Preventing Autoimmunity Protects Against the Development of Hypertension and Renal Injury

Keisa W. Mathis, Kedra Wallace, Elizabeth R. Flynn, Christine Marie-Bilkan, Babbette LaMarca, Michael J. Ryan

Abstract—Several studies suggest a link between autoimmunity and essential hypertension in humans. However, whether autoimmunity can drive the development of hypertension remains unclear. The autoimmune disease systemic lupus erythematosus is characterized by autoantibody production, and the prevalence of hypertension is increased markedly in this patient population compared with normal healthy women. We hypothesized that preventing the development of autoimmunity would prevent the development of hypertension in a mouse model of lupus. Female lupus (NZBWF1) and control mice (NZW) were treated weekly with anti-CD20 or immunoglobulin G antibodies (both 10 mg/kg, IV) starting at 20 weeks of age for 14 weeks. Anti-CD20 therapy markedly attenuated lupus disease progression as evidenced by reduced CD45R+ B cells and lower double-stranded DNA autoantibody activity. In addition, renal injury in the form of urinary albumin, glomerulosclerosis, and tubulointerstitial fibrosis, as well as tubular injury (indicated by renal cortical expression of neutrophil gelatinase-associated lipocalin) was prevented by anti-CD20 therapy in lupus mice. Finally, lupus mice treated with anti-CD20 antibody did not develop hypertension. The protection against the development of hypertension was associated with lower renal cortical tumor necrosis factor-α expression, a cytokine that has been previously reported by us to contribute to the hypertension in this model, as well as renal cortical monocyte chemoattractant protein-1 expression and circulating T cells. These data suggest that the development of autoimmunity and the resultant increase in renal inflammation are an important underlying factor in the prevalent hypertension that occurs during systemic lupus erythematosus. (Hypertension. 2014;64:792-800.) ● Online Data Supplement

Key Words: autoantibodies • B-lymphocytes • pressure • T-lymphocytes

Humoral immune system activation has a central role in the pathogenesis of autoimmune disorders, in part, by promoting antibody-mediated immune complex formation, leading to tissue injury and inflammation.1 Several studies show an association between the production of antibodies characteristic of autoimmune disease and human hypertension. For example, work from Kristensen et al2 showed that patients with hypertension were more likely to have increased circulating autoantibodies of the immunoglobulin G (IgG) and immunoglobulin M class.3 Similarly, Gudbrandsson et al4 showed high levels of antinuclear antibodies in patients with malignant hypertension.5 More recently, reports of specific activating antibodies that could promote hypertension have been reported.5,6 Although there is clear and growing evidence for humoral immune system activation in human hypertension, whether a general loss of immune tolerance leading to systemic autoimmunity can be an underlying causative factor in the pathogenesis of hypertension has not been directly tested.

Systemic lupus erythematosus (SLE), a chronic autoimmune disorder that predominantly affects women of childbearing age, is characterized by circulating antinuclear autoantibodies (eg, anti–double-stranded DNA antibodies) and is associated with a markedly increased prevalence of hypertension.7 Using an established female mouse model of SLE that displays these same characteristics, our laboratory has previously reported on several factors that contribute to the hypertension during SLE including endothelial dysfunction,8 impaired renal hemodynamic function,9,10 inflammatory cytokines,11 and oxidative stress.12 In the present study, we tested the hypothesis that the development of autoimmune disease is the underlying factor that leads to the prevalent hypertension associated with SLE. Our data show that administration of a mouse anti-CD20 antibody (the equivalent of rituximab in humans) to deplete B cells markedly attenuates autoantibody production and prevents the development of hypertension in a mouse model of SLE.

Methods

Animals
Twenty-week-old female NZBWF1 mice (Jackson Laboratories, Bar Harbor, ME), an established model that mimics characteristics of SLE...
in humans, and NZW/LacJ mice (controls; Jackson Laboratories) were used in this study because we previously showed that this age precedes the development of autoantibodies, albuminuria, and hypertension. Mice were maintained on a 12-hour light/dark cycle in temperature-controlled rooms with access to food and water ad libitum. All studies were approved by the University of Mississippi Medical Center Institutional Animal Care and Use Committee and were in accordance with National Institutes of Health Guide for the Care and Use of Laboratory Animals.

**Antibody Administration**
Starting at 20 weeks of age, mice were administered 100 μL of a monoclonal antibody to mouse CD20 (anti-CD20; provided by Genentech, South San Francisco, CA) or IgG (isotype control also provided by Genentech) retroorbitally at a dose of 10 mg/kg once per week for 14 weeks.

Treatment efficacy was assessed by measuring spleen weight and the percentage of CD45R+ B cells in freshly prepared splenocytes using flow-assisted cell sorting (fluorescence-activated cell sorter) as previously described.15

**Blood Pressure**
Blood pressure was recorded at the end of the 14-week protocol in conscious freely moving mice as previously reported by our laboratory.3,9,10,12,14

**Autoantibodies**
Plasma levels of anti–double-stranded DNA antibodies, a clinical hallmark of SLE, were measured at week 34 as previously described.9,10,12,14,16 and are presented as a positive antibody activity index per the manufacturer’s instructions and as previously published.15 An antibody activity <1 is considered negative, whereas activity >1 is considered positive.

**Renal Inflammation, Oxidative Stress, and Injury**
Markers of renal cortical inflammation, including tumor necrosis factor (TNF)-α and monocyte chemotactic protein (MCP)-1, were measured using Western blot as previously described by our laboratory in control mice and SLE mice with evidence of active disease (autoantibodies and albuminuria).12,14 Renal cortical expression of catalase, copper-zinc superoxide dismutase, and glutathione peroxidase was measured using Western blot as previously described.15

Albuminuria was determined in 24-hour urine samples either weekly using a dipstick assay or at the conclusion of the study by ELISA (presented as excretion rate in μg/d) as previously described.3,9,10,12,14,16 Paraffin-embedded kidneys were stained with periodic acid Schiff to assess glomerulosclerosis as previously described17 and Masson’s trichrome staining to assess tubulointerstitial fibrosis. Renal cortical expression of neutrophil gelatinase-associated lipocalin was measured as an index of renal tubular injury as previously reported by our laboratory.14

**Statistical Analysis**
Data are presented as mean±SEM. Statistical analyses were performed using SigmaPlot 11.0 software (Systat, San Jose, CA). Two-way ANOVA was used followed by the appropriate post hoc test to determine differences between multiple groups. Data were considered statistically different when P<0.05.

**Results**

**Anti-CD20 Therapy Attenuates the Development of Hypertension in Female Mice With SLE**
To determine whether autoimmunity contributes to the pathogenesis of hypertension during SLE, mean arterial pressure was measured in mice treated with vehicle (IgG) or anti-CD20 antibody. Mean arterial pressure was increased in SLE mice compared with controls (142±6 versus 120±3 mm Hg; P<0.001; Figure 1). Anti-CD20 treatment starting at 20 weeks of age prevented the increase in blood pressure in SLE mice (126±2 mm Hg; P<0.01) but had no effect in controls (121±3 mm Hg). Before starting the studies at 20 weeks of age, preliminary experiments were conducted beginning treatment at 26 weeks (8 total weeks of anti-CD20 antibody) and 30 weeks of age (4 total weeks of anti-CD20 antibody). The efficacy of the treatment corresponded with the length and timing of the treatment (please see online-only Data Supplement).

**Anti-CD20 Therapy Depletes B Cells and Attenuates the Development of SLE**
Anti-CD20 therapy reduced spleen weight in control (P<0.03) and SLE mice (P<0.001), as well as splenocyte count in control (P<0.001) and SLE mice (P<0.001; data not shown). SLE mice had increased B cells compared with controls, as indicated by a significant group effect on the percentage of splenic CD45R+ B cells (P<0.03; Figure 2A). Anti-CD20 therapy effectively depleted splenic B cells in control and SLE mice (treatment effect; P<0.01). Importantly, the percentage of splenic B cells in SLE mice after 14 weeks of anti-CD20 therapy (13.3±2%) was equivalent to control, vehicle-treated animals (11.4±4.6%). Anti–double-stranded DNA autoantibody activity was significantly elevated in SLE mice (4.51±0.78; P<0.05) compared with controls (0.52±0.08), as previously reported by our laboratory (Figure 2B).12,14,15 B-cell depletion did not significantly alter activity of anti–double-stranded DNA autoantibodies in control mice (0.18±0.08); however, autoantibody activity was attenuated by B-cell depletion in SLE mice (2.14±0.69; P<0.005). These data provide important functional evidence supporting the effective depletion of B cells achieved with 14 weeks of anti-CD20 therapy. Although the percentage of circulating CD3+ (pan) T cells was not different between control and SLE mice (21.5±8.5 and 20.5±7.1%, respectively), there was a significant treatment effect of anti-CD20 therapy on CD3+ T cells (Figure 3; P<0.01).

**Figure 1.** Mean arterial pressure measured in control and systemic lupus erythematosus (SLE) mice administered immunoglobulin G (IgG) or anti-CD20 for 14 weeks (n indicated on graph). *P<0.001 vs control/IgG; +P<0.01 vs SLE/IgG.
Anti-CD20 Therapy Attenuates the Development of Renal Inflammation in SLE Mice

Previous work from our laboratory demonstrated that TNF-α mechanistically contributes to the hypertension during SLE. In the present study, the impact of attenuating the development of autoimmunity on renal TNF-α protein expression was examined. Renal cortical expression of TNF-α was increased in vehicle-treated SLE mice compared with vehicle-treated controls (2.2±0.30 versus 0.81±0.13; P<0.005; Figure 4A). SLE mice treated with anti-CD20 antibody had significantly lower TNF-α (0.54±0.07; P<0.01) compared with vehicle-treated SLE mice. The treatment did not impact expression in control mice (0.27±0.03; P=0.103). Similarly, renal cortical expression of MCP-1 was assessed and was increased in SLE mice compared with controls (0.047±0.033 versus 0.006±0.002; P<0.005; Figure 4B). SLE mice treated with anti-CD20 antibody had lower MCP-1 (0.010±0.006; P<0.01), but expression was not altered in controls (0.013±0.006).

Expression of Antioxidant Enzymes Is Increased in Anti-CD20–Treated SLE Mice

We previously reported that oxidative stress contributes to hypertension during SLE. Therefore, renal cortical expression of antioxidant enzymes was assessed. Catalase was lower in SLE mice compared with controls (0.039±0.008 versus 0.058±0.004; P<0.03; Figure 5A). Catalase expression was increased in the kidneys of SLE mice treated with anti-CD20 antibodies (0.055±0.003; P<0.02). Renal copper-zinc superoxide dismutase was similar in control and SLE mice (0.054±0.005 and 0.040±0.012, respectively; Figure 5B); however, expression was increased in SLE mice treated with anti-CD20 antibodies (0.079±0.005; P<0.02). Protein expression of glutathione peroxidase in the renal cortex was not statistically different among all groups (Figure 5C).

Anti-CD20 Therapy Protects the Kidney in Mice With SLE

To test whether B cell depletion and the associated reduction in autoantibodies protected against the development of renal disease common to this model, several indices of renal injury were assessed. The prevalence of albuminuria in vehicle-treated SLE mice reached ≥67% by 34 weeks, whereas the prevalence of albuminuria in SLE mice treated with anti-CD20, as well as treated and untreated controls, remained <23% throughout the entire course of the 14-week study (Figure 6A). Urinary albumin excretion rate, measured...
by ELISA at the conclusion of the study, was increased in SLE mice compared with controls (833±183 versus 70±15 μg/d; \(P<0.001\); Figure 6B) as previously shown and was lower in anti-CD20–treated SLE mice (85±27; \(P<0.001\)). Glomerulosclerosis, defined as mesangial expansion and extracellular matrix deposition, was increased in SLE mice compared with controls (1.16±0.35 versus 0.23±0.03; \(P<0.001\); Figure 6C). Anti-CD20 therapy prevented the development of glomerulosclerosis in SLE mice (0.20±0.04; \(P<0.001\)). Similarly tubulointerstitial fibrosis, evident in both the renal cortex and outer medulla, was increased in SLE mice compared with controls (1.46±0.49 versus 0.28±0.08; \(P<0.001\); Figure 6D) as indicated by increased collagen deposition and tubular dilatation and atrophy and was prevented in anti-CD20–treated SLE mice (0.38±0.49; \(P<0.02\)). Representative histological sections for both glomerulosclerosis and tubulointerstitial fibrosis in SLE mice treated with IgG or anti-CD20 are shown in Figure 6E. Renal cortical neutrophil gelatinase-associated lipocalin, a marker of tubular injury, was increased in mice with SLE (Figure 6F), and this was prevented in anti-CD20–treated animals.

**Discussion**

Based on human studies showing a positive correlation between essential hypertension and serum levels of autoantibodies characteristic of systemic autoimmune disease,
the present study was designed to test whether the development of autoimmune disease can be an underlying factor in the pathogenesis of hypertension. To test this, a mouse anti-CD20 antibody, the equivalent of rituximab used in humans, was administered to an established female mouse model that develops SLE and hypertension. The major new findings of this study are that (1) treatment with an anti-CD20 antibody prevents the development of hypertension associated with SLE; (2) renal TNF-\(\alpha\), a cytokine that contributes to the development of hypertension in this model, is attenuated in
anti-CD20–treated animals, as is the expression of MCP-1; and (3) the expression of protective antioxidant enzymes is increased in the kidney of anti-CD20–treated SLE mice. In addition, the findings of this study support previous work showing that anti-CD20 therapy prevents the development of renal injury (albuminuria, glomerulosclerosis, and tubulointerstitial fibrosis) that is commonly associated with SLE disease progression. Therefore, these data provide an important proof of concept that autoimmunity may be an important factor underlying the development of hypertension, in part, through mechanisms that promote local renal inflammation and oxidative stress.

Autoimmunity Is Associated With Hypertension

Autoimmunity has long been implicated in the pathogenesis of hypertension. For example, an early study by White and Grollman demonstrated that vascular and renal antigens were associated with experimental hypertension in a renal infarction model, leading them to postulate that autoimmunity was an underlying factor. More recently, the idea that autoantigens perpetuate hypertension has been significantly advanced. Harrison and colleagues have proposed the concept that physiological stressors, including high angiotensin II, lead to the release of local antigens that ultimately promote adaptive immune system activation to sustain the hypertension.

In addition, Rodriguez-Iturbe and colleagues provided compelling evidence that heat shock protein 70 is an important antigen that promotes adaptive immune system activation and contributes to the development of salt-sensitive hypertension. The present study makes an important advance as it directly tests whether preventing autoimmunity can prevent the development of hypertension.

The association between circulating autoantibodies and hypertension has been recognized for many years with studies showing that IgG and anti-immunoglobulin M, antinuclear, and antiphospholipid antibodies are increased in patients with essential and malignant hypertension. The role of specific activating antibodies has also been reported in patients with hypertension. For example, activating angiotensin type 1 receptor antibodies are linked with preeclampsia while α1-adrenergic receptor antibodies have been reported in several studies of human hypertension. When patients with hypertension with α1-adrenergic autoantibodies were subjected to immunoadsorption therapy to remove those antibodies, blood pressure was reduced. As a whole, these data suggest that humoral immune system activation can promote the pathogenesis of hypertension either through specific activating antibodies or more generalized antibodies that characterize systemic autoimmune disease; however, whether autoimmunity per se is an underlying cause of hypertension remains uncertain. The current study provides evidence that humoral immune system activation associated with systemic autoimmune disease is an important factor in the pathogenesis of hypertension and thus addresses this previously unanswered question.

B-Cell Depletion During SLE

It is widely recognized that humoral immune system activation plays a central role in the pathogenesis of SLE. This is based on evidence that B cells, which differentiate into antibody-producing plasma cells, are increased during SLE. The production and release of autoantibodies contribute to immune complex formation, and their deposition into tissues initiates an inflammatory response that can lead to tissue injury and inflammation. The data from the current study show that anti-CD20 treatment in female SLE mice prevented the increase in B cells so that they remained similar to the vehicle-treated control animals that do not develop disease. Evidence of functional inhibition is also provided, given that antibody activity is significantly attenuated in SLE mice treated with anti-CD20 antibody. Therefore, these data confirm the effectiveness of systemic delivery of the antibody to suppress B-cell activity and provide a compelling case that we effectively prevented or delayed the onset of autoimmunity.

Because of their role in the development of SLE, B cells have been targeted for the treatment of SLE with therapeutics like rituximab (anti-CD20 antibody). However, the efficacy of rituximab to achieve the clinically desired outcome (ie, remission, antibody reduction, reduced renal injury) has been variable. The variable efficacy could be related to the fact that CD20 is not expressed on the antibody-producing plasma cells or attributed to differing severity or stages of disease in patients with SLE and their medications (ie, mycophenolate mofetil, azathioprine, prednisolone) at the time of the studies. In pilot studies, we administered anti-CD20 antibody to control and SLE mice for 4 and 8 weeks, starting at 30 and 26 weeks of age, respectively, an age in which autoantibodies are already present and active in SLE mice. The results show that 4 weeks of treatment starting at a later age did not affect blood pressure, and 8 weeks of treatment only modestly reduced blood pressure (please see online-only Data Supplement). This suggests that once the inflammatory process has begun and antibodies are being produced, the efficacy of B-cell depletion by anti-CD20 antibody may be limited. Importantly, these results are consistent with previous work showing a greater variability in the efficacy of anti-CD20 treatment as mice with SLE age and suggest that beginning treatment earlier may be more effective in controlling blood pressure and renal injury. In future studies, it will be important to determine whether specific B-cell populations are essential for autoimmune-mediated hypertension to occur.

Potential Mechanisms for Attenuated Hypertension After Prevention of Autoimmunity

The data herein suggest that the protection from developing hypertension afforded by preventing autoimmunity is dependent on reductions in renal inflammation, renal oxidative stress, and renal injury. Our laboratory previously reported that expression of the proinflammatory cytokine TNF-α is increased in the renal cortex from mice with SLE and that blockade of TNF-α biological activity with etanercept attenuates the hypertension in this model. The present study confirms that renal cortical TNF-α protein expression is increased in vehicle-treated SLE mice and demonstrates that this is completely prevented in the SLE mice treated with anti-CD20 antibody. Potential mechanisms by which TNF-α could regulate blood pressure are likely dependent on the balance between pro- and antihypertensive actions of TNF-α. For example, TNF-α promotes renal vasoconstriction, a prohypertensive
effect, whereas it has also been reported to have natriuretic effects through inhibition of the Na\(^+\)-K\(^-\)-2Cl\(^-\)-cotransporter.\(^{37}\)

Although the source of the renal TNF-\(\alpha\) in this model of SLE was not directly determined, it is likely that it comes from infiltrating immune cells such as macrophage and T cells, which have been reported by us and others to be elevated in the kidneys of these mice.\(^{11,17}\) We previously reported that renal MCP-1 expression, a chemokine that attracts monocytes and T cells, is increased in mice with SLE.\(^{14,17}\) Therefore, renal cortical protein expression of MCP-1 was assessed in the present study. The results show that MCP-1 expression is attenuated in SLE mice treated with anti-CD20 antibody, consistent with the concept that infiltrating immune cells in the kidney likely contribute to the increased TNF-\(\alpha\).

Inflammatory cytokines, including TNF-\(\alpha\), are associated with promoting oxidative stress. We previously demonstrated that TNF-\(\alpha\) contributes to renal oxidative stress in this model\(^{11}\) and that oxidative stress has a mechanistic role in the development of hypertension during SLE.\(^{12}\) In the current study, SLE mice treated with anti-CD20 antibodies exhibited increased expression of antioxidants enzymes, suggesting that improved regulation of reactive oxygen species may be an important protective mechanism as well.

By preventing the development of autoimmunity, several indices of renal injury (albuminuria, neutrophil gelatinase-associated lipocalin, glomerulosclerosis, and fibrosis) were markedly blunted in the SLE animals. This is consistent with the renal protective actions of anti-CD20 antibody therapy in the NZBWF1 model reported previously by others.\(^{34}\) Although it cannot be completely ruled out that the prevention of renal injury results from the prevention of the hypertension, we think that it is unlikely to be the case. The NZBWF1 model is a widely established experimental model of immune complex-mediated glomerulonephritis. Among the most commonly studied models of lupus nephritis in mice (ie, MRL/lpr and BXSB), only the NZBWF1 mouse develops hypertension, whereas all 3 strains develop lupus nephritis. In addition, we recently reported that the hypertension and renal injury in the NZBWF1 model mice are unrelated,\(^{14,15}\) thus supporting the concept that the renal injury is not merely pressure dependent.

Finally, it is important to recognize the possibility that T cells may have a mechanistic role in the pathogenesis of hypertension during SLE. T cells not only play a critical role in promoting antibody production through T-cell help but also mechanistically contribute to the progression of angiotensin II and salt-sensitive hypertension.\(^{38-42}\) Therefore, the data showing that circulating T cells are depleted in anti-CD20–treated mice suggests that this may have a mechanistic role in preventing the autoimmunity and the associated hypertension. Interestingly, similar effects of rituximab therapy on T cells have been demonstrated in humans and experimental models. In 1 study, peripheral blood mononuclear cells were isolated from the blood of newly diagnosed patients with non-Hodgkin lymphoma treated with rituximab as part of their therapy. In the absence of B cells, T-cell activation was reduced, most likely resulting from reduced antigen presentation after depletion of the antigen-presenting B cells.\(^{43}\) In a mouse model of experimental autoimmune encephalomyelitis with a humanized immune system, rituximab decreased CD4\(^+\) T cells.\(^{44}\) Consistent with our findings, mouse anti-CD20 (the mouse equivalent to rituximab) decreased activated T cells in the NZBWF1 mouse.\(^{44}\) These data support the overall hypothesis that autoimmunity contributes to the development of hypertension by promoting tissue inflammation and that humoral immune system activation is an essential component.

**Perspectives**

The work presented here makes significant advances on 2 important clinically relevant topics. During the past several years, it has become clear that adaptive immune system activation plays an important role in the development and maintenance of hypertension. Much of that work has focused on T cells in experimental hypertension and has implicated a role for autoimmunity in the pathogenesis of hypertension. In addition, studies in humans suggest that systemic autoimmunity is associated with essential hypertension. Therefore, the current study provides evidence that autoimmunity mechanistically contributes to the development of hypertension. The second advance is more directly related to SLE, a chronic autoimmune inflammatory disorder with prevalent hypertension for reasons that continue to be elucidated. The data provide important evidence that the hypertension associated with SLE is driven by humoral immune system activation that ultimately leads to renal inflammation and injury. The direct impact of immunosuppressive therapy on renal hemodynamic function has not been widely studied in SLE and will be an important consideration in future experiments. Overall, the current study may have important clinical implications as well. For example, immunosuppressive therapy would be ill-advised for patients with uncomplicated hypertension; however, our data suggest that immunosuppression may have an added benefit of helping to control blood pressure in patients with chronic inflammatory diseases. This idea is supported by a small clinical study showing that mycophenolate mofetil treatment reduces blood pressure in patients with hypertension with psoriasis or rheumatoid arthritis.\(^{45}\)

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**Disclosures**

None.

**References**


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**What Is New?**
- Preventing the development of systemic autoimmune disease attenuates the development of hypertension.

**What Is Relevant?**
- These studies demonstrate the potentially important underlying role that autoimmunity can have in the pathogenesis of hypertension.
- These studies advance the concept that immunosuppressive therapies may contribute to blood pressure control in patients with hypertension with chronic inflammatory disease.
- Because hypertension is a multifactorial disease with growing evidence of a role for autoimmunity and adaptive immune system activation, the use of experimental models with autoimmunity and hypertension may be particularly informative for better understanding the pathogenesis of the disease.

**Summary**

There is clear evidence supporting an association between autoimmunity and hypertension; however, whether systemic autoimmunity is mechanistically important remains unclear. Systemic lupus erythematosus is an autoimmune disease driven by loss of host tolerance, leading to humoral immune system activation and is associated with prevalent hypertension and renal disease. The current study set out to determine whether preventing autoimmune disease protects against the development of hypertension in an established female mouse model of systemic lupus erythematosus. The results show that chronic B-cell depletion attenuated the development of hypertension, renal injury, and renal inflammation and support the concept that autoimmune mechanisms can underlie the pathogenesis of hypertension.
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PREVENTING AUTOIMMUNITY PROTECTS AGAINST THE DEVELOPMENT OF HYPERTENSION AND RENAL INJURY

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METHODS

**Animals:** Female NZBWF1 mice, an established model that mimics characteristics of SLE in humans, and NZW/LacJ mice (controls) were obtained from Jackson Laboratories (Bar Harbor, ME). At 20 weeks of age, mice were randomly divided into four groups (n = 8-9 animals per group): control mice administered vehicle (Control/IgG), control mice subjected to B cell depletion (Control/Anti-CD20), SLE mice administered vehicle (SLE/IgG), and SLE mice subjected to B cell depletion (SLE/Anti-CD20). This study was designed to test whether anti-CD20 therapy can prevent the development of autoimmunity and the associated hypertension. Therefore, twenty-week-old SLE mice were used because we previously showed that this age precedes the development of autoantibodies, albuminuria and hypertension. Mice were maintained on a 12-hour light/dark cycle in temperature-controlled rooms with access to food and water *ad libitum*. All studies were approved by the University of Mississippi Medical Center Institutional Animal Care and Use Committee (IACUC) and were in accordance with National Institutes of Health (NIH) Guide for the Care and Use of Laboratory Animals.

**Antibody Administration:** Starting at either 20, 26, or 30 weeks of age, mice were administered 100 \( \mu \text{L} \) of a monoclonal antibody to mouse CD20 (anti-CD20; provided by Genentech, South San Francisco, CA) or IgG (the isotype control also provided by Genetech) retro-orbitally at a dose of 10 mg/kg once per week until the mice reached 34 weeks of age.

Treatment efficacy was assessed by measuring spleen weight and the percentage of CD45R\(^+\) B cells in freshly prepared splenocytes using flow-assisted cell sorting (FACS) as previously described. At the time of tissue harvest, splenocytes were isolated and washed in 10 mL of buffer consisting of RPMI and streptomycin/penicillin following red blood cell lysis. Splenocytes were labeled for 30 minutes at 4°C with an antibody against mouse CD45R conjugated to fluorescein isothiocyanate (FITC; Invitrogen, Frederick, MD). In addition, the percentage of CD3\(^+\) T cells was measured in freshly prepared peripheral blood mononuclear cells by labeling leukocytes with an antibody against mouse CD3 conjugated to phycoerythrin (PE; BD Biosciences). Following incubation, cells were washed and resuspended in 500\( \mu \text{l} \) of RPMI and analyzed by flow cytometry using a BD Gallios flow cytometer (BD Biosciences).
Figure S1: A) Mean arterial pressure measured in control and SLE mice administered IgG or anti-CD20 for 4 weeks (n indicated on graph). *p<0.05 vs. corresponding Control; and B) Mean arterial pressure measured in control and SLE mice administered IgG or anti-CD20 for 8 weeks (n indicated on graph). *p<0.05 vs. Control/IgG.
Figure S2: A) Absolute values of splenic CD45R+ B cells (cells/spleen) in control and SLE mice administered IgG or anti-CD20 for 14 weeks (n indicated on graph). *p<0.05 vs. corresponding IgG; and B) Absolute values of circulating CD3+ T cells (cells/mL) in control and SLE mice administered IgG or anti-CD20 for 14 weeks (n indicated on graph). *p<0.05 vs. Control/IgG.