valencia, Calif.) on an Option continuous fluorescence detector (MJ Research, Waltham, Mass) as previously described. Gene expression levels were normalized relative to the expression of cyclophilin (peptidylprolyl isomerase A), which served as the internal control, with results being determined in triplicate using the preferred method of Muller et al. For SREBP-1a target genes, the results in the transgene-positive rats (n=3) are displayed in comparison to the results in the transgene-negative rats (n=3). For the expression level of the human transgene encoding SREBP-1a, the results are displayed compared with the expression level of the endogenous rat gene encoding SREBP-1a. Expression of the transgene encoding human SREBP-1a was quantified using the following primers: upstream, 5’-tgc tga cca atc tga aag a; downstream, 5’-agg tgt aga cca gga ggt cag gc; and ACC upstream, 5’-tac atc aag tgg gac cac ag; FAS downstream, 5’-tgt tcg tca gca cct tct tg; SCD-1 upstream, 5’-ctc gga tgt tct ccc gag att ga; downstream, 5’-ctc ccc tgt gtt agg cta c-3’. The primers used to quantify expression of the SREBP-1a target genes were: FAS upstream, 5’-cag ctc atc aag tgt gag cac ag; FAS downstream, 5’-agg cta ccc ttg tgt gag g; downstream, 5’-cct cta ggc cag cag gc; and ACC upstream, 5’-acc itt ctc ctc tct ctc tct ctc tct cca gc.

Skeletal Muscle Glycogen Synthesis and Glucose Oxidation

Glycogen synthesis and glucose oxidation were determined in isolated soleus muscle by measuring the incorporation of [U-14C]U
glucose into glycogen and CO₂ as previously described.²¹,²⁵ The soleus muscles were attached to a stainless steel frame in situ at in vivo length by special clips and separated from other muscles and tendons and immediately incubated for 2 hours in Krebs-Ringer bicarbonate buffer, pH 7.4, that contained 5.5 mmol/L unlabeled glucose, 0.5 μCi/mL of 14C-U glucose, and 3 mg/mL bovine serum albumin (Armour, Fraction V) with or without 250 μU/mL insulin. After 2-hour incubation, 0.3 mL of 1 mol/L hyamine hydroxide was injected into the central compartment of the incubation vessel and 0.5 mL of 1 mol/L H₂SO₄ into the main compartment to liberate CO₂. The vessels were incubated for another 30 minutes, and the hyamine hydroxide was then quantitatively transferred into the scintillation vial containing 10 mL of toluene-based scintillation fluid for counting of radioactivity. Measurement of insulin-stimulated incorporation of glucose into glycogen was determined after extraction of glycogen as previously described.²¹,²⁵

Tissue Triglyceride Measurements
For determination of triglycerides in liver and soleus muscle, tissues were powdered under liquid N₂ and extracted for 16 hours in chloroform:methanol, after which 2% KH₂PO₄ was added, and the solution was centrifuged. The organic phase was removed and evaporated under N₂. The resulting pellet was dissolved in isopropyl alcohol, and triglyceride content was determined by enzymatic assay (Pliva-Lachema, Brno, Czech Republic).

Hemodynamic Studies
Arterial blood pressures were measured continuously in unanesthetized, unrestrained rats using radiotelemetry. All rats were allowed to recover for at least 10 to 14 days after surgical implantation of radiotelemetry transducers before the start of blood pressure recordings. Pulsatile pressures were recorded in 5-second bursts every 5 minutes throughout the day and 24-hour averages for systolic arterial blood pressure were calculated for each rat on each day of the study. The results from each rat in the same group were then averaged to obtain the group means. Blood pressure salt-sensitivity was also assessed by measuring arterial pressures during a 1-week period in which 1% NaCl solution was provided in place of drinking water.

Biochemical Analyses
Blood glucose levels were measured by the glucose oxidase assay (Pliva-Lachema) using tail vein blood drawn into 5% trichloracetic acid and promptly centrifuged. Serum nonesterified fatty acid levels were determined using an acyl-coenzyme A oxidase-based colorimetric kit (Roche Diagnostics GmbH, Mannheim, Germany). Serum triglyceride concentrations were measured by standard enzymatic methods (Pliva-Lachema). Serum insulin concentrations were determined using a rat insulin radioimmunoassay kit (Amersham Pharmacia Biotech UK Ltd). Serum levels of leptin and adiponectin were determined using radioimmunoassay kits from Linco Research (St. Charles, Mo).

Statistical Analysis
All data are expressed as means±SEM. Arterial pressures were analyzed by repeated measures analysis of variance (ANOVA). Individual group means for metabolic measurements were compared by t test. Statistical significance was defined as P<0.05.

Results
Expression of the Human Transgene Gene Encoding SREBP-1a
Transgene-positive SHR and transgene-negative controls were identified by genotyping offspring derived from crosses of a founder male with transgene-negative SHR females. Real-time PCR analysis confirmed abundant hepatic expression of the human transgene encoding SREBP-1a compared with expression of the endogenous SREBP-1a gene (Figure 1). Lower levels of transgene expression could also be
Table 1. Metabolic Features of 10-Week-Old Transgenic SHR Expressing Human SREBP-1a

<table>
<thead>
<tr>
<th>Parameter</th>
<th>SHR Controls (n=8)</th>
<th>SHR Transgenics (n=9)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Serum glucose, mmol/L</td>
<td>5.2±0.1</td>
<td>5.7±0.1†</td>
</tr>
<tr>
<td>Serum insulin, mmol/L</td>
<td>1.1±0.1</td>
<td>1.8±0.1†</td>
</tr>
<tr>
<td>Serum triglycerides, mmol/L</td>
<td>1.9±0.3</td>
<td>6.5±0.5‡</td>
</tr>
<tr>
<td>Serum fatty acids, mmol/L</td>
<td>0.4±0.02</td>
<td>0.7±0.04‡</td>
</tr>
<tr>
<td>Serum cholesterol, mmol/L</td>
<td>1.8±0.1</td>
<td>2.4±0.1†</td>
</tr>
<tr>
<td>Glucose tolerance AUC, mmol/L</td>
<td>813±13</td>
<td>797±7</td>
</tr>
<tr>
<td>Liver triglycerides, μmol/g</td>
<td>13.1±0.6</td>
<td>32.6±3.5‡</td>
</tr>
<tr>
<td>Liver cholesterol, μmol/g</td>
<td>6.9±0.2</td>
<td>10.8±0.6‡</td>
</tr>
<tr>
<td>Spleen triglycerides, μmol/g</td>
<td>4.9±0.5</td>
<td>9.2±1.2†</td>
</tr>
<tr>
<td>Body weight, g</td>
<td>253±3</td>
<td>235±8*</td>
</tr>
<tr>
<td>Liver weight, g</td>
<td>11.1±0.2</td>
<td>13.3±0.4‡</td>
</tr>
<tr>
<td>Epididymal fat pad weight, g</td>
<td>2.1±0.1</td>
<td>1.5±0.1†</td>
</tr>
<tr>
<td>Perirenal fat pad weight, g</td>
<td>1.4±0.2</td>
<td>0.6±0.1†</td>
</tr>
</tbody>
</table>

Measurements obtained after feeding rats a high-fructose diet for 2 weeks. Serum glucose and glucose tolerance testing area under the curve (AUC) obtained in the fasting state; other measurements obtained in the nonfasting state.

*P<0.05, †P<0.01, ‡P<0.001 compared to control.

Metabolic Effects of Transgenic Expression of SREBP-1a in Young Transgenic Rats

In 10-week-old transgenic SHR fed a high-fructose diet for 2 weeks, serum concentrations of glucose, insulin, triglycerides, and fatty acids were all significantly increased compared with the SHR transgene-negative controls (Table 1). However, there was no significant difference in the area under the curve of the oral glucose tolerance test between the 10-week-old SHR transgenics and the age-matched SHR controls (Table 1 and Figure 2).

The transgenic SHR weighed ≈10% less than the transgene-negative controls (Table 1). Epididymal and perirenal fat pad weights were also significantly reduced (P<0.01) in the transgenic SHR compared with the transgene-negative controls (Table 1). However, liver weight was significantly increased in the transgenic strain (Table 1). In addition, the transgenic SHR exhibited increased levels of triglycerides in liver and in soleus muscle compared with the SHR transgene-negative controls (Table 1). Hepatic levels of cholesterol were also significantly increased in the transgenic SHR compared with controls (Table 1). The changes in serum and tissue lipid levels induced by transgenic expression of human SREBP-1a in the SHR model are similar to those reported to be induced by expression of the same human SREBP-1a transgene in low-density lipoprotein receptor knockout mice.28

In soleus muscle isolated from 10-week-old transgenic SHR expressing human SREBP-1a, glucose oxidation and glucose incorporation into glycogen were significantly decreased compared with SHR transgene-negative controls either in the presence or in the absence of insulin (Table 2). Thus, basal and insulin-stimulated oxidative and nonoxidative glucose metabolism were impaired in skeletal muscle of transgenic rats compared with controls.

Metabolic Effects of Transgenic Expression of SREBP-1a in Old Transgenic Rats

To determine if glucose tolerance deteriorated with age, we performed oral glucose tolerance testing in 16-month-old transgenic SHR and in age-matched SHR controls after feeding a high-fructose diet for 2 weeks. In contrast to the results in the young rats, glucose tolerance was significantly impaired in the old transgenic SHR compared with age-

Figure 2. Oral glucose tolerance tests (OGTT) in young and old SHR transgenic rats and SHR controls. Left panel, OGTT in 10-week-old SHR transgenic rats (solid circles, n=9) and SHR controls (open symbols, n=8) fed a high-fructose diet for 2 weeks before study. Right panel, OGTT in 16-month-old SHR transgenic rats (solid circles, n=12) and SHR controls (open symbols, n=9) fed a high-fructose diet for 2 weeks before study. NS indicates no significant difference between the 2 groups in the area under the curve of the glucose tolerance tests. *P<0.01, significant difference between the 2 groups in the area under the curve of the glucose tolerance tests.
matched controls (Figure 2 and Table 3). Serum glucose, insulin, and lipid levels were also significantly increased in the 16-month-old transgenic SHR compared with the SHR transgene-negative controls (Table 3).

**Effects on Blood Pressure**

Despite significantly lower body weights throughout the course of the studies (Figure 3; Table 1), the transgenic SHR exhibited frank hypertension with blood pressures similar to those in transgene negative SHR controls (Figure 3). The blood pressure response to salt loading in the transgenic SHR was also similar to that in the SHR controls (Figure 3).

**Effects on Liver and Fat Pad Morphology and Adipocytokine Levels**

Autopsy of 24-week-old rats maintained on normal chow confirmed the presence of grossly enlarged fatty livers and reduced adipose tissue stores in the transgenic strain compared with the transgene negative control (Figure 4). The mean liver weight of the 24-week-old transgenic rats, 18.2 ± 0.9 grams, was significantly greater than that of controls, 12.8 ± 0.3 grams (P < 0.001). In the transgenic strain, histological analysis confirmed the presence of fatty liver as expected; however, there was no apparent increase in the amount of hepatic inflammation compared with the SHR control strain. However, distinct features of steatohepatitis were evident on histological analysis of livers from 16-month-old transgenic rats (data not shown).

**TABLE 2. Glucose Metabolism in Isolated Soleus Muscle from 10-Week-Old Transgenic SHR**

<table>
<thead>
<tr>
<th>Parameter</th>
<th>SHR Controls (n=8)</th>
<th>SHR Transgenics (n=9)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Soleus muscle</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Glucose oxidation, nmol glucose/g per 2 hours</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Without insulin</td>
<td>167 ± 13</td>
<td>93 ± 8‡</td>
</tr>
<tr>
<td>With insulin</td>
<td>268 ± 37</td>
<td>129 ± 14‡</td>
</tr>
<tr>
<td>Glycogenesis, nmol glucose/g per 2 hours</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Without insulin</td>
<td>925 ± 75</td>
<td>388 ± 82‡</td>
</tr>
<tr>
<td>With insulin</td>
<td>1915 ± 269</td>
<td>765 ± 162†</td>
</tr>
</tbody>
</table>

Measurements obtained after feeding rats a high-fructose diet for 2 weeks. *P < 0.05; †P < 0.01; ‡P < 0.001 compared with control.

Epididymal and perirenal fat weights in the transgenic strain, 0.9 ± 0.1 grams and 0.6 ± 0.04 grams, respectively, were significantly lower than in the transgene-negative control, 3.1 ± 0.3 grams and 2.8 ± 0.2 grams, respectively (both P < 0.0001). Although adipose tissue mass was decreased in the transgenic strain, the average volume of adipocytes in the epididymal fat pads was significantly increased in the transgenic rats compared with controls, 405 ± 18 picoliters versus 327 ± 28 picoliters, respectively (P < 0.05). The transgenic rats showed an increase in the percentage of large adipocytes with diameters > 120 μm compared with SHR controls: 10% of adipocytes in the transgenics were > 120 μm, whereas only 1% of the adipocytes were > 120 μm in the SHR controls. Serum levels of leptin and adiponectin in the SHR transgenic strain, 1.9 ± 0.3 and 2.2 ± 0.3 ng/mL, respectively, were significantly lower than in the SHR control strain, 4.2 ± 0.4 and 5.4 ± 0.5 ng/mL, respectively (both P < 0.005).

**Discussion**

In the current study in the most widely used rat model of essential hypertension, we have found that transgenic overexpression of a dominant-positive form of human SREBP-1a under control of the PEPCK promoter induces marked hepatic steatosis and biochemical features of the metabolic syndrome. Transgenic SHR exhibited fatty liver and hyperglycemia, hyperinsulinemia, and hypertriglyceridemia compared with nontransgenic SHR. Oral glucose tolerance was similar in 10-week-old transgenic SHR versus SHR controls but became frankly abnormal in older transgenic rats. In addition to glucose intolerance developing, histological fea-

**TABLE 3. Metabolic Features of 16 Month Old Transgenic SHR Expressing Human SREBP-1a**

<table>
<thead>
<tr>
<th>Parameter</th>
<th>SHR Controls (n=9)</th>
<th>SHR Transgenics (n=12)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Serum glucose (mmol/L)</td>
<td>5.3 ± 0.2</td>
<td>6.1 ± 0.2†</td>
</tr>
<tr>
<td>Serum insulin (mmol/L)</td>
<td>0.2 ± 0.03</td>
<td>0.9 ± 0.1‡</td>
</tr>
<tr>
<td>Serum triglycerides (mmol/L)</td>
<td>0.5 ± 0.03</td>
<td>5.6 ± 1.0‡</td>
</tr>
<tr>
<td>Serum fatty acids (mmol/L)</td>
<td>0.5 ± 0.01</td>
<td>0.8 ± 0.04‡</td>
</tr>
<tr>
<td>Serum cholesterol (mmol/L)</td>
<td>1.9 ± 0.1</td>
<td>2.5 ± 0.2†</td>
</tr>
<tr>
<td>Glucose tolerance AUC (mmol/L/2 hours)</td>
<td>792 ± 16</td>
<td>944 ± 35†</td>
</tr>
</tbody>
</table>

Measurements obtained after feeding rats a high fructose diet for two weeks. Serum glucose and glucose tolerance testing AUC (area under the curve) obtained in the fasting state; other measurements obtained in the non-fasting state. *P < 0.05; †P < 0.01; ‡P < 0.001 compared to control.
atures of steatohepatitis developed in the older transgenic SHR. Progressive impairment in glucose metabolism is known to occur in humans with the metabolic syndrome and some studies suggest that insulin resistance further deteriorates as patients progress from fatty liver to nonalcoholic steatohepatitis.29 The transgenic SHR were also characterized by impaired oxidative and nonoxidative glucose metabolism in skeletal muscle. However, the level of hypertension in the transgenic SHR was similar to that in the nontransgenic SHR despite lower body weights, reduced amounts of adipose tissue, and multiple disturbances in glucose and lipid metabolism. These findings suggest that the SHR transgenic strain represents a lipodystrophic form of the metabolic syndrome.

In both lean and obese individuals, ectopic deposition of fat in liver and muscle is widely suspected to play a role in the pathogenesis of the metabolic syndrome and diabetes by impairing hepatic and skeletal muscle glucose metabolism.30–32 Ectopic accumulation of fat has also been proposed to explain why patients with peripheral lipodystrophy are at increased risk for insulin resistance and diabetes despite having decreased total body fat.30,33,34 In the current study, the transgenic SHR overexpressing SREBP-1a exhibited hyperglycemia and hyperinsulinemia associated with increases in skeletal muscle and liver fat, decreases in adipose tissue stores, and decreases in body weight. These findings, together with evidence of impaired oxidative and nonoxidative glucose metabolism in isolated soleus muscle of the transgenic SHR, suggest that the metabolic disturbances in this strain are at least partly related to disordered peripheral glucose metabolism secondary to the increased deposition of fat in skeletal muscle.

Although we did not assess hepatic glucose metabolism, it is likely that the marked hepatic steatosis is also impairing glucose metabolism in the liver. Studies across multiple animal models with hepatic steatosis have shown an inverse relationship between hepatic triglyceride content and insulin-mediated inhibition of hepatic glucose production.13 Moreover, Samuel et al have found that in rats fed a high-fat diet, hepatic steatosis and hepatic resistance to insulin-mediated suppression of endogenous glucose production precede the peripheral accumulation of fat in skeletal muscle and the development of muscle insulin resistance.13 The decreased leptin and adiponectin levels observed in the transgenic SHR could also be contributing to their metabolic disturbances, because deficiencies in these adipocytokines are known to promote impaired glucose metabolism in liver and skeletal muscle.35–38 Thus, in future studies in the transgenic SHR, it will be of interest to simultaneously determine the time course for development of fat accumulation and insulin resistance in liver and skeletal muscle and to determine whether supplemental administration of leptin or adiponectin can ameliorate the hepatic steatosis and metabolic status of the transgenic SHR.

In contrast to the results in transgenic SHR expressing SREBP-1a, transgenic mice that express the same SREBP-1a construct used in this study do not display typical biochemical features of the metabolic syndrome despite marked hepatic steatosis and significantly reduced amounts of epididymal fat.20,39 However, severe hypertriglyceridemia and hypercholesterolemia develop when SREBP-1a is transgenically expressed on the genetic background of LDL low-density lipoprotein receptor knockout mice.28 Thus, when coupled with variants in other genes that influence lipid metabolism, changes in SREBP activity can have profound effects on circulating lipid levels. In the current study, we transgenically expressed SREBP-1a on the genetic background of the SHR that is known to harbor a spontaneous genetic defect in the CD36 fatty acid transporter.22,40,41 In previous studies, it has been shown that defective CD36 promotes increased susceptibility to insulin resistance and dyslipidemia in the SHR.22,40,41 These observations raise the possibility that the metabolic abnormalities in transgenic SHR expressing SREBP-1a may depend in part on the interaction between impaired fatty acid transport caused by mutant CD36 and disordered hepatic lipid metabolism caused by unregulated production of SREBP-1a. In future studies, it
should be possible to test the potential role of defective CD36 in modulating the metabolic effects of transgenic SREBP-1a by expressing the SREBP-1a transgene on the genetic background of SHR with wild-type CD36. Studies in additional transgenic lines will also be helpful to insure that the metabolic disturbances observed in our model are secondary to overexpression of SREBP-1a and not simply caused by an insertional mutagenesis event caused by the transgene.

Increased blood pressure is an important cardiovascular risk factor that frequently occurs as part of the metabolic syndrome. In the current study, the severity of hypertension in transgenic SHR was similar to that in SHR controls despite lower body weights and reduced adipose tissue stores in the transgenic strain. Given that reductions in body weight often lead to reductions in blood pressure, this observation raises the possibility that the metabolic abnormalities (eg, reduced adiponectin levels) and/or hepatic steatosis might be serving to maintain some portion of the hypertension in the transgenic strain. However, it is also possible that the hypertension in this model of the metabolic syndrome is entirely accounted for by the increased blood pressure characteristic of the SHR progenitor strain. In future studies, it should be possible to test the hypertensinogenic effects of overexpressing SREBP-1a by transferring the SREBP-1a transgene onto the genetic background of a normotensive strain.

Paterson et al have reported that selective overexpression of 11β-hydroxysteroid dehydrogenase type 1 (11β-HSD1) in liver of normotensive mice can produce hepatic steatosis, insulin resistance, dyslipidemia, and hypertension in the absence of obesity. In the SHR transgenic rats overexpressing SREBP-1a, the hepatic expression of 11β-HSD1 mRNA was similar to that in SHR controls (unpublished observations). Thus, it does not appear that the cardiovascular or metabolic effects of transgenic SREBP-1a are likely related to changes in expression of the gene encoding 11β-HSD1. Nevertheless, the findings of Paterson et al support the possibility that primary alterations in liver function may have the capacity to elicit hemodynamic as well as biochemical disturbances characteristic of the metabolic syndrome.

Perspectives
Transgenic SHR overexpressing SREBP-1a could provide valuable opportunities for investigating pathogenetic mechanisms that may relate fatty liver disease to the metabolic syndrome. In addition, because this model has been created on the background of a strain with increased blood pressure and an inherited defect in fatty acid transport, it could aid in studying the interaction between hepatic steatosis and genetic risk factors for insulin resistance, dyslipidemia, and hypertension. Finally, the availability of a rat model with hepatic steatosis and disordered glucose and lipid metabolism may also be helpful for investigating new therapies for fatty liver disease and the metabolic syndrome.

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