Urinary Plasmin Activates Collecting Duct ENaC Current in Preeclampsia

Kristian B. Buhl, Ulla G. Friis, Per Svenningsen, Ambika Gulaveerasingam, Per Ovesen, Britta Frederiksen-Møller, Bente Jespersen, Claus Bistrup, Boye L. Jensen

Abstract—In nephrotic syndrome, plasminogen is aberrantly filtered from plasma to the urinary space and activated along the tubular system. In vitro, plasmin increases ENaC current by proteolytic cleavage of the γ-subunit. It was hypothesized that preeclampsia is associated with renal ENaC activity present in urine samples from preeclampsia patients. The major protease was identified as plasmin aberrantly filtrated from plasma as plasminogen. In vitro, inward amiloride-sensitive current in collecting duct cells (M1 cells) by whole-cell patch clamp. In PE, urine plasminogen: creatinine ratio was elevated 40-fold (geometric mean, 160 versus 4 µg/g; P<0.0001) and urine aldosterone: creatinine ratio was suppressed to 25% of Ctrl (geometric mean, 27 versus 109 µg/g; P<0.001). A significant negative correlation was found in PE between urinary plasminogen and aldosterone (P<0.05). In PE, proteolytic activity was detected at 90 to 75 kD by gelatin zymography in 14 of 16 patients and confirmed by serine protease assay. Immunoblotting showed active plasmin in PE urine. Whole-cell inward current increased in M1 cells on exposure to urine from PE (173±21%; n=6; P<0.001). The increase in current was abolished by amiloride (2 µmol/L; P<0.001), α2-antiplasmin (1 µmol/L; P<0.001), and heat denaturation (P<0.001). Preeclampsia is associated with urinary excretion of plasminogen and plasmin-dependent activation of ENaC by urine. Proteolytic activation of ENaC by plasmin may contribute to Na+ retention and hypertension in preeclampsia.

Key Words: pregnancy ■ hypertension ■ kidney ■ edema ■ renin ■ aldosterone

Edema is a hallmark of acute nephrotic syndrome, and recent data favor overfill as the pathophysiological explanation with primary, renal hyperabsorption of NaCl through ENaC. Because plasma aldosterone is suppressed in nephrotic syndrome, the alternative pathway of ENaC activation by extracellular proteolysis has been investigated. Soluble proteolytic activity is present in urine samples from nephrotic syndrome patients. The major protease was identified as plasmin aberrantly filtrated from plasma as plasminogen. In vitro, inward amiloride-sensitive current in collecting duct cells is enhanced by exposure to nephrotic urine containing plasmin and by purified plasmin. Plasmin was shown to release an inhibitory peptide tract from the extrodomain of the γENaC subunit either by direct cleavage at high concentrations or through Glycosylphosphatidylinositol-anchored prostaclin at low concentrations. Addition of α2-antiplasmin and aprotinin inhibited the ability of nephrotic urine and plasmin to evoke current. The in vitro cleavage of γENaC is consistent with a distinct shift in migratory pattern of renin and aldosterone activity in manifest gels in proteinuric conditions. Sodium retention and ascites formation is mitigated by the ENaC blocker amiloride in experimental nephrotic syndrome in rats and in children with nephrotic syndrome. Proteinuria can predict the onset of hypertension and, in a Framingham study cohort, urinary albumin: creatinine ratio predicted the development of hypertension. Preeclampsia is a syndrome of pregnancy with abrupt onset of proteinuria and hypertension after 20 weeks of gestation. The cause of preeclampsia is thought to involve release of a soluble fms-like tyrosine kinase (a vascular endothelial growth factor receptor) from placenta leading to decreased bioavailability of circulating vascular endothelial growth factor and endothelial injury, decreased NO formation, vasoconstriction, and glomerular damage with endotheliosis. Preeclamptic (PE) patients exhibit suppressed renin-angiotensin-aldosterone system activity, in particular aldosterone, and avid NaCl retention. Treatment with furosemide in preeclampsia yields less stimulation of plasma renin concentration. These observations are compatible with a primary renal hyperabsorption of NaCl in manifest

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Preeclampsia is associated with fibrin deposition in glomeruli and injury to the glomerular barrier. PE patients display plasmin(ogen) immunoreactivity in urine, to an extent that urine from PE patients has been used as a source to recover human plasminogen and plasmin. These observations led us to hypothesize that manifest preeclampsia is associated with plasmin-dependent protease activity in urine capable of enhancing epithelial ENaC activity. The hypothesis was tested in a cross-sectional design where urine was obtained from patients with preeclampsia and compared with urine obtained from women with uncomplicated pregnancies. The urine samples were tested for their plasmin(ogen) level, proteolytic activity, and ability to activate ENaC.

**Materials**

A cross-sectional study was conducted in 16 patients with newly diagnosed preeclampsia. Preeclampsia was defined as pregnancy beyond 20 weeks associated with newly developed hypertension (office systolic blood pressure >140 mm Hg and diastolic blood pressure >90 mm Hg) and urine protein: creatinine ratio >30 mg/mmol. The patients were compared with 17 healthy (normotensive and nonproteinuric) pregnant women. Control urine samples were obtained from 5 nonpregnant healthy women (age 30±2 years). Spot urine samples were frozen immediately after voiding and kept temporarily at −20°C until storage at −80°C. Samples were thawed on ice, and aliquots were made and stored at −80°C until experiments were performed. The study was approved by the Region of Southern Denmark ethical committee system (Project ID: S-VF-20060095). All patients gave written informed consent to participate in the study.

**Methods**

Blood pressure was measured 3 times after 20 minutes of rest in the seated position using blood pressure monitors from Omron Healthcare Inc (Palatine, IL) and A&D Medical (San Jose, CA). Devices have been approved and validated for clinical use. Before experiments, aliquots of the urine samples were centrifuged at 16,060 G for 30 seconds and the supernatant was used. See the online-only Data Supplement for details concerning measurements of total protein, albumin, aldosterone, creatinine, total plasminogen (plasmino- gen, plasmin, and plasmin-antiplasmin complexes, abbreviated: plasmin[ogen]), and total soluble urokinase plasminogen activator receptor. Details of urine zymography, immunoprecipitation, Western immunoblotting, and serine protease activity assay can be found in the online-only Data Supplement. Single cell patch-clamp experiments were conducted on the cortical collecting duct cell line (M1), as described.

**Statistical Evaluation**

Data were tested for normal distribution. If data were not normally distributed, log-transformation was used to achieve normal distribution. Log-normally distributed data are presented in semilogarithmic diagrams with geometric means. Normally distributed data are presented as mean±SEM. Controls and PE patients were compared by unpaired Student t test. Correlation was evaluated using Pearson correlation (linear dependence). When comparing means of 3 groups (patch clamp data) 1-way ANOVA was used followed by Bonferroni multiple comparison post hoc test. For all data, P<0.05 were considered significant. For all analyses, GraphPad Prism 5 for Windows version 5.02 was used.

**Results**

The control group of healthy pregnant women was not different from the group of PE patients with respect to age or gestational age (Figure S1A and S1B in the online-only Data Supplement). PE patients excreted significantly (P<0.0001) more albumin and total protein in the urine and exhibited elevated systolic and diastolic blood pressures compared with normal pregnancies (P<0.0001; Figure S1C–S1F in the online-only Data Supplement). In PE patients, there was a significantly higher urinary excretion of immunoreactive plasmin(ogen), when normalized to urinary concentration of creatinine (P<0.0001; Figure 1A). Urine excretion of soluble urokinase plasminogen activator receptor was not different between the groups (Figure 1B). When urine proteins were separated by electrophoresis, samples from healthy pregnant women showed no detectable plasmin(ogen) by Western immunoblotting (Figure 2A), whereas urine samples from PE patients displayed prominent bands that migrated with molecular size corresponding with intact plasminogen (83–88 kDa) and active plasmin (83 kDa) and plasmin subunits (57, 26 kDa). Pure plasmin subjected to the same procedure migrated parallel to the most prominent urine band <75 kDa (Figure 2A). In some samples, the higher molecular weight product, likely intact plasminogen (88 kDa), was absent and only active plasmin was present. Immunoprecipitation of urine
yielded a product with molecular size that migrated similarly to pure human plasmin (molecular weight >80 kDa). PE: urine samples from PE patients. Control: urine samples from healthy pregnant women. P represents purified human plasmin 1 µg/85 µL H2O subjected to the same procedure. C, Representative gelatin-PAGE zymography of crude urine samples from PE patients and healthy pregnant women (control). The zymogram shows proteolytic activity at 60 to 90 kDa with a major product around 70 kDa, corresponding with the proteolytic activity of pure human plasmin (P) at 1 µg per well. Urine samples from the control group (Control) did not display protease activity. D, Dilution series for pure human plasmin by a fluorescence-substrate serine protease activity assay. The dilution series of pure plasmin generated the line with concentrations indicated at the x axis. Serine proteases were purified from 1-mL urine samples from PE patients and healthy controls by aprotinin-coated sepharose beads. Eluate was used for fluorescence assay (50 µL). Broken line indicates the geometric mean of serine protease activity obtained in 1 mL of PE patient’s urine (n=13). Urine samples from control pregnancies (n=14) yielded activity measurements not significantly different from 0 by this method. RFU indicates relative fluorescence unit. E, Correlation between urine concentrations of albumin (U-alb) and plasminogen (U-plg). Data were log-transformed to obtain normal distribution. (Pearson correlation; P<0.05; R²=0.2931).

samples for plasmin(ogen) corroborated the presence of predominantly active plasmin in PE urine samples (Figure 2B). Gelatine gel zymography (nonreducing conditions) of pure plasmin showed activity as doublet–triplet that migrated just <75 kDa (Figure 2C). Crude urine samples from patients showed bands that represented proteolysis at similar and slightly larger molecular size (Figure 2C). Significant proteolysis was present in 14 of 16 urine samples from the PE group, whereas 4 of 17 samples showed traces of activity in the control group (Figure 2C). A dilution series of purified plasmin yielded a significant linear regression in serine protease assay (y=0.3012x+1.0121; R²=1; P<0.0001; Figure 2D). Aprotinin-affinity purified urine samples (n=13) showed significant serine protease activity in the tested PE urine samples, whereas urine samples from the control group (n=14) showed no activity (Figure 2D). Mean relative activity corresponded to a plasmin concentration of 1.3 µg/mL (dotted line, Figure 2D). Significant positive correlation (P<0.05) existed between urine albumin concentration and urine plasmin(ogen) concentration (Figure 2E). Murine cortical collecting duct cells (M1) were used as biosensors to elucidate the effect of urine on whole-cell inward current. The voltage clamp protocol evoked minimal inward current in M1 cells by itself (set to 100%), but exposure to control urine produced an increase in inward current that was significantly different from zero, with a mean percentage of change of 42.6±5.7% (Figure 3A and 3D). Urine from nonpregnant women induced an inward current (44.1±23.0%; n=5; not shown), which was not different from healthy pregnant controls. When cells were exposed to urine from PE patients, inward current was significantly larger than measured with urine from healthy pregnant controls (Figure 3B and 3D; P<0.0001). The inward current evoked by PE patient urine samples was abolished when cells were treated with amiloride (2 µmol/L) before exposure to PE urine (Figure 3C and 3D). In the presence of amiloride, whole-cell current after the addition of PE urine was significantly lower than current measured with urine from healthy control pregnancies (P<0.05; Figure 3A and 3D). Treatment with α2-antiplasmin (1 µmol/L) abolished inward current in response to PE urine (P<0.001; Figure 3D). Heat denaturation of urine samples abolished the ability of PE urine to evoke inward current (P<0.001; Figure 3D). Inward current was at the level observed after amiloride and α2-antiplasmin treatment and significantly lower than control urine (P<0.01; Figure 3D). Urine Na/K ratio was not significantly different between Ctrl (2.0±0.3) and PE (1.8±0.3) patients. Urine aldosterone excretion was significantly lower in women with pre-eclampsia compared with healthy controls when normalized for creatinine concentration (Figure 4A) (P<0.001; geometric means). A significant negative correlation (P<0.05) was found between plasmin(ogen) and aldosterone in urine from PE patients (Figure 4B).

Discussion
The present set of data shows significant urinary excretion of plasmin(ogen) and active plasmin in patients with
preeclampsia. Plasmin activity confers to urine samples the ability to activate inward current in single collecting duct cells through ENaC as judged by sensitivity to α2-antiplasmin, amiloride, and heat denaturation. There was a significant inverse correlation between urine aldosterone and plasmin(ogen) and a direct correlation between urine plasmin(ogen) and albumin, which suggests aberrant filtration from plasma. The new information gained on ENaC activation by plasmin in urine from PE patients suggests that ENaC may contribute to an inappropriate renal conservation of Na+ and water and suppression of plasma aldosterone and contribute to hypertension in preeclampsia. Liddle syndrome of hypertension with suppressed aldosterone in which ENaC remains membrane-associated illustrates the significance of dysregulated ENaC for blood pressure.20 A pathological mechanism that includes ENaC-mediated, excessive renal conservation of NaCl in preeclampsia is consistent with a number of previous observations: (1) plasma renin and aldosterone is suppressed with temporal correlation between onset of proteinuria and suppression of renin and aldosterone;21,22 (2) extracellular volume expansion with elevated atrial natriuretic peptide;22–24 (3) more avid retention of NaCl after NaCl infusion;15,25 and (4) diminished responsiveness of renin secretion to furosemide but normal reaction of renin

Figure 3. A to C, Original current traces obtained in single cortical collecting duct cells (M1 line) by patch-clamp recordings. In A to C, gray traces display recordings from cells before superfusion with urine, whereas black traces show recordings from the same cells after superfusion with urine. A, A healthy pregnant control; B, a patient with preeclampsia; and C, urine from the same preeclamptic (PE) patient as in B but after the addition of amiloride. D, Bar graph showing the average changes in current obtained in patch-clamp experiments with single M1 cells in response to superfusion with urine from healthy pregnant controls (Ctrl) and urine from PE patients. The same PE patient urine samples were tested after addition of amiloride (PE+A, 2 µmol/L), α2-antiplasmin (PE+AP, 1 µmol/L), and heat denaturation of the samples (PE+HD). n=6 in all experiments. *P<0.0001; **P<0.001; ***P<0.05, by unpaired Student t test with Bonferroni correction.

Figure 4. A, Aldosterone concentration (U-aldo) normalized to creatinine concentration (U-crea) in spot urine samples from patients suffering from preeclampsia (PE, n=16) and healthy pregnant women (Ctrl, n=17). The thick lines indicate geometric means and thin lines the 95% CI (27 vs 109, µg/g, respectively). *P<0.001, unpaired Student t test. B, Correlation between plasmin(ogen)–(U-plg)/creatinine ratio and aldosterone (U-aldo)/creatinine ratio in spot urine samples from PE patients (n=16). Correlation was estimated by Pearson test; P<0.05; R2=0.2581.
release to postural changes. The K+-sparring antihypertensive drug amiloride normalizes sodium balance and resolves edema in a rat model of nephrotic syndrome and in children with nephrotic syndrome. Previous trials with thiazides and loop diuretics summarized in a Cochrane review have failed to show attenuation of preeclampsia. This could be because of exaggerated Na+ reabsorption along the collecting duct that might abrogate the effect of diuretics that target Na+ transport proteins in more proximal nephron segments. Proof-of-concept for ENaC involvement in clinical manifestations of preeclampsia requires a randomized intervention study. So far, only casuistic reports are available: a pregnant patient with Conn syndrome and 2 pregnant women with Bartter syndrome received amiloride therapy throughout pregnancy or beyond week 17 and in doses ≤15 to 20 mg/day in one of the cases. These pregnancies resulted in healthy offspring. Plasminogen is produced predominantly by the liver and circulates at slightly elevated plasma concentrations in normal human pregnancies. The present study confirms significant excretion of plasminogen and shows active plasmin in urine from PE patients. An integral membrane protein plasminogen receptor, Plg-Rγ, has been identified. Plg-RKT promotes activation of plasminogen and colocalizes with urokinase-type plasminogen activator receptor (uPAR). Urokinase-type plasminogen activator is abundant in urine and requires activation by uPAR. The present data show urinary excretion of soluble uPAR at similar levels between PE patients and healthy pregnant women. This is compatible with an epithelial origin of soluble uPAR immunoreactivity likely caused by shedding rather than aberrant filtration from plasma where it is slightly elevated in preeclampsia. Our previous data have shown the existence of a cell-attached cascade whereby low concentrations of plasmin (≤1 µg/mL) activates cell-anchored prostasin. At the measured concentrations of active plasmin in urine, involvement of amplification, that is, prostasin, in the activation of ENaC, is likely. Adjacent cleavage sites for plasmin and prostasin have been identified in the N terminal of γENaC. According to the stepwise proteolytic activation concept proposed by Kleyman et al, plasmin and prostasin elicit extracellular second hit proteolysis subsequent to intracellular furin to release an inhibitory peptide from γENaC. This causes full activation of the channel.

Perspectives

There is no causal treatment for preeclampsia. The present cross-sectional data show a new mechanism that may couple a defective glomerular barrier to activation of Na+ transport by the collecting duct epithelium in preeclampsia. Active plasmin confers to preurine the ability to activate ENaC. This may promote reabsorption of Na+ by the collecting duct and contribute to extracellular volume expansion, edema, suppression of renin and aldosterone, and hypertension. The present set of human data offers a mechanism that can be further tested for causality in intervention studies, and it provides a range of potential targets for pharmacologic intervention in severe preeclampsia, including: urine proteolytic activity/plasmin, plasminogen receptor, prostasin, urokinase-type plasminogen activator-uPAR, and ENaC.

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Disclosures

None.

References

What Is New?

The study provides a new disease mechanism for an old disease, toxemia of pregnancy, a syndrome of pregnancy that is composed of hypertension and protein excretion in urine. Aberrant loss of a plasma enzyme into preurine in the kidneys is coupled to activation of renal salt retention.

What Is Relevant?

Preeclampsia affects globally 3% to 5% of all pregnancies and is the leading cause of maternal and fetal mortality. No causal treatment exists for preeclampsia. The study provides a range of potential new drug targets that should be further explored in future research.

Summary

The study shows a new coupling between aberrant urine enzyme activity and inappropriate renal NaCl retention in preeclampsia. This mechanism may explain the development of hypertension and edema and provide new drug targets to alleviate symptoms.
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Urinary plasmin activates collecting duct ENaC current in preeclampsia

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Supplementary Methods

Urinalysis

Before experiments, aliquots of the urine samples were centrifuged at 13,000 rpm for 30 seconds and experiments were done using the supernatant. Concentrations of urinary total protein and urinary albumin were determined on the Cobas Mira Plus device using ABX Pentra reagents (urinary total protein CP, ref: A11A01642 and Micro-albumin CP, ref: A11A01623; Triolab A/S, Brøndby, Denmark) following the manufacturer’s protocol. Urine aldosterone concentration was measured using Coat-A-Count® Aldosterone kit (Siemens Healthcare Diagnostics Inc. Los Angeles, CA, USA) following the manufacturer’s instructions. Urinary creatinine concentrations were determined spectrophotometrically using Microlab 300 (Vital Scientific BV, AC Dieren, NL).

Measurement of total plasminogen (plasminogen, plasmin and plasmin-anti-plasmin complexes, abbreviated: plasmin(ogen)) content in urine samples was performed using a commercial ELISA kit (Human Plasminogen Total Antigen Assay (Cat# IHPLGKT-TOT, Innovative Research, Novi, Michigan, USA). Measurement of total soluble urokinase plasminogen activator receptor (SuPAR) content in urine samples was performed using a commercial ELISA assay (SuPARnostic® Standard Kit, Virogates A/S, Birkeroed, DK). Urinary serine protease activity was tested using 10% Zymogram (gelatin) gel. For quantitative protease assay and additional western immunoblotting, urine samples were pretreated using aprotinin (USB, Cleveland, OH, USA) coupled to CNBr-activated Sepharose 4B (Amersham Bioscience, Hillerod, DK) as previously described using 1 mL of urine.

Immunoprecipitation Plasmin(ogen) was immunoprecipitated from 85 µl urine using polyclonal goat anti-plasminogen (ab 6189-100, Abcam, Cambridge, UK) and the Protein G immunoprecipitation kit (IP50, Sigma, St. Louis, Missouri, USA) following the manufacturer’s protocol.

Western Immunoblotting The immunoprecipitate or the aprotinin plasmin(ogen) purification eluate (see quantitative protease assay below) was mixed with NuPAGE® Sample Reducing Agent (10X) (Invitrogen, Carlsbad, CA, USA), NuPAGE® LDS Sample Buffer (4X) (Invitrogen), and heat denatured. Samples were run on a Ready Gel 7.5% Tris-HCl (BioRad Laboratories Inc. Hercules, CA, USA), and subsequently blotted onto a Immobilon™ polyvinylidene difluoride membrane (Millipore, Immobilon-P transfer membrane, pore size 0.45µm, Millipore Corporation, Bedford, MA, USA). The polyclonal goat anti-human plasminogen ab 6189-100 (Abcam) was used as primary antibody and HRP-conjugated anti-goat (DakoCytomation, Glostrup, Denmark) was used as secondary antibody. Blots were developed using AmershamTM ECL™ system.

Serine protease activity Urinary protease activity was tested using 10% Zymogram (gelatin) gel, 1.0 mm x 10 well (Invitrogen, Carlsbad, CA, USA) according to manufacturer’s protocol loading 10 µL urine and 10 µL Novonex® Tris-Glycine SDS Sample buffer (x2) (Invitrogen, Carlsbad, CA, USA) per well. For quantitative protease assay, urine samples were pretreated using Aprotinin (USB, Cleveland, OH, USA) coupled to CNBr-activated Sepharose 4B (Amersham Bioscience, Hillerod, DK) as previously described (Svenningsen et al 2009) using 1 mL of urine. The beads were pelleted and washed thoroughly. Bound proteins were eluted using 100µL PBS, pH 2 added 10 µL 1 mol/L Trizma per sample. Quantitative measurements of serine protease activity in the urine samples were conducted using EnzChek® Peptidase/Protease Assay Kit (E33758), (Molecular Probes™, Eugene, OR, USA) following the manufacturer’s protocol. Readings were done using Fusion™, Packard using software Fusion Instrument Control application v4.0, Perkin Elmer®.
Single cell (CCD M1) Patch-clamp experiments The experiments were conducted on the cortical collecting duct cell line (M1) from ATCC (Boras, Sweden) as previously described. Briefly, M-1 cells were grown to confluence in 25 cm² flasks (Nunc, Roskilde, Denmark) followed by trypsination and seeding onto coverslips in DMEM:F12 (Life Technologies, Taastrup, Denmark) added 5 µmol/L dexamethasone (Sigma, Brøndby, Denmark). M-1 cells incubated at 37°C/5% CO₂ and single cell patch clamp experiments were performed 24 to 48 hours after the cells were seeded. Experiments were performed at room temperature in the tight-seal whole-cell configuration of the patch-clamp technique with heat-polished patch pipettes with resistances of 5-7 MΩ. Seal resistances were in the range of 2-15 GΩ. High resolution membrane currents were recorded with an EPC-9 patch-clamp amplifier (HEKA) controlled by PULSE v8.11 software on a Power Macintosh G3 computer. The current was monitored by the response to a voltage step of -160 mV for 200 ms from a holding potential of -60 mV (this pulse was repeated every 3rd second throughout the entire experiment). After 30-60 seconds, the cell was gently flushed with urine and the current monitored.

List of References

Figure S1

A

Gestational Age Days

250

150

PE Ctrl

B

Age Years

40

30

20

10

0

PE Ctrl

C

Systolic BP mm Hg

150

50

PE Ctrl

D

Diastolic BP mm Hg

150

50

PE Ctrl

E

ACR mg/g

100

10

PE Ctrl

F

U-protein total/U-crea mg/g

10000

100

PE Ctrl
**Figure legends S1**

Description of the two groups showing values from preeclamptic patients and healthy pregnant controls with respect to gestational age, age, arterial blood pressure, urine albumin and protein. Preeclamptic patients (PE, n=17) compared to healthy pregnant controls (Ctrl, n=16).

**A-B,** Figures show gestational age (GA) in days (A) and age of each individual in years (B) for preeclamptic patients (PE) and healthy pregnant controls (Ctrl). Mean values GA: PE 242.4 ± 6.5 days vs. Ctrl 241.9 ± 4.8 days, Age: PE 32.0 ± 1.6 years vs. Ctrl 31.6 ± 1.4 years,

**C-D,** Figures show distribution of systolic (C) and diastolic (D) blood pressures in preeclamptic patients (PE) and healthy pregnant controls (Ctrl). Systolic blood pressure means; PE: 158.5 ± 3.1 mmHg vs. Ctrl: 116.9 ± 2.1 mmHg, diastolic blood pressure means; PE: 100.5 ± 2.2 mmHg vs. Ctrl: 74.9 ± 1.8 mmHg, *P < 0.0001, unpaired Student’s t-test.

**E,** The figure shows distribution of the urine albumin-creatinine-ratio (ACR) in the two groups PE and Ctrl. (Geometric mean 367.3 mg/g vs. 3.6 mg/g), *P < 0.0001, unpaired Student’s t-test.

**F,** The figure shows distribution of total urine protein concentration normalized to urine creatinine concentration in the two groups PE patients and control. (Geometric mean: 1002.3 mg/g vs. 89.9 mg/g), *P < 0.0001, unpaired Student’s t-test.