Addition of Angiotensin II Type 1 Receptor Blocker to CCR2 Antagonist Markedly Attenuates Crescentic Glomerulonephritis

Maki Urushihara, Naro Ohashi, Kayoko Miyata, Ryousuke Satou, Omar W. Acres, Hiroyuki Kobori

Abstract—The monocyte chemoattractant protein-1 (MCP-1)/CC-chemokine receptor 2 (CCR2) pathway plays a critical role in the development of antiglomerular basement membrane (anti-GBM) nephritis. We recently showed angiotensin II (Ang II) infusion in rats activated MCP-1 and transforming growth factor-β1 (TGF-β1), which in turn induced macrophage infiltration of renal tissues. This study was performed to demonstrate that combination therapy with a CCR2 antagonist (CA) and an Ang II type 1 receptor blocker (ARB) ameliorated renal injury in the anti-GBM nephritis model. An anti-GBM nephritis rat model developed progressive proteinuria and glomerular crescent formation, accompanied by increased macrophage infiltration and glomerular expression of MCP-1, angiotensinogen, Ang II, and TGF-β1. Treatment with CA alone or ARB alone moderately ameliorated kidney injury; however, the combination treatment with CA and ARB dramatically prevented proteinuria and markedly reduced glomerular crescent formation. The combination treatment also suppressed the induction of macrophage infiltration, MCP-1, angiotensinogen, Ang II, and TGF-β1 and reversed the fibrotic change in the glomeruli. Next, primary cultured glomerular mesangial cells (MCs) stimulated by Ang II showed significant increases in MCP-1 and TGF-β1 expression. Furthermore, cocultured model consisting of MCs, parietal epithelial cells, and macrophages showed an increase in Ang II-induced cell proliferation and collagen secretion. ARB treatment attenuated these augmentations. These data suggest that Ang II enhances glomerular crescent formation of anti-GBM nephritis. Moreover, our results demonstrate that inhibition of the MCP-1/CCR2 pathway with a combination of ARB effectively reduces renal injury in anti-GBM nephritis. (Hypertension. 2011;57[part 2]:586-593.)  ● Online Data Supplement

Key Words: renin-angiotensin system  ■ crescentic glomerulonephritis  ■ MCP-1  ■ CCR2 antagonist  ■ TGF-β1

Crescentic glomerulonephritis (GN), also known as antiglomerular basement membrane (anti-GBM) disease or Goodpasture’s syndrome, is characterized by the formation and deposition of antibodies on the basement membranes of glomeruli and alveoli.1 The disease progresses rapidly, and patients present with renal failure, dyspnea, hemoptysis, a sudden decrease in the hemoglobin level, pallor, and circulatory disturbances. Most patients with advanced disease do not respond to plasmapheresis or immunosuppression therapy.2 While kidney transplantation is an option, because of the risk of recurrence, a patient should wait for 6 months or after the disappearance of serum anti-GBM antibodies before undergoing kidney transplantation.1 Therefore, a novel therapeutic strategy is needed. Studies based on anti-GBM antibody have focused on elucidating the molecular and cellular mechanisms involved in the pathogenesis of this disease. Understanding the mechanisms of proinflammatory responses help facilitate the identification of therapeutic targets for arresting the progression of anti-GBM disease. In Wistar-Kyoto (WKY) rats, the administration of a minute dose of anti-GBM antibodies induces severe proliferative and necrotizing GN with crescent formation.3 In rat models of anti-GBM disease, glomerular infiltration by T lymphocytes, monocytes/macrophages, and a few neutrophils is the earliest and the most prominent pathological change.3 Recent studies have revealed that monocyte chemoattractant protein-1 (MCP-1) is involved in the pathogenesis of crescentic GN.4 Various methods for blocking the actions of proinflammatory cytokines and chemokines have been evaluated in animal models3,5,6 However, the current knowledge regarding effective therapies for anti-GBM disease and the mechanism underlying its pathogenesis is still limited.

Recently, we showed that chronic angiotensin II (Ang II) infusion in rats activated MCP-1 and transforming growth factor-β1 (TGF-β1), which in turn induced macrophage infiltration in renal tissues.7 Furthermore, we reported that TGF-β1 is associated with crescent formation in GN.8 MCP-1 plays a pivotal role in crescentic GN. The release of MCP-1 and TGF-β1 is mediated by the renin-angiotensin system (RAS), and these molecules are considered to be key targets in the treatment of anti-GBM disease. In this study, we hypothesize that the therapeutic management of anti-GBM disease cognition by the therapeutic intervention of monocyte chemoattractant protein-1 (MCP-1)/CC-chemokine receptor 2 (CCR2) pathway.
disease could focus on blocking the MCP-1/CC chemokine receptor 2 (CCR2) signaling pathway and RAS. Our central hypothesis is that the interaction between the MCP-1/CCR2 pathway and the RAS is important for the development of anti-GBM disease.

**Methods**

**Animal Preparation (In Vivo Study)**
The Institutional Animal Care and Use Committee of the Tulane University Health Sciences Center approved all procedures and protocols used in this study. To investigate whether the blockade of MCP-1 and RAS attenuates anti-GBM disease, we treated anti-GBM disease-affected rats with a CCR2 antagonist (CA) and an Ang II type 1 (AT1) receptor blocker (ARB). Progressive anti-GBM GN was induced in 7-week–old male WKY rats by a single intraperitoneal injection of 100 µg of rat monoclonal GBM antibody. RS102895 (CA, 10 mg/kg per day) and/or olmesartan (ARB, 10 mg/kg per day) were mandatory injected into their mouths from day 1. All rats were killed under anesthesia (pentobarbitone) at 2 weeks after the injection of anti-GBM antibodies. RS102895 (Tocris Bioscience) is a novel member of specific CA, and this compound has been shown to inhibit MCP-1/CCR2 signaling in vivo in rodents. The dose of olmesartan that was used in this experiment was high enough to inhibit Ang II receptor binding to kidney tissue. Control rats were nondiseased rats without anti-GBM antibodies or any drug treatment. The urine was obtained in a 24-hour collection using metabolic cages. The amount of protein excreted into the urine was measured by the pyrogallol red method. Blood samples were obtained from all rats at the time of sacrifice. Urinary concentrations of angiotensinogen (AGT) were measured with a commercially available ELISA kit (IBL). Serum creatinine levels were measured by quantitative colorimetric determination (BioAssay Systems). Systolic blood pressure was measured in conscious rats using tail-cuff plethysmography (Visitech) as described previously. All rats were euthanized at 2 weeks after the injection of anti-GBM antibodies, and kidneys were immediately harvested for protein or RNA extraction or for histological analysis.

**Cell Preparation (In Vitro Study)**
Rat cultured mesangial cells (MCs) were established from intact glomeruli as described previously. The normal alveolar macrophages NR8383 were purchased from American Type Culture Collection (ATCC). Parietal epithelial cells (PECs) and macrophages were placed into the lower compartment of Transwell cluster plates (Costa Corning) separated by the chamber with a 0.4-µm polyester membrane filter. MCs were added to the upper chamber and incubated at 37°C in a humidified 5% CO2 incubator. Supernatants were collected from MCs, and the MCP-1 or TGF-β1 concentration was measured with a commercially available ELISA kit. A 4-[3-[4-(iodophenyl)-2-(4-nitrophenyl)-2H-5-tetrazolio]-1,3-benzene disulfonate (WST-1) assay was performed to determine the cell proliferation ability, and the Sircol collagen assay kit was used to measure the collagen concentration from cultured cells.

**Kidney Histology and Immunohistochemistry, Quantitative Real-Time RT-PCR, Western Blot Analysis, Rat PEC Culture, Cytokine Determination by ELISA, and Cell Proliferation Assay and Collagen Measurement**
See http://hyper.ahajournals.org for the detailed Methods.

**Statistical Analysis**
Statistical analysis was performed using a one-way factorial ANOVA with the post hoc Scheffe’s F test. All data are presented as means ±SEM, and probability values <0.05 were considered significant.

**Results**

**Effects of Treatment With the CA and/or the ARB on Systolic Blood Pressure, Urinary Protein Excretion, and Plasma Creatinine Levels**
In contrast to the non-GN rats, systolic blood pressure of the vehicle-treated GN rats was elevated. Treatment with the CA alone showed slightly lower systolic blood pressure than the vehicle-treated GN rats. In comparison, the systolic blood pressure was significantly reduced in the groups treated with the ARB alone or with the combination of CA plus ARB (Figure 1A). As shown in Figure 1B, the vehicle-treated GN rats developed time-dependent, progressive proteinuria, with the urinary protein excretion level increased almost 20-fold at day 14. CA or ARB treatment moderately ameliorated the development of proteinuria. However, CA plus ARB treatment appeared to be attenuated better than other treatments. Consistently, plasma creatinine levels at day 14 were elevated in vehicle-treated rats, which were significantly reduced by CA plus ARB treatment (supplemental Figure S1A). Body weight, daily total water intake, and urine volume were comparable among all groups (supplemental Figure S1B, S1C, and S1D).

**Histological Study**
We examined the effects of treatment with the CA and/or the ARB on renal histology by using periodic acid-Schiff-stained sections (Figure 1C). The most prominent change in the vehicle-treated GN rats was severe glomerular crescent formation. The degree of crescent formation was lower in the CA- or the ARB-treated rats than in rats treated with the vehicle (Figure 1D). Moreover, the combination of the CA and the ARB significantly inhibit crescent formation and GN. The crescentic score was also highest in vehicle-treated GN rats, and the combination therapy with a CA and an ARB showed the significant reduction of the score (Figure 1E).

Furthermore, we quantified the fibrotic changes in glomeruli (Figure 2). Compared with the non-GN rats, vehicle-treated GN rats showed elevation of the percentage of fibrocellular crescent in counted glomerular crescent (Figure 2B) and glomerular fibrotic area (Figure 2C) as well as collagen type I expression (Figure 2A, 2D, and 2E). The inductions of these factors were suppressed by CA and ARB treatment, consistent with the improved level of urinary protein in cotreated rats. Therefore, combination therapy with a CA and an ARB markedly prevents the development of glomerular fibrosis during the course of crescentic GN.

**Glomerular Infiltration by Macrophage and Expression of MCP-1**
The number of CD68-positive macrophages in nphritic glomeruli was markedly increased in vehicle-treated GN rats (Figure 3A and 3B) compared with the non-GN rats. Treatment with CA significantly inhibited the macrophage accumulation in glomeruli. Moreover, CA plus ARB treatment dramatically reduced the macrophage infiltration into glomeruli. Consistently, glomerular expression of MCP-1 was induced in vehicle-treated GN rats (Figure 3A, 3C, 3D, and 3E), as determined by quantitative polymerase chain reaction (PCR) (Figure 3C), immunostaining (Figure 3A and 3D), and
Western blotting (Figure 3E). Treatment with the CA and ARB combination prevented the induction of MCP-1 at the both the mRNA and protein levels in the GN rats.

RAS Activation in Crescentic GN
To determine whether CA plus ARB treatment more effectively suppressed the activation of RAS, Ang II expression levels in glomeruli were examined. Immunostaining showed marked Ang II accumulation in nephritic glomeruli (Figure 4A and 4C). While CA or ARB treatment moderately reduced the increase of Ang II expression level in glomeruli, CA plus ARB treatment further prevented the Ang II accumulation. Consistently, CA plus ARB treatment drastically suppressed AGT expression in glomeruli, whereas CA or ARB treatment mildly attenuated the AGT increase in glomeruli (Figure 4A and 4B). These observations were confirmed by urinary AGT levels (Figure 4E). Furthermore, AT1 receptor expression levels were paralleled with Ang II and AGT expression levels (Figure 4A and 4D).

Expression of TGF-β1 in Crescentic GN
We quantified the expression of TGF-β1 by immunostaining (Figure 5A and B), quantitative PCR (supplemental Figure S2A), and Western blotting (supplemental Figure S2B). Compared with control rats, vehicle-treated GN rats showed elevation of TGF-β1 expression. The drug treatments, particularly CA plus ARB treatment, mostly attenuate these augmentations, consistent with the improved glomerular fibrotic levels and RAS activation. TGF-β1 type 1 receptor (TβR1) expression levels in glomeruli were closely paralleled with TGF-β1 expressions (Figure 5A and 5C).

Effects of Ang II on MCP-1 and TGF-β1 mRNA Levels in Cultured MCs
Following incubation of quiescent MCs with each concentration of Ang II, quantitative PCR revealed an increase in MCP-1 and TGF-β1 mRNA levels (supplemental Figure S3A and S3B). ARB treatment reduced the elevated expression levels of MCP-1 and TGF-β1 (supplemental Figure S3C and S3D). These observations were confirmed by ELISA (supplemental Figure S3E and S3F).

Effects of Ang II on Cell Proliferation and Collagen Secretion in Cocultured PECs and Macrophages With/Without MCs
We examined cell proliferation and collagen secretion in the cocultured PECs and macrophages with/without MCs stimulated with Ang II using the WST-1 assay or Sircol collagen assay, respectively. As a result, ARB treatment completely prevented an increase in cell proliferation or collagen secretion stimulated by Ang II, while CA or pan-specific neutralizing TGF-β antibody alone moderately reduced increases (supplemental Figure S4A and S4B). On the other hand, there were no significant differences between each group in cocol-
tured PECs and macrophages without MCs (supplemental Figure S4C and S4D).

Discussion

To our knowledge, our results provided the first evidence that the combination therapy of CA and ARB markedly attenuates proteinuria and the progression of crescentic GN. MCP-1 is presumed to be a key molecule in chemotaxis and activation of macrophages. CCR2, a cognate receptor of MCP-1 expressed mainly on monocytes, has been reported to be involved in human crescentic GN.15 The strategy of blocking MCP-1/CCR2 interaction might be effective in preventing macrophage-induced tissue damage. Supporting this notion, neutralization of MCP-1 has been reported to reduce macrophage infiltration and progressive kidney damage.3,16,17 Newly developed antagonists against chemokine receptors are now available and have been used as therapeutic agents in kidney injury.18,19 In addition, RS102895 also has the capacity to inhibit MCP-1–induced chemotaxis and renal inflammation in the hypertensive rat model, where MCP-1 plays a role.10 However, few studies have provided direct evidence that the blockade of CCR2 might be effective for the treatment of crescentic GN.20

The RAS plays an important role in the development of hypertension, in fluid and electrolyte homeostasis, and in the progression of renal disease.21,22 Recently, the focus of interest on the RAS has shifted toward the role of the local/tissue RAS in specific tissues.23 The local RAS in the kidney has several pathophysiologic functions, for not only regulating blood pressure but also renal cell growth and production of glomerulosclerosis, which is included in the development of renal fibrosis.24,25 Indeed, previous studies have shown that RAS blockades have beneficial effects in rats and in humans with various renal diseases, and these

Figure 2. Effects of the treatments on the development of glomerular fibrosis in an anti-GBM disease rat model. A, Masson’s trichrome-stained sections and collagen type I (COL I)-immunostained kidney sections. Original magnification ×400 (top, Masson’s trichrome; bottom, COL I); ×200 (middle, Masson’s trichrome). B, Percentage of fibro-cellular crescent glomeruli. C, Quantitative assessment of the glomerular fibrotic area. D, Quantitative real-time reverse transcription-PCR of COL I mRNA in the isolated glomeruli of all treatment groups. E, Densitometric analysis of COL I expression determined by immunostaining. Data are mean±SEM *P<0.05, **P<0.01, and ***P<0.001 between groups as indicated.
produce cytokines and chemokines.19 Infiltration of monocytes into renal tissues is an important mechanism for the development of crescentic GN. From these findings, we believe that suppressing RAS is a key target in the treatment of anti-GBM disease.

Glomerular crescents are defined as the presence of ≈2 layers of cells in the Bowman space. Monocyte/macrophages and PECs are the principle mediators of crescent formation.3 The presence of crescents in glomeruli is a marker of severe injury.28 In the present study, we demonstrated that CA or ARB alone moderately normalized the crescent formation, preventing the infiltration of macrophages. Consistently, the combination therapy markedly reduced crescent formation. The reversibility of crescents correlates with the relative predominance of cellular components.38 The progression or resolution of crescents may depend on the integrity of the Bowman capsule and the resulting cellular composition of the crescent. The progression of crescents to the fibrous stage is often considerably more significant than their suppressive effects on blood pressure.26,27 Based on these principles, here we demonstrated that combination administration of a CA and an ARB very effectively blocks the development of crescentic GN in the anti-GBM disease animal model.

The disturbance in the expression of these components likely plays an important role in the pathogenesis of the crescentic formation in GN. Furthermore, it is reported that Ang II upregulated AGT and Ang II receptor expressions and ARB prevents the increase of AGT, suggesting positive Ang II feedback in kidney.34 Interestingly, ARB treatment prevented increases in kidney and renal interstitial fluid Ang II concentration in the Ang II-infused rat.35 Thus, in our study, combination therapy suppressed these expressions more effectively than CA or ARB alone, cutting intrarenal RAS activation. In previous studies, the RAS activation was shown to be involved in the formation of glomerular crescent.36,37 Together, these data clearly indicate that blocking the RAS is a key target in the treatment of anti-GBM disease. Our results cannot utterly exclude that the lowering blood pressure level by ARB treatment affects the present data. However, it has been shown that RAS blockade has the protective effect for renal injury independent of systemic blood pressure.24,26,27 Furthermore, we have clarified ARB suppressed fibrotic changes of glomerular crescent and the expression of collagen type I in the in vivo study. Also, ARB inhibited cell proliferation and collagen secretion for PECs in the in vitro study. From these findings, we believe that suppressing RAS activation is effective for the attenuation of the progression of crescentic GN.

The expression of MCP-1 is elevated in the glomeruli of patients with crescentic GN and is a mediator of progressive renal injury in GN.30–33 In this anti-GBM disease model, the glomerular expression levels of RAS components were increased compared to control rats. MCP-1-immunostained (bottom) kidney sections. Original magnification ×400. B, Number of CD68-positive macrophages per glomerular cross-section. C, Quantitative real-time reverse transcription-PCR of MCP-1 mRNA in the isolated glomeruli of all treatment groups. D, MCP-1 expression determined by immunostaining. E, Western blot analyses of MCP-1 protein levels in isolated glomeruli of all treatment groups. Data are mean±SEM. *P<0.05, **P<0.01, and ***P<0.001 between groups as indicated.
correlates with irreversible glomerular sclerosis. The combination of CA and ARB also reduced fibrous changes in the glomerular crescent, which was accompanied by suppression of collagen type I induction at both mRNA and protein levels. TGF-β1 is a major profibrotic factor that plays a key role in glomerular sclerosis in GN. Moreover, it is reported that Ang II is the important inducer for TGF-β1. In the present study, ARB treatment reduced the elevated level of TGF-β1 in the crescentic glomeruli. Based on these findings, it suggests that RAS activation enhanced TGF-β1 expression in crescentic GN. Monocyte/macrophage infiltration into the kidney, stimulated by MCP-1 produced from kidney cells in GN, promotes kidney fibrosis and renal injury through secretion of inflammatory cytokines. Although the MCP-1

Figure 4. Effects of the treatments on intrarenal AGT, Ang II, and AT1 receptor (AT1R) expressions and urinary AGT excretion in an anti-GBM disease rat model. A, AGT (top)-, Ang II (middle)-, and AT1R (bottom)-immunostained kidney sections. Original magnification ×400. B, Densitometric analyses of AGT expressions determined by immunostaining. C, Densitometric analyses of Ang II expressions determined by immunostaining. D, Densitometric analyses of AT1R expressions determined by immunostaining. Data are mean±SEM. *P<0.05 and **P<0.01 between groups as indicated. E, Urinary AGT excretion. Data are mean±SEM. *P<0.05 (vs vehicle); **P<0.01 (vs vehicle); ***P<0.001 (vs vehicle).

Figure 5. Effects of the treatments on TGF-β1 expression in an anti-GBM disease rat model. A, TGF-β1-immunostained (top) and TGF-β1 type 1 receptor (TβR1)-immunostained (bottom) kidney sections. Original magnification ×400. B, Densitometric analysis of TGF-β1 expression determined by immunostaining. C, Densitometric analysis TβR1 expression determined by immunostaining. Data are mean±SEM. *P<0.05 and **P<0.01 between groups as indicated.
Ang II induces the expression of MCP-1 and TGF-β1 in cultured MCs, the suppression of these factors by the cotreatment is likely to arrest macrophage infiltrations and fibrous changes for glomerular crescent effectively. From these findings, combination therapy with ARB and CA may confer strong, synergistic effects on glomerular crescent formations by suppressing inflammatory process with macrophage infiltrations and by preventing fibrous changes associated with TGF-β1 overexpressions in crescentic GN.

The next study was performed to determine the molecular and cellular mechanisms underlying the involvement of MCP-1 and the RAS in anti-GBM disease. Glomerular MCs express the relevant receptors for immune complexes and inflammatory cytokines that are responsible for initiation and progression of crescentic GN. In addition, through the stimulation of these cell-surface receptors, MCs produce a range of chemokines and inflammatory mediators that are relevant to the pathogenesis of crescentic GN. The mechanisms from cultured MCs have been shown to be important in subsequent intervention studies in crescentic GN. Furthermore, it is well known that cellular crescents consist of PECs and macrophage. In our hypothesis, MCP-1 and TGF-β1 released from Ang II-stimulated MCs, induces PEC proliferation and fibrosis with macrophage in the course of crescentic GN. Therefore, this in vitro study was designed to elucidate the mechanism of crescent formation, MCP-1 and RAS signal mediators, via glomerular MCs, leading to PEC proliferation and fibrosis. Our in vitro data suggested that Ang II induces the expression of MCP-1 and TGF-β1 in MCs.

Perspectives

The present study reveals that a combination therapy of RAS inhibition with MCP-1/CCR2 signal inhibition can be used to block the MCP-1 and TGF-β1 increases, and thus mitigate renal injury in crescentic GN. Additional studies will be needed to determine the relationship between RAS activation and MCP-1/CCR2 signal in crescentic GN and clarify the mechanism for Ang II-induced fibrotic change of glomeruli. Furthermore, it will be important to determine whether MCP-1/CCR2 signal affects MCP-1 itself, TGF-β1 expression, and which glomerular cell components, such as MCs, PECs, or macrophages, mainly proliferate by the stimulation of MCP-1 in the glomerular crescent. However, it is clear that the combination therapy prevents the infiltration of macrophage and crescent formation in this animal model. We propose that therapeutic effects of the combination could provide a novel pharmacological strategy of anti-GBM disease.

Acknowledgments

We acknowledge critical discussion and/or excellent technical assistance from L. Gabriel Navar, Toshie Saito, Masumi Kamiyama, Akemi Katsurada, M. Patrick Sweeney, G. Michael Upchurch, Nina A. Perrault, Jessica L. Mucci, and Salem I. Elkhayat.

Sources of Funding

This study was supported by grants from the National Institute of Diabetes and Digestive and Kidney Diseases (R01DK072408) and the National Center for Research Resources (P20RR017659).

Disclosures

None.

References

15. Segret S, Cui Y, Hudsins KL, Goodpaster T, Eitner F, Mack M, Schindorff D, Alpers CE. Expression of the chemokine monocyte che-


Addition of Angiotensin II Type 1 Receptor Blocker to CCR2 Antagonist Markedly Attenuates Crescentic Glomerulonephritis

Maki Urushihara, Naro Ohashi, Kayoko Miyata, Ryousuke Satou, Omar W. Acres and Hiroyuki Kobori

_Hypertension_. 2011;57:586-593; originally published online January 31, 2011;
doi: 10.1161/HYPERTENSIONAHA.110.165704

_Hypertension_ is published by the American Heart Association, 7272 Greenville Avenue, Dallas, TX 75231
Copyright © 2011 American Heart Association, Inc. All rights reserved.
Print ISSN: 0194-911X. Online ISSN: 1524-4563

The online version of this article, along with updated information and services, is located on the World Wide Web at:
http://hyper.ahajournals.org/content/57/3/586

Data Supplement (unedited) at:
http://hyper.ahajournals.org/content/suppl/2011/01/28/HYPERTENSIONAHA.110.165704.DC1

Permissions: Requests for permissions to reproduce figures, tables, or portions of articles originally published in _Hypertension_ can be obtained via RightsLink, a service of the Copyright Clearance Center, not the Editorial Office. Once the online version of the published article for which permission is being requested is located, click Request Permissions in the middle column of the Web page under Services. Further information about this process is available in the Permissions and Rights Question and Answer document.

Reprints: Information about reprints can be found online at:
http://www.lww.com/reprints

Subscriptions: Information about subscribing to _Hypertension_ is online at:
http://hyper.ahajournals.org//subscriptions/
ONLINE SUPPLEMENT

Full title of manuscript

ADDITION OF ANGIOTENSIN II TYPE 1 RECEPTOR BLOCKER TO CCR2 ANTAGONIST MARKEDLY ATTENUATES CRESCENTIC GLOMERULONEPHRITIS

Author’s full names and affiliations

Maki Urushihara, MD, PhD, Naro Ohashi, MD, PhD, Kayoko Miyata, PhD, Ryousuke Satou, PhD, Omar W. Acres, MS, Hiroyuki Kobori, MD, PhD

Department of Physiology, and Hypertension and Renal Center of Excellence,
Tulane University Health Sciences Center, New Orleans, LA

Short title

CCR2 and Ang II Receptor on Crescentic Nephritis

Word count of Text: 5,463 (including Title Page, Abstract, Text, References, Tables and Figure Legends), Word count of Abstract: 235, Number of Figures: 5, Number of Supplemental Figures: 4, Number of Tables: 1, Number of Color Figures: 5

Corresponding author

Hiroyuki Kobori, MD, PhD, FAHA, FASN, FACP

Associate Professor of Departments of Medicine and Physiology
Director of the Molecular Core in Hypertension and Renal Center of Excellence
Tulane University Health Sciences Center
1430 Tulane Avenue, #SL39/M720, New Orleans, LA 70112-2699, USA

Tel: +1-504-988-2591, Fax: +1-504-988-0911, E-mail: hkobori@tulane.edu
Detailed Methods

**Regents and Materials.** Anti-collagen type I antibody was purchased from Chemicon International. Anti-CD68 antibody was from Serotec. Anti-MCP-1, AT1 receptor, TGF-β1 and TβR1 antibodies were from Santa Cruz Biotechnology. Anti-Ang II antibody was from Phoenix Pharmaceuticals. Anti-AGT antibody was from Immuno-Biological Laboratories (IBL). Pan-specific TGF-β neutralizing antibody and TGF-β1 Immunoassay kit were from R and D Systems. Rat MCP-1 ELISA Kit was from Thermo Scientific. A 4-[3-(4-iodophenyl)-2-(4-nitrophenoxy)-2H-5-tetrazolio]-1,3-benzene disulfonate (WST-1) was from Roche Applied Science. A Sircol collagen assay was from Biocolor. ARB of olmesartan was kindly provided by Daiichi Sankyo.

**Kidney Histology and Immunohistochemistry**

Glomeruli are considered to exhibit crescent formation when at least 2 layers of cells are observed in the Bowman space. The number of crescents was determined from 50 glomeruli per rat, and this value was expressed as the mean percentage for each group. The ratio of area occupied by crescents in each glomeruli was estimated and assigned one of the following scores: 0, absent; 1, < ¼; 2, between ¼ and ½; 3, between ½ and ¾; and 4, more than ¾ of the whole glomerulus. The mean score of estimated glomeruli was then calculated as the crescent score. The extent of glomerular fibrotic area was quantitatively evaluated by an automatic image analysis, which determined the area occupied by glomerular staining positively for collagen in Masson’s trichrome-stained section (Mass Histology), as described previously. Paraffin-embedded kidney tissues were sectioned, deparaffinized, and incubated with either antibody. Immunohistochemistry was performed by a robotic system (Dako) as previously described. The fraction of fibrotic (blue) or immuno-reactive area (brown) in the glomeruli was measured using the Image-Pro Plus software (Media Cybermetics). For each rat, 20 glomeruli were examined and mean percentages of the affected lesions were calculated. CD68-positive cells (brown) were counted in the glomeruli in each microscopic field. The averaged numbers of macrophages/monocytes in the glomeruli were then obtained for each rat.

**Quantitative Real-Time RT-PCR**

Total RNA was extracted using a commercially available kit (Qiagen) and subjected to DNase 1 treatment (Invitrogen) to eliminate contaminant genomic DNA. Quantitative real-time RT-PCR was performed as previously described; the data thus obtained were normalized to glyceraldehyde-3-phosphate dehydrogenase (GAPDH) mRNA expression levels. The PCR primers used in this study are listed in Supplemental Table S1.

**Western Blot Analysis**

Protein extraction and western blot analysis were performed on samples using an infrared imaging system (LI-COR Biosciences), as described. Appropriate secondary antibodies were purchased from LI-COR.

**Rat Parietal Epithelial Cell (PCE) culture**

Glomerular PECs were cultured using a previously method, with some modification. Glomeruli were isolated from rat kidney by multiple sieving techniques and placed under an inverted tissue culture microscope with phase-contrast optics. Encapsulated glomeruli were selected from tissue suspension by sucking into a micropipette and placed on collagen-coated culture dish. The cultured PECs were out growths from glomeruli and were polygonal, with a cobble-like appearance. Cell clones were proliferated in K1-3T3 medium.

**Cytokine Determination by ELISA**

We carried out sandwich ELISAs for rat MCP-1 or TGF-β1 with supernatants form MCs in accordance with the manufactures’ specifications. MCs were seeded in completed medium into
12-well plate. Twenty-four hours later, cells were washed with PBS and then covered with serum free medium for additional 24 h. For measurement of total TGF-β1, supernatants were acidified with 1 N HCl for 10 min and then neutralized with 1.2 N NaOH right before application.

**Cell Proliferation Assay and Collagen Measurement**

Cell proliferation ability was evaluated by the WST-1 assay according to the manufacturer’s protocol. Briefly, co-cultured cells were grown to sub-confluent in medium containing serum, and subsequently starved for 24 h. Then, cells were cultured for 24 h with or without treatment, and WST-1 reagent was added to cell culture medium in each well. After incubation for 4 h, the absorbency of the samples was measured with a microplate reader at wavelength of 450 nm.

The soluble collagen levels in culture supernatants were measured by using a Sircol collagen assay after incubation for 24 h. This assay measured total secreted collagen from cultured cells. One milliliter of Sirius red, an anionic dye that specifically reacts with basic side chain groups of collagens, was added to the supernatant and incubated with gentle rotation for 30 min at room temperature. After centrifugation, the collagen-bound dye was re-solublized in 0.5 M NaOH, and the absorbance at 540 nm was measured.

**References**


### Supplemental Table S1

<table>
<thead>
<tr>
<th>Gene</th>
<th>Sense Primer</th>
<th>Antisense Primer</th>
<th>Probe</th>
</tr>
</thead>
<tbody>
<tr>
<td>GAPDH</td>
<td>CAGAACATCATCCCTGC ATC</td>
<td>CTGCTTCACCACCTCTTG A</td>
<td>CCTGGAGAAAACCTGCCAAGTATGATGA</td>
</tr>
<tr>
<td>MCP-1</td>
<td>AGCACCCTTTGAATGTGA ACT</td>
<td>AGAAGTGCTTGAGGTGT GT</td>
<td>CCCATAAAATCTGAAGCTAATGCATCC</td>
</tr>
<tr>
<td>TGF-β1</td>
<td>TACCATGCCAACCTCTGTTC</td>
<td>AAGGACCTTGCTGACTGTGT</td>
<td>CCCTACATTTGGAGCCTGGAC</td>
</tr>
<tr>
<td>Collagen type1</td>
<td>GCATCAAGGTCTACTGCAACA</td>
<td>CAGACATGCTTCTTCTCCTT</td>
<td>CCAGAAGAACTGGTACATCAGCCCA</td>
</tr>
</tbody>
</table>
Supplemental Figure S1. The level of plasma creatinine, body weight, total water intake, and daily urine volume in anti-GBM disease rat model. (A) The level of plasma creatinine, (B) body weight, (C) total water intake, and (D) daily urinary volume. Data are mean +/- SEM. * P < 0.05, and ** P < 0.01 between groups as indicated.
Supplemental Figure S2. Effects of the treatment on TGF-β1 expression in anti-GBM disease rat model. (A) Quantitative real-time RT-PCR of TGF-β1 mRNA in the isolated glomerulo of all tratment groups. (B) Western blot analyses of TGF-β1 protein levels in isolated glomeruli of all treatment groups. Data are mean +/- SEM. * P < 0.05, and ** P < 0.01 between groups as indicated.
Supplemental Figure S3. Effects of angiotensin II (Ang II) on cultured mesangial cells (MCs). (A and B) mRNA levels of MCP-1 (A) and TGF-β1 (B) in cultured MCs. MCs were stimulated with Ang II for indicated concentrations and analyzed by quantitative real-time RT-PCR. (C and D) mRNA levels of MCP-1 (C) and TGF-β1 (D) in cultured MCs. MCs were pretreated with Ang II type 1 receptor blocker (ARB) and subsequently stimulated with 100 nM Ang II. Sandwich ELISA for secretion of MCP-1 (E) and TGF-β1 (F) in MCs. MCs were pretreated with ARB (100 nM) and subsequently stimulated with 100 nM Ang II. Data are mean +/- SEM. ** P < 0.01 between groups as indicated. N.S., not significant.
Supplemental Figure S4. Effects of angiotensin II (Ang II) on cultured mesangial cells (MCs) and cultured parietal epithelial cells (PECs) co-cultured with macrophages with/without MCs. (A) Cell proliferation in PECs co-cultured with macrophages and MCs using WST-1 assay. (B) Collagen secretion in PECs co-cultured with macrophages and MCs using Sircol assay. (C) Cell proliferation in PECs co-cultured with macrophages without MCs using WST-1 assay. (D) Collagen secretion in PECs co-cultured with macrophages without MCs using Sircol assay. PECs co-cultured with macrophages were pretreated with ARB (100 nM), CCR2 antagonist (CA, 10 μM) or pan-specific TGF-β neutralizing antibody (TGFβ Ab, 10 μg/ml) and subsequently stimulated with 100 nM Ang II. Data are mean +/- SEM. ** P < 0.01 between groups as indicated. N.S., not significant.