Angiotensin Type 2 Receptor Stimulation Ameliorates Left Ventricular Fibrosis and Dysfunction via Regulation of Tissue Inhibitor of Matrix Metalloproteinase 1/Matrix Metalloproteinase 9 Axis and Transforming Growth Factor β1 in the Rat Heart

Dilyara Lauer, Svetlana Slavic, Manuela Sommerfeld, Christa Thöne-Reineke, Yuliya Sharkovska, Anders Hallberg, Bjorn Dahlfö, Ulrich Kintscher, Thomas Unger, Ulrike Muscha Steckelings,* Elena Kaschina*  

Abstract—Left ventricular (LV) remodeling is the main reason for the development of progressive cardiac dysfunction after myocardial infarction (MI). This study investigated whether stimulation of the angiotensin type 2 receptor is able to ameliorate post-MI cardiac remodeling and what the underlying mechanisms may be. MI was induced in Wistar rats by permanent ligation of the left coronary artery. Treatment with the angiotensin type 2 receptor agonist compound 21 (0.03 mg/kg) was started 6 hours post-MI and continued for 6 weeks. Hemodynamic parameters were measured by echocardiography and intracardiac catheter. Effects on proteolysis were studied in heart tissue and primary cardiac fibroblasts. Compound 21 significantly improved systolic and diastolic functions, resulting in improved ejection fraction (71.2±4.7% versus 53.4±7.0%; P<0.001), fractional shortening (P<0.05), LV internal dimension in systole (P<0.05), LV end-diastolic pressure (16.9±1.2 versus 22.1±1.4 mm Hg; P<0.05), ratio of early (E) to late (A) ventricular filling velocities, and maximum and minimum rate of LV pressure rise (P<0.05). Compound 21 improved arterial stiffness parameters and reduced collagen content in peri-infarct myocardium. Tissue inhibitor of matrix metalloproteinase 1 was strongly upregulated, whereas matrix metalloproteinases 2 and 9 and transforming growth factor β1 were diminished in LV of treated animals. In cardiac fibroblasts, compound 21 initially induced tissue inhibitor of matrix metalloproteinase 1 expression followed by attenuated matrix metalloproteinase 9 and transforming growth factor β1 secretion. In conclusion, angiotensin type 2 receptor stimulation improves cardiac function and prevents cardiac remodeling in the late stage after MI, suggesting that angiotensin type 2 receptor agonists may be considered a future pharmacological approach for the improvement of post-MI cardiac dysfunction. (Hypertension. 2014;63:e60-e67.)  

Key Words: matrix metalloproteinase 9 • myocardial infarction • receptor, angiotensin, type 2 • TIMP1 protein, human

Heart failure (HF) secondary to myocardial infarction (MI) remains a major source of morbidity and mortality, despite significant advances in therapies developed for this disease. The major determinant of survival after recovery from MI is left ventricular (LV) remodeling, which has been strongly associated with clinical outcomes in numerous HF trials.  

The cardiac renin–angiotensin–aldosterone system, which is highly activated after MI and in HF, has been implicated in the process of vascular and cardiac remodeling. Angiotensin II through its type 1 receptor (AT1R) induces myocardial hypertrophy, fibrosis, and cardiomyocyte apoptosis. Consequently, AT1R antagonists inhibit fibrosis in post-MI hearts. Both angiotensin-converting enzyme inhibitors and AT1R antagonists are cardioprotective with regard to post-MI LV dysfunction. Beneficial effects of AT1R blockade have at least, in part, been attributed to stimulation of unopposed angiotensin.
by guest on May 29, 2017 http://hyper.ahajournals.org/ Downloaded from

Cardiac AT2R overexpression attenuates LV remodeling after MI. We studied the potential underlying mechanisms of promoting antigrowth, antiproliferation, and antifibrosis.6

We have recently demonstrated a profound improvement of cardiac function 7 days after MI by direct AT2R stimulation.15 Furthermore, C21 prevented vascular remodeling in L-NAME (N \textsuperscript{-} nitro-L-arginine methyl ester) hypertensive rats and stroke-prone spontaneously hypertensive rats,17 and it preserved renal integrity by preventing inflammatory cell infiltration and collagen accumulation.18 However, it has not been investigated yet whether chronic AT2R stimulation has any effect on cardiac remodeling and resulting HF after MI.

The present study was designed to assess the long-term effects of treatment with the nonpeptide AT2R agonist C21 on cardiac function and structural myocardial alterations 6 weeks after MI. We studied the potential underlying mechanisms of AT2R activation on the heart by exploring extracellular matrix turnover processes of the postischemic myocardium, especially with regard to the tissue inhibitor of metalloprotease 1 (TIMP1) and matrix metalloprotease 9 (MMP9) axis.

Methods

Animals
This study was performed in strict accordance with National and European guidelines for animal experiments with approval by the ethics commission of the regulatory authorities of the City of Berlin, Germany, the Landesamt fuer Gesundheit und Soziales (registration number G0307/06).

Experimental Protocol
MI was induced by ligation of proximal left anterior descending coronary artery as previously described.13 Male Wistar rats were randomly assigned to the following groups: C21 treatment (0.03 mg/kg daily IP), vehicle treatment (0.9% NaCl), and sham-operated controls. Treatment was started 6 hours after MI. Animals with ejection fraction <35% were excluded from the experiment before assignment to groups. C21 was kindly provided by Vicore Pharma (Gothenburg, Sweden).

Analysis of Hemodynamic Parameters
Transsthoracic Doppler echocardiography (M-mode and Doppler measurements) was performed 1 and 6 weeks after MI in rats anesthetized with isoflurane (2%) with the use of the high-resolution imaging system Vevo 770 (VisualSonics, Canada), and invasive hemodynamic assessment was performed at the end of the study using a fiber-optic pressure transducer catheter (Samba Sensors, Sweden) and Chart5 software for analysis as previously described.15,19

Quantitative Real-Time Reverse Transcription Polymerase Chain Reaction, Western Blot Analysis, and ELISA
Quantitative real-time reverse transcription polymerase chain reaction for measurement of MMP9, MMP2, and transforming growth factor β1 (TGF-β1) mRNA in heart tissues and TGF-β1 in cardiac fibroblasts (CFs) in vitro, as well as immunoblotting for MMP2, MMP9, TGF-β1, and TIMP1 protein expression in LV, was performed according to standard protocols. TGF-β1 concentration in plasma was determined by ELISA. TGF-β1 expression in LV was studied immunohistochemically. Details of primers and antibodies can be found in the online-only Data Supplement.

Histology and Hydroxyproline Content in the Heart
Paraffin-embedded cross-sections of the heart were stained for collagen (sirius red/picric acid). Myocytometer diameter was quantified by means of computer-aided histomorphometry by using of a χ40 objective (χ60 on monitor). The diameters were determined on a minimum of 30 myocytes from each animal (n=4) in noninfarcted regions. Interstitial fibrosis was quantified by means of computer-aided histomorphometry (magnification χ60). Results were calculated as percentage of sirius red–positive areas in randomly chosen pictures from cardiac sections. The amount of collagen in the LV was analyzed by quantification of the tissue hydroxyproline content using the modified method based on alkaline hydrolysis as previously described.16,19

Cell Culture Experiments
Primary rat CFs were isolated as described by Agocha et al.20 CFs were incubated with interleukin 1β (IL-1β; 20 ng/mL, Sigma) in the presence or absence of C21 (0.25, 0.5, 1 μmol/L) and PD123319 (10 μmol/L). MMP2 and MMP9 activity was analyzed by gelatin zymography as described previously.21,22 mRNA expression of MMP9, MMP2, and TGF-β1 was evaluated by quantitative real-time reverse transcription polymerase chain reaction.

Fluorescence Microscopy
MMP9 and TIMP1 release was studied by fluorescence microscopy (Biorveo BZ-9000, Keyence Japan) in single CFs after stimulation with IL-1β (20 ng/mL) in the presence or absence of C21 (1 μmol/L) for 1, 4, 24, and 48 hours. Antibodies are specified in the online-only Data Supplement.

Statistical Analysis
Results are expressed as mean±SEM. Multiple comparisons were analyzed with 1-way ANOVA followed by the Bonferroni post hoc test. Two-group comparisons were analyzed by the 2-tailed Student unpaired t test for independent samples. Differences were considered statistically significant at a value of P<0.05.

Results
C21 Improves Systolic and Diastolic Cardiac Functions 6 Weeks After MI
Postinfarct mortality was 31%. Animal losses all occurred within the first 6 hours after MI before animals were assigned to the respective treatment groups.

Echocardiographic outcomes obtained 6 weeks after MI are presented in the Table. MI induced a significant LV dilation reflected by increased systolic LV inner diameter (P<0.05). Systolic function was impaired as evidenced by a decrease in LV ejection fraction (P<0.001), fractional shortening (P<0.01), and stroke volume (P<0.05). MI also deteriorated LV diastolic function, leading to an increase in the ratio of early (E) to late (A) ventricular filling velocities (E/A ratio; P<0.05) and shortening of E-wave deceleration time (P<0.01; Table).

Chronic treatment with C21 significantly improved systolic function, raising ejection fraction by 17.8% (P<0.001) and fractional shortening by 9.9% (P<0.05) versus vehicle. C21
also reduced systolic LV inner diameter from 5.2±0.7 mm in vehicle-treated to 4.2±0.4 mm (P<0.05) in C21-treated rats and diastolic LV inner diameter from 7.8±0.6 to 7.8±0.5 mm (P<0.05), respectively. C21 further improved cardiac wall motion (Figure S1A in the online-only Data Supplement). Improvement of diastolic function by C21 was evidenced by a 17.4% decrease in E/A ratio (P<0.05; versus vehicle; Table; Figure S1B).

Hemodynamic parameters measured via intracardiac Samba catheter are presented in Figure 1. Rats with MI developed systolic dysfunction, as evidenced by decreased contractility index (125.4±4.2 versus sham 142.3±2.0 L/s; P<0.05; Figure 1A) and maximal peak rate of LV pressure increase (dP/dt\text{max}; 936±369 versus sham 11959±420 mm Hg/s; P<0.05; Figure 1B).

Rats with MI also exhibited a severe diastolic dysfunction, as defined by elevated LV end-diastolic pressure (22.1±1.4 versus sham 14.4±1.0 mm Hg; P<0.05; Figure 2D), increased minimal peak rate of LV pressure increase (dP/dt\text{min}; –10262±464 versus sham –12524±529 mm Hg/s; P<0.05; Figure 1E), and reduced diastolic duration time (P<0.01; Figure 1F).

C21 significantly improved systolic cardiac function, enhancing contractility index (141.7±4.2 versus vehicle 125.4±4.2 L/s; P<0.05) and dP/dt\text{max} (11540±594 mm Hg/s versus 9364±369 mm Hg/s; P<0.05; Figure 1A and 1B) and reducing LV maximal pressure (P<0.05; Figure 1C).

C21 demonstrated favorable effects on the diastolic function by reducing LV end-diastolic pressure (16.9±1.2 versus vehicle 22.1±1.4 mm Hg; P<0.05) and dP/dt\text{min} (–11 805±398 versus vehicle –10 262±464 mm Hg/s; P<0.05; Figure 1D and 1E) and by increasing diastolic duration time (P<0.05; Figure 1F).

To assess indirect parameters of arterial stiffness, pulse wave analysis was performed on pressure curves obtained from the ascending aorta. The augmentation index in vehicle-treated animals was increased (25.4±1.2 versus sham 15.57±1.3; P<0.05), and C21 attenuated this increase (17.8±1.2 versus vehicle 25.4±1.2; P<0.05).

### Table. Hemodynamic Variables in Wistar Rats Measured 6 Weeks After MI by Transthoracic Doppler Echocardiography

<table>
<thead>
<tr>
<th>Parameter</th>
<th>Sham</th>
<th>Vehicle</th>
<th>C21</th>
</tr>
</thead>
<tbody>
<tr>
<td>EF, %</td>
<td>74.6±2.51</td>
<td>53.4±7.03</td>
<td>71.2±4.74†</td>
</tr>
<tr>
<td>FS, %</td>
<td>50.5±5.19</td>
<td>30.9±3.08‡</td>
<td>40.8±2.32§</td>
</tr>
<tr>
<td>LVIDd, mm</td>
<td>7.9±0.14</td>
<td>8.7±0.63</td>
<td>7.8±0.54§</td>
</tr>
<tr>
<td>LVIDs, mm</td>
<td>4.7±0.47</td>
<td>5.2±0.76</td>
<td></td>
</tr>
<tr>
<td>E/A</td>
<td>1.8±0.17</td>
<td>2.3±0.13</td>
<td></td>
</tr>
<tr>
<td>EDT, ms</td>
<td>17.8±2.29</td>
<td>12.4±0.78</td>
<td></td>
</tr>
<tr>
<td>Stroke volume, µL</td>
<td>44.3±5.37</td>
<td>35.6±1.42</td>
<td></td>
</tr>
<tr>
<td>Heart rate, bpm</td>
<td>339.3±11.84</td>
<td>357.8±6.46</td>
<td></td>
</tr>
</tbody>
</table>

Also, C21 prevented the IL-1α-stimulated secretion of MMP9 in a concentration-dependent manner, whereas MMP2 release was not influenced. AT1R blockade with PD123319 thoroughly abolished the effect of C21 on the regulation of MMP2, MMP9, and TGF-β1 in the heart 6 weeks after MI.

**Myocyte Diameter, Collagen Content, and Expression of TGF-β1 in the Heart 6 Weeks After MI**

Myocyte diameter was significantly increased 6 weeks post-MI (12.57±1.85 versus sham 10.56±1.50 µm; P<0.01), and C21 treatment further increased the diameter of myocytes (15.38±1.30 versus vehicle 12.57±1.85 µm; P<0.001; Figure S2).

Less collagen accumulation was found in the heart after C21 treatment as demonstrated by staining with the collagen-specific dye sirius red (Figure S3). Interstitial fibrosis was markedly increased in the noninfarcted region of vehicle-treated post-MI animals (5.5±0.3% versus sham 2.8±0.3%; P<0.01); treatment with C21 completely prevented this effect (3.0±0.2% versus vehicle 5.5±0.3%; P<0.001; Figure S4A).

Collagen content was studied by measuring tissue hydroxyproline concentration. Hydroxyproline increased by 24% (P<0.01) post-MI compared with sham group, whereas C21 attenuated this increase (P<0.05; Figure S4B).

Because fibrosis is promoted by TGF-β1, its expression was investigated in the peri-infarcted zone. In the vehicle group, TGF-β1 was strongly enhanced in fibrotic tissues and cardiomyocytes (Figure 2A). C21 prevented the overexpression of TGF-β1.

Accordingly, TGF-β1 expression in the LV, which was significantly upregulated both on the mRNA (1.3-fold; P<0.05) and particularly on protein levels (40-fold; P<0.01) post-MI, was completely abolished by C21 (P<0.01; Figure 2B and 2C).

Furthermore, serum concentration of active TGF-β1 tended to be elevated 6 weeks post-MI, whereas the latent form of TGF-β1 was decreased (P<0.05; Figure 2D and 2E). C21 treatment normalized the levels of both forms (P<0.05).

### Regulation of TIMP1, MMP2, and MMP9 by C21 in the LV 6 Weeks After MI

As demonstrated in Figure 3A and 3B, MMP9 was upregulated post-MI both at the mRNA (2.9-fold; P<0.05) and protein levels (3.6-fold; P<0.001). C21 treatment significantly attenuated this increase.

MMP2 protein but not mRNA was also upregulated (Figure S5). Zymographic analysis of the LV post-MI extracts demonstrated an increased release of active MMP9 (Figure 3C and 3D). Again, C21 significantly prevented MMP9 secretion, whereas there was no statistically significant effect on MMP2 activity (Figure 3E). Protein expression of TIMP1, the native inhibitor of MMP9, was lower after MI (Figure 3F). C21 strongly reversed this decrease (17.7-fold; P<0.01) and, in turn, attenuated the increased MMP9/TIMP1 ratio (Figure 3G).

### Regulation of TIMP1, MMP2, MMP9, and TGF-β1 Expression in Rat Primary CFs

Zymographic analysis of MMP9 and MMP2 activity in CFs is presented in Figure 4. C21 prevented the IL-1α-stimulated secretion of MMP9 in a concentration-dependent manner, whereas MMP2 release was not influenced. AT1R blockade with PD123319 thoroughly abolished the effect of C21 on the regulation of TIMP1, MMP2, and MMP9.
IL-1α–induced MMP9 secretion (Figure S6). C21 also significantly attenuated TGF-β1 mRNA upregulation (Figure S7).

The results of TIMP1 and MMP9 immunostaining in CFs are presented in Figure 5. After stimulation with IL-1α, MMP9 (red stain) was expressed in cytoplasm already at 4 hours and reached its maximum at 48 hours. TIMP1 (green stain) appeared 24 hours after stimulation, was colocalized (yellow stain) with MMP9, and disappeared at 48 hours. In C21-treated cells, TIMP1 was strongly expressed already 4 hours after IL-1α stimulation, reached maximum at 24 hours, and was distinguishable until 48 hours in a colocalization with MMP9. In C21-treated cells, even at 48 hours after stimulation not linked MMP9 (red stain) was undistinguishable.

**Discussion**

This study describes protective effects of long-term selective AT₂R stimulation with the AT₂R agonist C21 in a chronic model of MI. C21 improved both systolic and diastolic heart functions, largely prevented post-MI LV dysfunction, and reduced arterial stiffness.

The present study is in line with our previous data obtained 7 days after MI, which showed an improvement of cardiac function in the acute phase after MI. In this earlier study, the favorable effect of AT₂R stimulation with C21 on cardiac function was prevented by the AT₂R antagonist, PD123319, confirming AT₂R specificity of C21 actions. Hemodynamic data are also in agreement with recent investigations performed in an AT₂R overexpression model, where the AT₂R protected the heart from ischemic injury 4 weeks post-MI. However, in a study looking at cardiac function and remodeling in a model of MI induced by coronary occlusion–reperfusion in mice, no attenuation of post-MI LV remodeling by a 4-week treatment with C21 was observed. The lack of treatment effect of C21 in this study was probably because of the application of the same dose of C21 that is usually used for bolus injections (0.3 mg/kg bw) by minipump for 24 hours, which considering a plasma half-life of C21 of maximum 4 hours results in too low plasma levels and thus in underdosing.

In the late phase post-MI, adverse postischemic cardiac remodeling is characterized by fibrotic processes and collagen deposition in the remote myocardium, which have the potential of increasing LV mass at the cost of LV function. Therefore, the role of AT₂R stimulation in post-MI cardiac
fibrosis and the underlying mechanisms were the focus of this study.

We found that C21 decreased interstitial fibrosis and collagen accumulation in the remote myocardium after ischemia. In parallel, C21 reduced the increased augmentation index, which characterizes arterial stiffness and correlates with vascular fibrosis, as well as with the severity of LV hypertrophy and coronary heart disease.26 These data are in agreement with previous reports demonstrating an antifibrotic effect of the AT2R agonist in the aorta of L-NAME–treated hypertensive rats16 and in the vessels and heart of SHR.17

Interestingly, C21 administration reinforced post-MI compensatory hypertrophy of cardiac myocytes and by this mechanism possibly contributed to functional improvement of the heart.

A similar trend was seen by Voros et al.,27 but in their study did not reach statistical significance probably because of limited power of the study as the authors state in the discussion.

We further investigated the effect of AT2R stimulation on TGF-β1 expression, a profibrotic cytokine that is extensively expressed after ischemia and is upregulated by angiotensin II via AT2R activation in cardiac myocytes and fibroblasts.28,29 This study is the first to demonstrate that AT2R stimulation prevents the increase in TGF-β1 expression after MI, both at protein and transcriptional levels. The elevated serum concentration of active TGF-β1 was also decreased by C21. In vitro experiments in primary CFs further demonstrated that C21 attenuated the IL-1α–induced increase in TGF-β1 mRNA expression, suggesting its regulation through a direct effect of AT2R stimulation on the regulation of TGF-β1 expression.

The degradation of matrix components by MMPs and the resulting side-by-side slippage of cardiomyocytes are other mechanisms that may contribute to LV wall thinning in the remote region after MI.25 It is, therefore, suggested that the interplay of MMPs and their regulators is important in the development of myocardial fibrosis and post-MI LV dysfunction and that HF can be modified by modulating MMP activity.

Gelatinases MMP2 and MMP9 have been shown to be main regulators of ECM (extracellular matrix) in cardiac tissue because they possess substrate affinity for denatured fibrillar collagen and exhibit proteolytic activity against elastin and proteoglycans.30

MI induced the proteolytic activity of MMP9 in the LV, which was attenuated by C21 treatment. Similarly, C21 dose dependently decreased gelatinolytic activity of MMP9 in CFs. Furthermore, C21 regulated the expression of TIMP1, the natural inhibitor of MMP9. Deficiency of TIMP1 is known to promote LV dilatation and increase LV end-diastolic volume31 and has also been demonstrated to exacerbate LV remodeling after MI in mice after MI.32

In our study, the higher concentration of MMP9 and the increased MMP9/TIMP1 protein expression ratio in cardiac tissue post-MI evidenced the proteolytic dysbalance, which was ameliorated by treatment with C21. The molecular mechanisms, by which the AT2R inhibits proteolysis, are complex and require further investigation, but several possible interactions exist.

First, the AT2R seems to be involved in the interplay between MMPs and TGF-β1. Increasing evidence points toward the
possibility that TGF-β1 increases MMP activity within the myocardium. However, MMP9 and MMP2 are both able to cleave latent TGF-β1, releasing its active form, which in turn activates transcription of TIMPs. This suggests that a feed-forward loop could develop in cardiac remodeling, in which the roles of MMPs and TGF-β1 are intertwined.
TIMP1 activation may contribute to antiproteolytic effects of C21 in the heart. Our immunostaining experiments performed in single CFs and determining the time course of AT_2R-induced expression of TIMP1 and MMP9 suggest that C21 in a first step activates TIMP1 followed by inhibition of MMP9. TIMP1 is also known to possess antiapoptotic activity, and this fact is in accordance with antiapoptotic properties of C21 demonstrated previously. Thus, TIMP1 may be a major mediator of the cardioprotective effects of C21. Given that MMPs generally play a pivotal role in extracellular matrix degradation, the antiproteolytic effect of C21 might be of great relevance for tissue remodeling in various pathologies.

It is a limitation of this study that we did not have an animal group treated with C21 plus an AT_2R antagonist (PD123319) to prove AT2R specificity of the observed protective effects of C21. However, AT_2R specificity of a beneficial effect of C21 on post-MI cardiac performance has been shown by us in a previous study with shorter duration. Furthermore, we demonstrate in this study in vitro experiments that the inhibitory effect of C21 on MMP activity, which is an entirely new finding, is AT2R specific, that is, reversible by PD123319.

**Perspectives**

This study demonstrates that long-term stimulation of the AT_2R with C21 improved postischemic cardiac function by preventing adverse myocardial remodeling and reducing proteolysis. Furthermore, C21 activated TIMP1, inhibited MMP9, and decreased TGF-β1 expression as well as collagen accumulation in the heart.

This study is of clinical relevance because it supports the hypothesis that AT_2R stimulation could represent a novel therapeutic concept in postischemic LV remodeling and HF treatment.

**Sources of Funding**

This work was supported by the EUREKA’s Eurostars program of the European Union and the German Ministry of Research and Technology.

**Disclosures**

B. Dahlöf and T. Unger received speaker fees from Vicore Pharma. T. Unger has a modest interest in Vicore Pharma. U.M. Steckelings received modest research support from Vicore Pharma. B. Dahlöf has a significant interest in Mintgate Scientific, the owner of Vicore Pharma. The other authors report no conflicts.

**References**

Shimizu M, Kario K. Role of the augmentation inde

French B

receptor agonist does not attenuate postmyocardial infarction left ven-
Kemp BA, Carey RM, Kramer CM. A nonpeptide angiotensin II type 2
J
tricular remodeling in mice.

AT2R stimulation prevented fibrosis of peri-infarct myocardium and im-


Shimizu M, Kario K. Role of the augmentation inde


Summary
AT2R stimulation could represent a novel therapeutic concept in postischemic left ventricular remodeling and heart failure treatment.
Angiotensin Type 2 Receptor Stimulation Ameliorates Left Ventricular Fibrosis and Dysfunction via Regulation of Tissue Inhibitor of Matrix Metalloproteinase 1/Matrix Metalloproteinase 9 Axis and Transforming Growth Factor β1 in the Rat Heart

Dilyara Lauer, Svetlana Slavic, Manuela Sommerfeld, Christa Thöne-Reineke, Yuliya Sharkovska, Anders Hallberg, Bjorn Dahlöf, Ulrich Kintscher, Thomas Unger, Ulrike Muscha Steckelings and Elena Kaschina

Hypertension. 2014;63:e60-e67; originally published online December 30, 2013; doi: 10.1161/HYPERTENSIONAHA.113.02522

Hypertension is published by the American Heart Association, 7272 Greenville Avenue, Dallas, TX 75231
Copyright © 2013 American Heart Association, Inc. All rights reserved.
Print ISSN: 0194-911X. Online ISSN: 1524-4563

The online version of this article, along with updated information and services, is located on the World Wide Web at:
http://hyper.ahajournals.org/content/63/3/e60

Data Supplement (unedited) at:
http://hyper.ahajournals.org/content/suppl/2013/12/30/HYPERTENSIONAHA.113.02522.DC1

Permissions: Requests for permissions to reproduce figures, tables, or portions of articles originally published in Hypertension can be obtained via RightsLink, a service of the Copyright Clearance Center, not the Editorial Office. Once the online version of the published article for which permission is being requested is located, click Request Permissions in the middle column of the Web page under Services. Further information about this process is available in the Permissions and Rights Question and Answer document.

Reprints: Information about reprints can be found online at:
http://www.lww.com/reprints

Subscriptions: Information about subscribing to Hypertension is online at:
http://hyper.ahajournals.org//subscriptions/
ONLINE SUPPLEMENT

ANGIOTENSIN TYPE 2 RECEPTOR STIMULATION AMELIORATES LEFT VENTRICULAR FIBROSIS AND DYSFUNCTION VIA REGULATION OF TIMP1/MMP9 AXIS AND TGF-β1 IN THE RAT HEART

Dilyara Lauer1, Svetlana Slavic1,2, Manuela Sommerfeld1, Christa Thöne-Reineke1,3, Yuliya Sharkovska4, Anders Hallberg5, Bjorn Dahlöf6, Ulrich Kintscher1, Thomas Unger1,7, Ulrike Muscha Steckelings1,8*, Elena Kaschina1*

1Center for Cardiovascular Research (CCR) and Institute of Pharmacology, Charité – Universitätsmedizin, Berlin, Germany; 2Institute of Physiology, Pathophysiology and Biophysics, Department of Biomedical Sciences University of Veterinary Medicine, Vienna, Austria; 3Forschungseinrichtung für Experimentelle Medizin, Charité – Universitätsmedizin Berlin, Germany; 4Institute of vegetative Anatomy, Charité – Universitätsmedizin, Berlin, Germany; 5Department of Medicinal Chemistry, Uppsala University, Sweden; 6Sahlgrenska University Hospital, Gothenburg, Sweden; 7CARIM - School for Cardiovascular Diseases Maastricht University, Maastricht, The Netherlands; 81MM – Department of Cardiovascular and Renal Research, University of Southern Denmark, Odense, Denmark

* Both authors contributed equally

Short title: AT2-receptor in post-MI cardiac remodeling

Corresponding author: Elena Kaschina, Center for Cardiovascular Research (CCR) and Institute of Pharmacology, Charité – Universitätsmedizin, Berlin, Hessische Strasse 3-4, D-10115 Berlin, Germany; phone: +49-30-450-525-024; fax: +49-30-450-525-901;

e-mail: elena.kaschina@charite.de.
Expanded Materials and Methods

Animals
Male normotensive Wistar rats (weight 200 to 220g; Harlan Winkelmann, Borchen, Germany) were kept in a specific pathogen-free barrier under standardized conditions with respect to temperature (23±1°C) and humidity and were housed on a 12-hour light/dark cycle in groups of 5 animals with food and water ad libitum.

Myocardial infarction
Under anaesthesia with ketamine/xylazine (80/10 mg/kg i.p.) rats were intubated and connected with a small-animal ventilator with room air at a rate of 75 cycles/min and 3.5mL tidal volume. Left lateral thoracotomy was performed, and a suture was tightened around the proximal left anterior descending coronary artery. Sham-operated rats underwent the same surgical procedure with the exception of the coronary ligature. Analgesic Flunixin-Meglumine (1,0 mg/kg i.p.) was applied 45 min before anesthesia and daily 3 days after operation. The number of animals per group was 12 in C21 and vehicle treated and 8 in sham operated groups at the time of euthanasia. C21 was dissolved in 0,9%NaCl and administered 0,03mg/kg daily i.p.

Western Blot Analysis
Protein lysates were prepared according to standard protocols. Protein expression was normalized to glyceraldehyde-3-phosphate dehydrogenase (GAPDH). Immunoreactive bands were detected by enhanced chemiluminescence (GE Health Care, Germany) and quantified with ImageJ software.

Antibodies for Western Blot analysis

<table>
<thead>
<tr>
<th>Antibodies</th>
<th>Source</th>
<th>Dilution</th>
<th>Manufacturer</th>
</tr>
</thead>
<tbody>
<tr>
<td>MMP-2</td>
<td>Polyclonal rabbit</td>
<td>1:1000</td>
<td>Abcam</td>
</tr>
<tr>
<td>MMP-9</td>
<td>Polyclonal rabbit</td>
<td>1:1000</td>
<td>Abcam</td>
</tr>
<tr>
<td>TIMP-1</td>
<td>Polyclonal rabbit</td>
<td>1:500</td>
<td>Santa Cruz</td>
</tr>
<tr>
<td>TGF-β</td>
<td>Polyclonal rabbit</td>
<td>1:500</td>
<td>Santa Cruz</td>
</tr>
<tr>
<td>GAPDH</td>
<td>Monoclonal mouse</td>
<td>1:500</td>
<td>MerckMillipor</td>
</tr>
</tbody>
</table>

Antibodies for fluorescence microscopy

<table>
<thead>
<tr>
<th>Antibodies</th>
<th>Source</th>
<th>Dilution</th>
<th>Manufacturer</th>
</tr>
</thead>
<tbody>
<tr>
<td>MMP-2</td>
<td>Polyclonal mouse</td>
<td>1:200</td>
<td>Santa Cruz</td>
</tr>
<tr>
<td>MMP-9</td>
<td>Polyclonal mouse</td>
<td>1:200</td>
<td>Santa Cruz</td>
</tr>
<tr>
<td>TIMP-1</td>
<td>Polyclonal rabbit</td>
<td>1:200</td>
<td>Santa Cruz</td>
</tr>
<tr>
<td>TGF-β</td>
<td>Polyclonal rabbit</td>
<td>1:200</td>
<td>Santa Cruz</td>
</tr>
<tr>
<td>anti-mouse Cys3</td>
<td>Polyclonal mouse</td>
<td>1:250</td>
<td>Dianova</td>
</tr>
<tr>
<td>anti-mouse FITC</td>
<td>Polyclonal rabbit</td>
<td>1:250</td>
<td>Dianova</td>
</tr>
<tr>
<td>anti-rabbit</td>
<td>Polyclonal goat</td>
<td>1:200</td>
<td>DAKO</td>
</tr>
</tbody>
</table>
**Immunohistochemistry**

Immunohistochemistry of TGF-β1 was performed in paraffin-embedded tissue sections (5μm) using the avidin-biotin complex method according to the manufacture's instructions (Vectastain ABC; Vector Laboratories, Burlingame, California, USA). Peroxidase activity was visualized by 3-amino-9-ethylcarbazole (AEC, Vector Laboratories). Primary antibodies against TGF-β1 (1:50, Santa Cruz Biotechnology Inc., Heidelberg, Germany) were used.

**mRNA analysis**

Total RNA from LV tissue and CFs was isolated by TRIZOL reagent (Invitrogen) and subsequently treated with DNase-I (Promega). qRT-PCR was performed with MX3005p QPCR System (Stratagene) using SYBR green reaction mix and primers indicated in supplement. All samples were measured in triplicate, and expression values were normalized to 18s rRNA. Data analysis was done using the MxPro™ ET QPCR software (Stratagene) and ΔΔCt-method.

**Primers**

<table>
<thead>
<tr>
<th>Gene (Rat)</th>
<th>Primer sequences for SYBR Green® real-time PCR</th>
</tr>
</thead>
<tbody>
<tr>
<td>18s</td>
<td>Forward: 5’- GGGAGGTAGTGACGAAAAATAAACAT-3’</td>
</tr>
<tr>
<td></td>
<td>Reverse: 5’- TTGCCCTCCAAATGGATCCT-3’</td>
</tr>
<tr>
<td>MMP 2</td>
<td>Forward: 5’- GACGCTGGGAGCATGGAG-3’</td>
</tr>
<tr>
<td></td>
<td>Reverse: 5’- TTACGCGGACCACTTGCTCC-3’</td>
</tr>
<tr>
<td>MMP 9</td>
<td>Forward: 5’- CTTCTCTGGGCGCAAAATG-3’</td>
</tr>
<tr>
<td></td>
<td>Reverse: 5’- CCGGTGACCAGGGTTACCT-3’</td>
</tr>
<tr>
<td>TIMP1</td>
<td>Forward: 5’- TCTGGCATCCTCTTTGTAG-3’</td>
</tr>
<tr>
<td></td>
<td>Reverse: 5’- CACAGCCAGCAGTATAGGTTCTT-3’</td>
</tr>
<tr>
<td>TGF-β1</td>
<td>Forward: 5’- GGTGGACCGCAACAAGCCTATA-3’</td>
</tr>
<tr>
<td></td>
<td>Reverse: 5’- GGGTGGCCACGAGGAGCAAA-3’</td>
</tr>
</tbody>
</table>

**Figures**

**A**

Sham Vehicle C21

LVIDd LVIDs

**B**

Sham Vehicle C21

E A
**Figure S1.** Effect of the C21 treatment on cardiac function 6 weeks post-MI. (A): M-mode echocardiograms of LV. Note LV dilatation, thinning and akinesis of the anterior wall in vehicle-treated rats. Diastolic (LVIDd) and systolic (LVIDs) inner dimensions are increased compared with sham-operated animals. C21 prevented LV dilation and improved anterior wall motion.

(B): Pulsed-wave Doppler spectra of mitral inflow. Note an augmentation of E-wave (early diastolic filling velocity), reduction of A-wave (atrial contraction filling velocity) in vehicle-treated rats vs. healthy rats. C21 reversed MI-induced alterations in E and A-waves almost back to sham level.

![Graph showing myocyte diameter](image)

**Figure S2:** Myocyte diameter was quantified in cardiac sections by means of computer-aided histomorphometry, magnification x60. All of the values are given as mean±SEM; **p<0.01 versus sham group; †††p<0.001 versus vehicle; one-way ANOVA.

![Cross-sections from LV 6 weeks after MI](image)

**Figure S3:** Representative cross-sections from LV 6 weeks after MI. Sirius red stain. Collagen fibers are stained red and cardiac myocytes are yellow, 40x. Note diffuse interstitial fibrosis in vehicle-treated animals which is partly prevented by C21 treatment.
**Figure S4**: (A) Intersitial fibrosis was quantified in Sirius Red-stained cardiac sections by means of computer-aided histomorphometry, magnification x200. Results were calculated as percentage of Sirius-Red-positive area in randomly chosen pictures from cardiac sections. (B) Hydroxyproline concentration in septum 6 weeks after MI (n=3); All of the values are given as mean±SEM; ***p<0.001, **p<0.01 versus sham group; †††p< 0.001, †p<0.05 versus vehicle; one-way ANOVA.

**Figure S5**: (A) MMP2 expression measured in LV 6 weeks post-MI. Western blot analysis (n=6). Representative image and densitometric data of proteins are mean ± SEM; (B): MMP2 mRNA. **p<0.01, *p<0.05 versus sham; †p<0.05 versus vehicle; one-way ANOVA.
Figure S6. Effect of C21 and combination of C21 (1µM) plus PD 123319 (10µM) on cytokine-induced MMP9 and MMP2 secretion in CFs. Activity of MMP9 and MMP2 in CFs determined via gelatine zymography. PD 123319 abolished the effect of C21 on the IL-1 alpha induced MMP9 secretion.

Figure S7. TGF-β1 mRNA expression measured in primary rat cardiac fibroblasts. Bars represent densitometric data of three independent experiments (mean ±S.E.M). C21 reduces IL 1α-stimulated release of TGF-β1 by cardiac fibroblasts. **p<0.01 versus sham; ††p<0.01, †p<0.05 versus vehicle; one-way ANOVA.