Heart Rate Variability

Biophysical Characterization of the Underappreciated and Important Relationship Between Heart Rate Variability and Heart Rate

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Abstract—Heart rate (HR) variability (HRV; beat-to-beat changes in the R-wave to R-wave interval) has attracted considerable attention during the past 30+ years (PubMed currently lists >17000 publications). Clinically, a decrease in HRV is correlated to higher morbidity and mortality in diverse conditions, from heart disease to fetal distress. It is usually attributed to fluctuation in cardiac autonomic nerve activity. We calculated HRV parameters from a variety of cardiac preparations (including humans, living animals, Langendorff-perfused heart, and single sinoatrial nodal cell) in diverse species, combining this with data from previously published articles. We show that regardless of conditions, there is a universal exponential decay-like relationship between HRV and HR. Using 2 biophysical models, we develop a theory for this and confirm that HRV is primarily dependent on HR and cannot be used in any simple way to assess autonomic nerve activity to the heart. We suggest that the correlation between a change in HRV and altered morbidity and mortality is substantially attributable to the concurrent change in HR. This calls for re-evaluation of the findings from many articles that have not adjusted properly or at all for HR differences when comparing HRV in multiple circumstances. (Hypertension. 2014;64:1334-1343.) ● Online Data Supplement

Key Words: autonomic nervous system ■ ion channels ■ physiology ■ sinoatrial node

Early scientists thought that the heart beat was metronomic until in 1733, Reverend Stephen Hales1 made the observation that the pulse rate varied with respiration. In the 1960s, advances in ECG recording allowed quantification of beat-to-beat variation in to R-wave interval (R-R interval) dubbed heart rate (HR) variability (HRV).2 Thereafter, an explosion of investigation into HRV occurred—PubMed currently lists >17000 HRV-related articles. Diverse articles describing different characteristics of HRV have been proposed.3 In the general population, having a low HRV is associated with increased morbidity and mortality from various causes, not all cardiac.4 For example, Dekker et al5 demonstrated that decreased HRV was a predictor of death from all causes, including cancer. There is ample evidence too of changes in HRV that occur in response to disease, both cardiac and noncardiac. For example, decreased HRV is correlated to higher morbidity and mortality in patients following myocardial infarction.6 In addition to ischemic heart disease, significant changes in HRV are also documented in many other common conditions, including heart failure, hypertension, before arrhythmia onset, left ventricular hypertrophy, hypertrophic cardiomyopathy, and in noncardiac conditions, including sepsis, fetal distress, diabetes mellitus, stroke, depression, and obstructive airways disease (for review, see Billman7). HRV was heralded as a useful noninvasive method for predicting clinical risk in these diverse disease states.

Underlying HRV is thought to be fluctuating behavior in the limbs of the cardiac autonomic nervous system. Before the advent of HRV, the ability of scientists and physicians to noninvasively estimate cardiac autonomic innervation was limited. A significant literature concerning HRV from bench to bedside continues to be produced today.

In this study, we have investigated HRV in a variety of species and cardiac preparations. Our results argue that, rather than being a pure marker of cardiac autonomic nervous system activity, HRV is primarily dependent on HR, with HRV increasing when the R-R interval increases (ie, when HR slows).
and decreasing when the R-R interval decreases (ie, when HR quickens). This lends weight to the theory that the correlation between a decrease in HRV and higher morbidity and mortality is the consequence of the concurrent increase in HR. It follows that observed differences in HRV between ≥2 scenarios should, therefore, always take into account the HR present when HRV was measured, otherwise conclusions drawn may be flawed.

**Methods**

Experiments performed for this study focused on 3 different cardiac preparations: the conscious in vivo human and rat, isolated denervated Langendorff-perfused heart (rabbit, rat), and isolated sinoatrial node cell (SANC, rabbit). The species used in each of these situations varied, as did the experimental conditions used, and these are described in the online-only Data Supplement. Human studies were approved by the local ethics committee at the Manchester Royal Infirmary, and subjects gave their informed consent. Studies on the rat in vivo were approved by the Norwegian Council for Animal Research; the protocol used had the ID number 1980 and was performed in accordance with the Guide for the Care and Use of Laboratory Animals by the European Commission Directive 86/609/EEC. Studies on the Langendorff-perfused rabbit and rat heart were performed in accordance with the Animals (Scientific Procedures) Act 1986 from the UK Home Office, with animals being euthanized using approved Schedule 1 procedures. Studies on the isolated SANC from the rabbit were conducted in accordance with the National Institutes of Health Guide for the Care and Use of Laboratory Animals.

**HRV Analysis**

Continuous recordings of single-channel ECG data from the conscious human or rat and the isolated rabbit and rat Langendorff-perfused heart and spontaneous action potentials recorded from the isolated rabbit SANC were subjected to HRV analysis in multiple domains along identical lines, as detailed in the online-only Data Supplement. Stationary time epochs of 2.5 minutes (150 s) were used. Because of the varying HRs in different species, the number of data points differed among species. We repeated the data analysis with a fixed number of data points (500 heart beats) and did not observe significant differences in the results compared with the analysis using fixed time epochs of 2.5 minutes (data not shown). For simplicity, only the SD of normal beat to normal beat intervals (SDNN) and root mean square of successive differences (RMSSD) are presented as parameters reflecting HRV herein.

**Computer Modeling**

The deterministic ordinary differential equation model for the electrophysiology of a rabbit central SANC by Zhang et al was implemented in this study for the advanced computer modeling. Further details of the model are given in the online-only Data Supplement.

**Results**

We measured baseline HRV in the conscious human (n=11) and rat (n=11), isolated denervated Langendorff-perfused heart from the rabbit (n=58) and rat (n=8), and isolated denervated rabbit SANC (n=67). Figure 1A to 1E shows tachograms for the different preparations, demonstrating marked differences in HRV under baseline conditions; the data are summarized in Figure 1F to 1H. Corresponding power spectra and total power summary data are shown in Figure S1 in the online-only Data Supplement. HRV as a result of fluctuation in autonomic nerve activity is expected to be present in the conscious animal, but absent in the isolated denervated preparations; however, the pattern of HRV did not conform to this assumption. As expected, HRV in terms of SDNN, RMSSD, and total power was high in the conscious human (Figure 1A, 1G, and 1H and Figure S1) and low in the isolated denervated preparations (Figure 1B–1D, 1G, and 1H and Figure S1). However, in the conscious rat with an intact autonomic nervous system, HRV was also low (Figure 1E, 1G, and 1H and Figure S1). The baseline cycle length (CL; same as R-R or NN interval) also varied widely between preparations (Figure 1F).

The preparation with the longest CL was the conscious human (mean±SEM, 839±50 ms). The Langendorff-perfused rabbit heart (428±10 ms) and rabbit SANC (327±7 ms) had the next-longest CLs, followed by the Langendorff-perfused rat heart (229±8 ms). The preparation with the shortest CL was the conscious rat (161±4 ms). Below we argue that HRV is strongly dependent on CL: the shorter the CL, the less the HRV. This explains the low HRV in the conscious rat, which had the shortest CL (Figure 1E and 1F). However, there is not a perfect correspondence between HRV and CL: the HRV in the 2 Langendorff heart preparations is lower than expected on the basis of CL alone. We re-examine the differences in baseline HRV in the 5 preparations in Figure 5A.

Despite clear interpreparation differences in HRV, β-adrenergic stimulation has the same effect on HRV in the conscious animal, isolated denervated heart, and isolated denervated SANC: Figure 2A shows the effect of the β-adrenergic agonist, dobutamine, on the CL and HRV in the conscious human (n=11). As expected, dobutamine caused a dramatic decrease in the CL (Figure 2A and 2E) and along with it a decrease in measured parameters of HRV (Figure 2F and 2G). The effect of β-adrenergic stimulation (using isoprenaline as the agonist) was also investigated in the rabbit SANC (Figure 2B) and in the Langendorff-perfused rabbit (Figure 2C) and rat (Figure 2D) heart. β-Adrenergic stimulation had a similar effect on CL, causing it to significantly shorten in all preparations studied (Figure 2E). It also had a similar effect on HRV, causing it to decrease in all preparations studied (Figure 2F and 2G). Could the decrease in HRV in all preparations simply be accounted for by the decrease in CL?

**Relationship Between HRV and HR**

Figure 3A summarizes all data obtained from the conscious human and rat, the Langendorff-perfused rabbit and rat heart, and the rabbit SANC: SDNN is used as the parameter of HRV and is plotted against HR. Figure 3B shows the same data, but only up to HRs of 240 bpm. Figure 3A and 3B also shows data from many other studies, including data from the healthy conscious human, athletically trained conscious human, conscious human exposed to autonomic blockade, conscious human with heart failure, conscious human with hypertrophic cardiomyopathy, conscious human with myocardial infarction, conscious human heart transplant recipient, conscious mouse (wild type and transgenic), and rabbit SANC (control and exposed to acetylcholine)—see online-only Data Supplement for details of studies included. Despite the fact that the SDNN (along with other HRV parameters) will have been measured in different ways (over different sampling time periods, for example) and that the data are from different species, different preparations, different disease states, different conditions, and different laboratories, all the data are clustered and approximately fall along a common exponential decay-like
curve: there is an exponential decay-like decrease in SDNN when HR increases (Figure 3A and 3B). Figure S2 shows that subgroups of the data in Figure 3A and 3B show the same relationship between HRV and HR as the whole data (Figure 3A and 3B). Figure 3C shows a semilogarithmic plot of the same data: the natural logarithm of the SDNN is plotted against the HR and there is a linear relationship, as is expected for an exponential decay-like process; it can be calculated from this linear relationship that for every 10 bpm increase in HR, $\log_{e}(\text{SDNN})$ decreases by 0.169 ms ($\text{SEE}=0.41$; $R^2=0.68$). It follows that the SDNN at a given HR is given by the following:

$$\text{SDNN} = \text{SDNN}_{\text{reference}} e^{\frac{(HR-HR_{\text{reference}})}{55.5}} \quad (1)$$

where $\text{SDNN}_{\text{reference}}$ is the SDNN at a reference HR, $HR_{\text{reference}}$.

**A Simple Mathematical Model of the Relationship Between HRV and HR**

In the sinoatrial node, the membrane depolarizes throughout diastole (the pacemaker potential). If the pacemaker potential is monophasic, the rate of change of membrane potential, $\frac{dV_m}{dt}$, is roughly constant. This is driven by a roughly
constant inward ionic current, $I_{\text{tot}}$, the total of the various ionic currents flowing during diastole, including rapid and slow delayed rectifier $K^+$ currents ($I_{Kr}$ and $I_{Ks}$), funny current ($I_f$), background $Na^+$ current ($I_{b,Na}$), $T$-type $Ca^{2+}$ current ($I_{\text{Ca,T}}$), and $Na^+\text{-}Ca^{2+}$ exchange current ($Ca^{2+}$ clock current, $I_{\text{NaCa}}$), etc.

$dV_m/dt$ is related to $I_{\text{tot}}$ in the following way:

$$\frac{dV_m}{dt} = -\frac{I_{\text{tot}}}{C_m}$$  \hspace{1cm} (2)$$

where $C_m$ is cell capacitance. If $\Delta V_m$ is the difference between the maximum diastolic potential (peak negative potential after the action potential; $\approx -60 \text{ mV}$) and the threshold potential ($\approx -40 \text{ mV}$), then the diastolic interval (DI; time between spontaneous action potentials) is given by:

$$\text{DI} = \frac{\Delta V_m}{dV_m/dt} \text{ or } \text{DI} = -\frac{\Delta V_m C_m}{I_{\text{tot}}}$$  \hspace{1cm} (3)$$

Figure 2. Effect of $\beta$-agonists on heart rate variability (HRV) in different cardiac preparations. A to D, Tachograms demonstrating the effect of $\beta$-agonists on HRV. A, Before and after dobutamine in conscious humans. B, Data before and after 100 nmol/L isoprenaline in rabbit sinoatrial node cell (SANC). C, Data before and after 100 nmol/L isoprenaline in Langendorff-perfused rabbit hearts. D, Data before and after 100 nmol/L isoprenaline in Langendorff-perfused rat hearts. Baseline data are shown in red, and data in presence of $\beta$-adrenergic agonist are shown in green. E to G, summary of the effect of $\beta$-adrenergic agonists on HRV. Mean (+SEM) CL (E), SD of normal beat to normal beat intervals (SDNN; F), and root mean square of successive differences (RMSSD) (G) under baseline conditions (=B; red bars) and with $\beta$-adrenergic agonist (=C; green bars) for the different preparations. Asterisk and bar indicate statistically different ($P<0.05$; 1-way ANOVA). Sword and bar indicate $P=0.1>P>0.05$. 

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Consider a fluctuating perturbing force that results in HRV (perhaps a change in autonomic nerve activity in the conscious animal, some unknown process in the isolated heart, or a fluctuation in the Ca\(^{2+}\) clock in the isolated SANC). This results in a fluctuating ionic current, \(I\), across the SANC membrane. During diastole (i.e., the pacemaker potential), this will change \(\frac{dV_m}{dt}\). The change in \(\frac{dV_m}{dt}\), that is, \(\Delta \frac{dV_m}{dt}\), is given by:

\[
\Delta \frac{dV_m}{dt} = \frac{I_{\text{per}}}{C_m}
\]

(4)

If one assumes that \(I_{\text{tot}}\) and \(I_{\text{per}}\) are constant throughout the pacemaker potential, the rate of change of membrane potential during the pacemaker potential will now be \(\frac{dV_m}{dt} + \Delta \frac{dV_m}{dt}\), in other words \(-\frac{I_{\text{tot}}}{C_m} + \frac{I_{\text{per}}}{C_m}\) or \(-\left(\frac{I_{\text{tot}} + I_{\text{per}}}{C_m}\right)\). This will result in a change in DI (\(\Delta\) DI):

\[
\Delta DI = \frac{AV_mC_m}{I_{\text{tot}}} - \frac{AV_mC_m}{I_{\text{tot}} + I_{\text{per}}} \text{ or } \Delta DI = AV_mC_m\left(\frac{1}{I_{\text{tot}}} - \frac{1}{I_{\text{tot}} + I_{\text{per}}}\right)
\]

(5)

Because

\[
CL = DI + APD,
\]

(6)

where APD is action potential duration, there will be a change in CL (ACL) which will be the same as the change in DI, that is, \(\Delta CL = \Delta DI\) (assuming APD is constant). \(I_{\text{per}}\) fluctuates from beat to beat (thus explaining HRV). If it is assumed that during a single DI the amplitude of \(I_{\text{per}}\) is sufficient to change the CL by 1 SD, then (from Equations 5 and 6):

\[
SDNN = -AV_mC_m\left(\frac{1}{I_{\text{tot}}} - \frac{1}{I_{\text{tot}} + I_{\text{per}}}\right)
\]

(7)

An increase in HR (decrease in CL) is achieved by an increase in \(I_{\text{tot}}\) (from 30 to 100 bpm, calculation using Equations 3 and 6 shows that \(I_{\text{tot}}\) will increase \(\approx 4.2\) times, from \(-0.22\) to \(-0.91\) pA). Equation 6 shows that as the HR increases (i.e., as \(I_{\text{tot}}\) increases), the SDNN decreases, because \(\frac{1}{I_{\text{tot}}} - \frac{1}{I_{\text{tot}} + I_{\text{per}}}\) will tend to zero. Equation 7 was solved numerically assuming that \(\Delta V_m = 20\) mV, \(C_m = 20\) pF, and \(I_{\text{per}} = 0.085\) pA. HR was calculated from the CL given by Equations 3, 5, and 6 (APD assumed to be 160 ms). The solid line in Figure 3A and 3B shows the calculated relationship between the SDNN and HR based on
this model. It is a steep exponential decay-like relationship and the line goes through the majority of the experimentally measured points. This suggests that HR is a major determinant of the changes in HRV in the clinical and experimental studies.

The model highlights 2 reasons for the decrease in SDNN at high HRs. These reasons are robust and model independent. First, a perturbing influence will produce a change in \( \frac{dV}{dt} \) that is likely to be roughly constant at all rates, and a change in slope of the pacemaker potential will obviously have a greater effect on the DI at longer baseline DIs. Second, the relationship between DI (SDNN is a DI) and HR is nonlinear and hyperbolic, because the 2 are inversely related. This acts to steepen the relationship between SDNN and HR.

Figure S3 shows the effect of \( I_{\text{p}} \) on the calculated relationship between HRV in terms of SDNN and HR: an increase in \( I_{\text{p}} \) effectively shifts the curve to higher HRs.

**Relationship Between HRV and HR as Determined by a Biophysically Detailed Model**

To confirm the dependence of HRV on HR, simulations were performed using a biophysically detailed model of the sinoatrial node action potential. The model of a rabbit central SANC (with \( C_m = 20 \text{ pF} \)) from Zhang et al. was used. Figure 4Aii shows simulated pacemaker action potentials with no perturbing current (red traces), and with a perturbing current (green traces). The maximum \( I_{\text{p}} \) was 20 pA, but the average value of \( I_{\text{p}} \) was 0.132 pA, similar to the value of \( I_{\text{p}} \) used in the simple model. Figure 4Ai shows the corresponding tachogram. The pacemaker rate of the model was modulated by changing the modeled acetylcholine concentration (0–0.04 \( \mu \text{mol/L} \)). The maximum amplitude of \( I_{\text{p}} \) (20 pA) was kept constant. The pacemaker rate was fastest in Figure 4A (acetylcholine concentration, 0 \( \mu \text{mol/L} \)) and slowest in Figure 4C (acetylcholine concentration, 0.04 \( \mu \text{mol/L} \)). In the absence of acetylcholine, when the pacemaker rate was high, the perturbing current produced a small beat-to-beat change in the CL (Figure 4Ai). When the pacemaker rate was slowed by acetylcholine, the identical perturbing current produced a greater beat-to-beat change in CL (Figure 4Ci). The tachograms from the simulations confirm that the HRV was greater at the slower HRs (Figure 4A–4Ci).

The calculated SDNN from the model is plotted against the HR in Figure 4D. The data from Figure 4 are shown in Figure 3A as gray squares. It is clear that they fall comfortably within the scatter of clinical and experimental data points.

**A Detailed Analysis of Changes in HRV in Different Preparations at Baseline and With \( \beta \)-Adrenergic Stimulation**

With the above considerations in mind, we were able to produce curves of expected change in SDNN with HR (shown as the green lines in Figure 5) and compare these with experimentally observed changes in HRV, to determine whether an experimentally observed difference in HRV (in terms of SDNN) between 2 experimental conditions can be explained simply by the difference in HR. Figure 5A shows the relationship between SDNN and HR in several different experimental preparations: the conscious human and rat, Langendorff-perfused rabbit and rat heart, and rabbit SANC (same data as shown in Figure 1).

The green line demonstrates the expected change in SDNN with HR (based on a decrease in \( \log_{10}(\text{SDNN}) \) of 0.169 ms for every 10 bpm increase in HR, as calculated from Figure 3C), arbitrarily using data for the conscious human as the starting point (thus explaining why the line passes exactly through the conscious human data point; the choice of starting point makes no difference to the analysis). It is apparent that the decrease in HRV in the rabbit SANC, Langendorff-perfused rat heart, and conscious rat can be largely accounted for by the higher HR in these preparations (the data points lie above the green line).

However, the decrease in HRV in the Langendorff-perfused rabbit heart may not be completely accounted for by the higher HR in this preparation (the point lies below the green line).

Figure 5B to 5E shows a similar analysis of the effect of \( \beta \)-stimulation on HRV in the conscious human (Figure 5B), rabbit SANC (Figure 5C), and Langendorff-perfused rabbit (Figure 5D) and rat (Figure 5E) heart. In each case, the decrease in HRV seen following exposure to catecholamine can be completely explained by the increase in HR (the dobutamine/isoprenaline points lie on or above the green lines). Such an understanding of the expected change in HRV where HR is different between experimental preparations or conditions is essential when trying to determine whether there is an HR-independent difference in HRV between \( \geq 2 \) situations.

Figure 5 is a graphical method of correcting for changes in HR. It is also possible to correct for HR numerically:

\[
\text{cSDNN} = \frac{\text{SDNN}}{e^{\frac{\text{HR}}{58.8}}} \tag{8}
\]

where \( \text{cSDNN} \) is the corrected SDNN (at an HR of 0 bpm). Figure S4 shows that it is possible to eliminate the HR dependence of the HRV data in Figure 3 using this correction factor. Equation 8 can be rewritten to give corrected SDNN at any reference HR \( (HR_{\text{ref}}) \):

\[
\text{SDNN}_{\text{ref}} = \frac{\text{SDNN}}{e^{\frac{(\text{HR}_{\text{ref}} - \text{HR})}{58.8}}} \tag{9}
\]

**Discussion**

We have demonstrated in detail that substantial HRV exists in isolated cardiac preparations without autonomic connections. This HRV exists under baseline conditions and is modified by drugs that modify beating rate, including exposure to a \( \beta \)-adrenergic agonist. We have shown that regardless of species, preparation, conditions, method of determination, and laboratory, there is a unique exponential decay-like relationship between HRV (as measured by the SDNN) and HR (Figure 3). This is true both in the healthy and diseased heart. Using mathematical modeling, we propose a biophysical explanation for the relationship between HRV and HR: at high HRs, the ratio of the perturbing current, \( I_{\text{p}} \) (regardless of its nature), to the intrinsic ionic current, \( I_{\text{p}} \), driving the pacemaker potential becomes vanishingly small and the effect of \( I_{\text{p}} \) becomes negligible. This can be stated in another way: \( I_{\text{p}} \) will change the slope of the pacemaker potential by roughly the same amount regardless of rate; the effect of this change will be smaller the higher the HR. As such, HRV is primarily dependent on HR and cannot be used in any simple way...
to assess autonomic nerve activity to the heart. Although our analysis has focused on SDNN, our arguments are germane to many commonly used methods to measure HRV.

**Earlier Indications of a Relationship Between HRV and HR**

It has long been suspected that HR has a significant effect on HRV. Mangin et al observed this relationship in conscious rats, showing that SDNN, RMSSD, and the sum of low- and high-frequency power are all significantly correlated with CL. Coumel et al also demonstrated a significant relationship between SDNN and HR (correlation coefficient, 0.79). Neither of these studies quantitatively established the impact of HR on HRV. Zaza and Lombardi, however, did. Using isolated rabbit SANCs and modifying HR using acetylcholine, they were able to demonstrate that the relationship between

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**Figure 4.** Relationship between heart rate variability (HRV) and heart rate predicted by a biophysically detailed model. Computed action potentials from a sinoatrial node cell (SANC) model with (green) and without (red) a maximum 20-pA perturbing current ($I_{\text{per}}$) at fast (top, eg, $A_1$) to slow (bottom, eg, $C_1$) rates, and corresponding tachograms ($A_{\text{ii}}$–$C_{\text{ii}}$). Rate was varied by altering ACh-activated K+ current ($I_{\text{K,ACH}}$). D. Relationship between SD of normal beat to normal beat intervals (SDNN) and heart rate in this model.
concentration of acetylcholine and CL was not linear, nor was the relationship between CL and diastolic depolarization rate. This hinted at a strong rate dependency of several commonly used indices of HRV, including SDNN. Despite this article being published >10 years ago, there continues to be a marked underappreciation of this relationship. Our work aims to further clarify the importance of this and the need to make allowances for it.

Zaza and Lombardi used model data to calculate that logn(SDNN) decreases by 0.17 ms for every 10 bpm increase in HR. Similarly, Tsuji et al. used 2-hour snapshots of ambulatory cardiac monitoring to define that log n(SDNN) decreased by 0.16 ms for every 10 bpm increase in HR. In our study, logn(SDNN) decreased by 0.169 ms for every 10 bpm increase in HR (Figure 3B). The similarity of these numbers demonstrates the highly conserved relationship between HRV and HR. Similarly, our simple model predicts that logn(SDNN) decreases by 0.24 ms per 10 beat/min increase in HR (Figure 3A and 3B, solid line).

Sacha and Pluta have pointed out that the variation in CL (R-R interval) for a given variation in HR is greater at slower HRs, because of the inverse relationship between HR and CL, and suggested a correction procedure, which could eliminate this effect. We agree that this is one reason why SDNN varies with HR, as stated above it acts to steepen the relationship between SDNN and HR. However, the primary reason why SDNN varies with HR is the nonlinear relationship between the change in CL (ΔCL) and CL (for a given Iper): the correction procedure from Sacha and Pluta does not take this into account.

Figure 5. Application of a correcting factor facilitates clarification of whether changes in heart rate variability (HRV) are attributable to heart rate differences alone. Analysis of baseline differences in HRV among different preparations (A); analysis of changes in HRV after administration of β-agonists to different preparations (B–E). SD of normal beat to normal beat intervals (SDNN) is plotted against heart rate. Predicted effect of heart rate on SDNN is plotted (green lines) using calculated correcting factor from Figure 3C. Conscious human data (A) or baseline data (B–E) are arbitrarily used as starting point from which to calculate the effect of heart rate on SDNN.
Multiple nonlinear methods for the interpretation of HRV have been developed during the past several years (eg, approximate entropy), yet their take-up has been limited. It remains unclear whether these do indeed give a true HR-independent means with which to assess HRV.

**Can HRV Be Corrected for the Effect of HR?**

Previously, several authors have attempted to correct for the effect of HR on HRV by normalizing HRV parameters by simply dividing them by the HR at that point in time (eg, Hayano et al\textsuperscript{15}). Such linear corrections are inadequate to correct the nonlinear relationship between HRV and HR. This is demonstrated by Figure S5. In the power spectrum of HRV, the high-frequency band is said to represent parasympathetic activity, the low-frequency (at least in part) sympathetic activity, and Mortality

The higher morbidity and mortality associated with decreased HRV has been presumed to be the consequence of autonomic imbalance (sympathetic excess, parasympathetic withdrawal, even bilateral autonomic withdrawal).\textsuperscript{2,3} This must now be reinterpreted. We argue that HRV is a nonlinear measure of autonomic nerve activity to the heart,\textsuperscript{10} and undoubtedly the mechanisms proposed in the present study are not expected to affect this ratio. However, for independent reasons, the validity of this ratio has been challenged.\textsuperscript{16}

Tsuji et al\textsuperscript{11,12} used a good method. They studied HRV (SDNN) in 736 human subjects enrolled in the Framingham study, finding that HRV was correlated with the HR (in the manner that can be explained by the present study). However, they\textsuperscript{11,12} showed that age was an independent factor determining HRV, with HRV being lower in older subjects for a given HR. They were able to separate age from HR, because of the large number of subjects investigated. If it is assumed that HRV is primarily the result of fluctuations in autonomic nerve activity in the conscious human (see above), the study of Tsuji et al\textsuperscript{12} suggests a decrease in autonomic nerve activity with age.

Figure 5 shows a graphical method of separating an independent factor from the effect of HR. For example, Figure 5A suggests that HRV in Langendorff-perfused rabbit hearts (compared with the conscious human) is lower than can be accounted for by the higher HR, suggesting that $I_{\text{ref}}$ in the Langendorff-perfused rabbit heart is lower than in the conscious human. This is not unexpected: there is no fluctuating autonomic nerve activity in Langendorff-perfused rabbit hearts.

Finally, the easiest method to correct for HR is to use Equation 8 as shown by Figure S4.


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**Novelty and Significance**

**What Is New?**

- A unique relationship exists between heart rate variability (HRV) and heart rate across diverse cardiac preparations, both innervated and denervated, from the living animal all the way down to the fundamental building block of automaticity, the sinoatrial node cell.
- This relationship is independent of the conditions or duration over which HRV is recorded and persists even when drugs or gene modification are used to affect cellular processes.
- We have modeled this relationship using diverse biophysical models of differing complexity.
- We have suggested how to adequately correct for the phenomenon.

**What Is Relevant?**

- Our findings are significant because many articles have previously been published using HRV data that are not corrected for heart rate.
- Articles continue to be published apace with this fundamental flaw, and yet with simple correction, the effect of heart rate could be removed and true differences in HRV revealed.
- This has ramifications for the handling of blood pressure variability data.

**Summary**

HRV data should always adequately take into the account the heart rate at which they were measured.
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BIOPHYSICAL CHARACTERISATION OF THE UNDER-APPRECIATED AND IMPORTANT RELATIONSHIP BETWEEN HEART RATE VARIABILITY AND HEART RATE

Short title: Heart rate variability and heart rate

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Supplementary Methods

Experiments performed for this study focussed upon three different cardiac preparations: conscious animals, isolated denervated Langendorff-perfused hearts, and isolated sinoatrial node cells. The species employed in each of these situations varied, as did the experimental conditions used.

Conscious human study

Male and female patients (mean age, 58-2 years, n=11, 55% male) attending the Manchester Heart Centre (Manchester Royal Infirmary, Oxford Road, Manchester, UK) for out-patient dobutamine stress echocardiography to aid in their pre-operative assessment were recruited to the study. Ethical approval for the study had been granted by the local ethics committee. All experiments were performed between 09:00 and 12:00 hours to minimise interference from circadian factors. Patients in the supine position were exposed to an incremental infusion of dobutamine every 3 min, beginning (after a period of baseline ECG recording) with 5 µg/kg/min, progressing through 10, 20, 30, and reaching a peak at 40 µg/kg/min. The target was an 85% increase in heart rate based on their maximum predicted heart rate by age and sex. Patients were given information regarding the study prior to undergoing the test, and were advised not to drink alcohol or caffeinated beverages for 24 hours leading up to the test. Those patients taking β-blocking medication were advised to discontinue this 72 hours prior to the day of the test. Patients indicated their consent by signing a hospital-approved consent form. Adhesive electrodes were placed on the right and left shoulders and left leg of the patient, and were connected to a Finapres Finometer ECG module (Finapres Medical Systems, Amsterdam, Netherlands). This was connected to a CED 1401 analogue to digital converter (Cambridge Electronic Design, Cambridge, UK) sampling the signal at 1000 Hz, which in turn was connected to a personal computer, where Spike2 software (Cambridge Electronic Design) made continuous recordings of the ECG data in the form of .cxd files. These were exported directly into LabChart Pro software (AD Instruments, BellaVista, NSW, Australia) and full analysis of HRV was made after the ECG signal had been subjectively checked for stationarity.

Conscious rat study

Three-month old Sprague-Dawley male rats (n=11) were fitted with intra-abdominal ETA-F20 biopotential transmitters (DSI, St Paul, MN, USA; weight 3-9 g, volume 1-9 ml) under isoflurane general anaesthesia whilst spontaneous respiration continued. The two flexible electrodes emanating from the transmitter box (helices of stainless steel wire covered by insulating silicone tubing) were implanted subcutaneously in the pre-cordial region of the rat, one rostrally and one caudally. Dissolvable sutures were used to close the operated site. Rats were allowed one week of recovery before first recordings were made. The rats were not handled excessively, or treated differently to prior to the insertion of telemetry devices. ECG recordings at baseline were made by placing the plastic housing of the rat’s cage onto an RPC-1 PhysioTel receiver (DSI, St Paul, MN, USA), which in turn was remotely connected to a Data Exchange Matrix (DSI, St Paul, MN, USA) to facilitate multiple simultaneous recordings from several rats. All telemetry data were recorded in the morning hours, between 09:00-11:00. The data was passed to a personal computer for storage and offline analysis, which was performed by LabChart Pro software. The transmitters could be turned on and off via the use of a magnetic switch to conserve battery life for the periods of recording only. The first 150 s of stationary data were selected for analysis of HRV data.

Langendorff-perfused rabbit and rat heart study

Three-month old male New Zealand white rabbits (n=58) were sacrificed by lethal injection of 2-3 ml 20% sodium pentobarbital into a pre-anaesthetised (with EMLA cream: 25 mg/g lidocaine, 25
mg/g prilocaine, AstraZeneca, Alderley Park, Cheshire, UK) dorsal ear vein, in accordance with the requirements set out in the Animals (Scientific Procedures) Act 1986. The heart was immediately removed, and placed into 250 ml ice cold Tyrode’s solution (containing, in mM, 120 NaCl, 4 KCl, 1·3 MgSO₄*7H₂O, 1·2 NaH₂PO₄*2H₂O, 1·2 CaCl₂, 25·2 NaHCO₃, 5·8 glucose), containing 1000 units heparin sodium (LEO Pharma, Buckinghamshire, UK) to prevent coronary thrombosis prior to the re-establishment of coronary flow upon connecting the Langendorff perfusion rig. Following removal of the heart, the aorta was flushed retrogradely with 25-50 ml ice-cold heparin-containing Tyrode’s solution. It was then rapidly transferred (immersed in ice cold heparinised Tyrode’s solution) to the electrophysiology rig, where curved forceps were used to mount it onto a custom-designed steel cannula through which was passing a constant flow of Tyrode’s solution (not containing heparin; equilibrated with 95% O₂/5% CO₂; 20 ml/min via a peristaltic pump - Minipuls3, Gilson, Middleton, WI, USA). The Tyrode’s solution was maintained at a constant temperature of 37±0·4°C by heat exchanging glassware plumbed into a water bath (constant temperature of the solution was verified at the superior end of the heat exchanging column by a temperature monitoring system (Space Labs, Washington DC, USA).

Hearts generally recovered spontaneous beating within 2 min. Any heart that failed to re-establish spontaneous beating within 5 min was discarded. Following re-establishment of regular beating, a stabilisation period of 20 min was allowed. Spring-loaded ECG electrodes (Harvard Apparatus) were positioned over the surface of the right atrium and on the free wall of the left ventricle. Obvious vascular structures were avoided during positioning of the electrodes. The ECG electrodes, along with the ground electrode, were connected to a head-stage (Digitimer, Welwyn Garden City, UK), and then via a filter/amplifier (Digitimer) to an analogue-to-digital converter (CED 1401), and finally to a personal computer where Spike2 software was used to record the signal. Online filtering and amplification of the signal was performed at frequencies of D.C. for the low pass filter, and 1 kHz for the high pass filter. Further offline analysis was achieved using LabChart Pro software. Following the 20 min recovery period, a period of 500 s of baseline ECG recording was made. Thereafter, the effect of several chronotropically active drugs, isoprenaline (100 nM, n=8), carbachol (2x10⁻⁷ M, n=9) and caesium chloride (2mM, n=8), was studied by switching a three-way tap to facilitate access to a second water-jacketed reservoir. The drug was allowed 10 min to achieve its effect on the heart, following which time a further 500 s of ECG recording was performed.

The basic procedure outlined above in the rabbit is almost identical to the Langendorff-perfusion experiments performed on the rat. Three-month old Wistar rats (n=8) were killed by stunning and cervical dislocation in accordance with the Animals and Scientific Procedures Act (1986). Following an identical procedure for removal of the heart, it was flushed with 20 ml of ice cold heparinised Tyrode’s solution before transfer to the Langendorff perfusion rig, where the flow rate had been reduced to 10 ml/min through a smaller metal cannula designed especially for the rat aorta. The procedure for testing the effects of chronotropically active drugs were identical to those described above (n=8 for isoprenaline).

**Isolated rabbit sinoatrial node cell study**

Studies in isolated SANC from rabbits were conducted in accordance with the National Institutes of Health Guide for the Care and Use of Laboratory Animals. The isolation and preparation of sinoatrial node cells has been described elsewhere¹ and identical techniques were used in this study. Three-month old New Zealand white rabbits were sacrificed by approved methods. The sinoatrial node was dissected free from the rest of the heart and subjected to triple enzymatic digestion, yielding a concentrated sinoatrial node cell-containing solution. 15-50 μl (depending on cell yield of the individual isolation) of the sinoatrial node cell-containing solution was pipetted into the centre of a custom-built chamber (capacity, 400 μl) attached to the stage of a Zeiss
Axiovert 100 inverted microscope (Carl Zeiss, Inc., Germany) containing Ca$^{2+}$-containing HEPES-based cell perfusion buffer (containing, in mM, 140 NaCl, 5 HEPES, 0-33 NaH$_2$PO$_4$*H$_2$O, 5-4 KCl, 5-5 glucose, 0-5 MgCl$_2$*6H$_2$O, 50 taurine) by a dropper pipette – at this point, circulation of the cell perfusion buffer and temperature regulating equipment were switched off.

Sinoatrial node cells were added from a Gilson pipette, and allowed to settle to the bottom of the chamber for 20 min. Following this, circulation of the HEPES buffer was commenced at a low rate (1-2 ml/min) using a gravity driven perfusion system. A dual thermoregulation system was activated (Analog TC2BIP 2/3Ch Bipolar Temperature Controller; Cell Micro Controls, Norfolk, VA, USA) that incorporated a thin-film, transparent ITO (indium-tin-oxide) plate heater of the glass onto which the cells settled, and a pre-heating coil surrounding the entry tube for the fluid prior to its entrance into the chamber. Both of the heaters were controlled thermostatically by a thermistor located within the bath itself. This setup allowed strict maintenance of the HEPES based buffer solution at 35±0·1°C. Upon commencement of circulation of the buffer and the heating of the chamber, the sinoatrial node cells began to spontaneously contract.

Borosilicate glass micro-pipettes (World Precision Instruments, Sarasota FL, USA) were generated (diameter, ~2 μm, confirmed by visual inspection under a microscope; typical resistance, 3-5 MΩ) using a horizontaly-loading micro-pipette puller (Sutter Instruments, Novato, CA, USA), and were filled with high K+ concentration fluid containing ATP (containing, in mM, 120 potassium gluconate, 5 NaCl, 5 MgATP, 5 HEPES, 20 KCl), and 35-50 μmol/l β-escin (Sigma Aldrich, St Louis MO, USA) as the pore-forming active ingredient. An electronically operated micromanipulator (EXFO Life Sciences, Ontario, Canada) was used to guide the micro-pipette to the chosen cell under direct vision, initially using a 10x objective lens, switching to a 40x or 63x objective lens for the final approach to the cell. Action potentials were recorded from healthy and spontaneously beating cells (n=67) using an Axopatch-200 B patch-clamp amplifier (Axon Instruments, Foster City, CA). Action potential measurements were performed in a standard zero-current-clamp (fast) mode using a gap-free acquisition protocol with a sampling interval of 0.1 ms. Recordings were only made from cells with acceptable electrophysiological parameters of a maximum diastolic potential more negative than -50 mV and a peak potential of >20 mV. Action potential signals were recorded as ‘.abf’ files on the hard drive of the personal computer for further analysis.

The effect of several perfusates, isoprenaline (100 nM, n=6), carbachol (1x10$^{-7}$ M, n=4), caesium chloride (2mM, n=9), zatebradine (20 μM, n=4) and ryanodine (3 μM, n=23), was investigated on the spontaneous beating of the isolated rabbit sinoatrial node cells. These substances were made up to the desired concentration in the cell perfusion buffer (containing, in mM, 140 NaCl, 5 HEPES, 0-33 NaH$_2$PO$_4$*H$_2$O, 5-4 KCl, 5-5 glucose, 0-5 MgCl$_2$*6H$_2$O, 1-8 CaCl$_2$; pH corrected to 7-35 using 1 M NaOH) and directly added to the chamber via a separate gravity driven perfusion system at an identical rate to the baseline perfusate. Switching between the two was achieved via a simple three-way tap.

Action potentials were recorded and stored on a personal computer. Each individually patched cell was allowed 20 min to undergo β-escin-induced membrane perforation, i.e. to achieve acceptable electrophysiological parameters. If perforation was not achieved during this time, the cell was discarded. Once perforation was achieved, baseline recording was immediately commenced. The period of baseline recording was 5 min. If there was a substantial change in beating rate or morphological appearance of the cell during this period sufficient to suggest that the patching process had damaged the cell, then the cell was discarded. If, however, beating rate and cellular appearance remained stable, the experiment was stored for offline analysis. When other substances were added, following the 5 min period of baseline recording, the perfusate was switched from the HEPES-based cell perfusion buffer, to the isoprenaline/carbachol/caesium/zatebradine/ryanodine-containing HEPES-based cell perfusion
buffer. This was allowed to run in over the following 5 min. Following this, a further 5 min period of recording was commenced to demonstrate the effects of the intervention on variability. Only cells with an initial cycle length of <500 ms in the baseline period of recording were further analysed. From the 5 min baseline and 5 min isoprenaline records, a 2.5 min (or 150 s) segment of action potential data was objectively selected on the basis of stationarity, lack of ectopic or artefactual beats and quality of recording. No attention was paid to the actual interbeat intervals making up this selected segment during the process of its selection; indeed such parameters had not been calculated at this stage. For the analysis of HRV, the .abf files were directly imported into LabChart Pro software.

**HRV analysis**
Continuous recordings of single channel ECG data from the conscious human or rat and the isolated Langendorff-perfused rabbit and rat heart or spontaneous action potentials recorded from the isolated rabbit sinoatrial node cell were subjected to HRV analysis along identical lines. Time epochs of 2.5 min/150 s were chosen for HRV analysis on the basis of subjective stationarity. A stationary time series is “one whose statistical properties such as mean, variance, autocorrelation, etc. are all constant over time”. For the purposes of HRV tachograms, stationarity refers to a series of RR intervals without trend, but with constant variance over time, in addition to a constant autocorrelation structure over time and no periodic fluctuations. Strict rules for data pre-processing were employed in order that meaningful HRV data could be obtained, without over-processing of the signal and therefore loss of data characteristics. HRV parameters were calculated in the time-domain – we calculated the standard deviation of normal to normal beats (SDNN; i.e. the standard deviation of cycle length between normal, not ectopic, beats), standard error of the mean, normalised standard deviation, and root-mean-square of the successive differences (RMSSD). HRV parameters were also calculated in the frequency domain (total power present within the signal at all frequencies; power in very-low-frequency/low-frequency/high-frequency bands; low-frequency to high-frequency power ratio) and the non-linear domain (Poincaré plots), enabling a broad-based appreciation of the HRV present within the recorded signals. For simplicity, only SDNN and total power are presented as the time- and frequency-domain parameters reflecting HRV in figures.

**Computer modelling**
The Zhang et al. deterministic ordinary differential equation (ODE) model for the electrophysiology of a rabbit central sinoatrial node cell was implemented in this study. This model incorporates the concentration-dependent chronotropic effects of the ACh-activated K⁺ current \( (I_{K,ACH}) \) and the actions of ACh on the conductance and kinetics of the L-type Ca²⁺ current \( (I_{Ca,L}) \) and the funny current \( (I_f) \). In general, the equation governing the time-dependent change in membrane potential \( (V_m) \) with the effect of ACh is:

\[
C_m \frac{dV_m}{dt} = -(I_{tot} + I_{K,ACH}),
\]

where \( C_m \) is the cell capacitance, \( t \) is time and \( I_{tot} \) the total ionic current in a sinoatrial node cell. Full details of equations and parameters of the Zhang et al. models of sinoatrial node cells and the actions of ACh are documented in our previous studies and working codes in C are available on request. Fluctuations in cycle length (HRV) were simulated by including a stochastic perturbing current in Equation [1]:

\[
\]
\[ C_m \frac{dV_m}{dt} = -(I_{\text{tot}} + I_{K,ACb} + I_{\text{per}} \xi(t)), \]  

where \(I_{\text{per}}\) is the maximum amplitude of the perturbing current (20 pA), and \(\xi(t)\) is a random fluctuation term (-1≤\(\xi(t)\)≤1) and was generated by the intrinsic function \textit{rand}(l) of the C complier with uniform deviates. Equations [7] and [8] were numerically integrated by the fourth-order Runge-Kutta method with a time step of 0.2 ms, which was sufficiently small for stable solutions.

Supplementary Discussion
Speculating on the nature of \(I_{\text{per}}\)
Our arguments are independent of the nature of \(I_{\text{per}}\), and we have not explored its nature in this study. In the conscious animal, there is no reason to believe that \(I_{\text{per}}\) does not represent fluctuations in autonomic nerve activity to the heart. Although there is no evidence to reject this, there is also no evidence to confirm it either. Although autonomic blockade abolishes HRV in the conscious human, \(^5\-\(^7\) it also increases heart rate, and the abolition of HRV could be significantly contributed to by the increase in heart rate. Further work is required to resolve this. Ideally, autonomic tone should be directly measured by electrodes.

In the isolated sinoatrial node cell, the considerable HRV cannot be the result of fluctuations in autonomic nerve activity. Instead evidence from our own group suggests that it is intrinsic to the sinoatrial node cell.\(^8\) According to the coupled clock theory, the heart rate is set via mutual entrainment of the Ca\(^{2+}\) and membrane clock pacemaker mechanisms.\(^9,\,10\) This concept has been tested via perturbations of the Ca\(^{2+}\) clock using cyclopiazonic acid and the membrane clock with ivabradine.\(^11\) In the isolated sinoatrial node cell, HRV has been shown to be the result of beat-to-beat fluctuations in the Ca\(^{2+}\) clock.\(^8\) In the present study, we either modelled \(I_{\text{per}}\) as a constant current during the pacemaker potential (simple model) or as a randomly fluctuating current during the pacemaker potential (biophysically-detailed model), but both approaches predicted a similar relationship between SDNN and heart rate (see Fig. 3A, solid line and Fig. 4D). Nevertheless, it is not known which approach most closely represents reality when HRV is generated by fluctuations in autonomic tone or the Ca\(^{2+}\) clock. In the case of the Ca\(^{2+}\) clock, the amplitude of the associated current (\(I_{\text{per}}=I_{NaCa}\)) is increased during the late phase of the pacemaker potential shortly before the action potential upstroke.\(^12\) A crucial parameter defining CL duration is the \textit{phase} of Ca\(^{2+}\) release, rather than its \textit{amplitude}.\(^8\) Thus, the increase in Ca\(^{2+}\) release amplitude and \(I_{NaCa}\) occurring during the late phase of the pacemaker potential may not necessarily generate substantial CL fluctuations. These ideas are in line with those of Jalife \textit{et al.},\(^13\) who showed that the CL is more influenced by a current perturbation if it is applied in the so-called “1:1 entrainment zone” (≈20% of the CL before an excitation, i.e. the late phase of the pacemaker potential). These complexities, however, do not negate the central conclusion of this study - regardless of whether \(I_{\text{per}}\) is constant or fluctuating and whether it preferentially occurs during one part of the pacemaker potential, the effect of \(I_{\text{per}}\) on CL is expected to be less at high heart rates when \(I_{\text{tot}}\) during the pacemaker potential is greater.

In rabbit sinoatrial node cells, Bogdanov \textit{et al.}\(^12\) showed that fluctuations in the rate of diastolic depolarization were larger after the application of the \(\beta\)-adrenergic agonist, isoprenaline. Computer simulations suggested that the fluctuations in \(I_{NaCa}\) during the latter part of the diastolic depolarization substantially increase.\(^12\) This result is not inconsistent with the finding that \(\beta\)-adrenergic agonist decreases HRV (Fig. 2), a finding that has also been reported previously.\(^14\) The data from Bogdanov \textit{et al.}\(^12\) suggests that \(I_{\text{per}}\) \textit{increases} with \(\beta\)-adrenergic agonist. However, the fact that \(\beta\)-adrenergic agonist \textit{decreases} HRV in the conscious human, Langendorff-perfused rabbit and rat heart and rabbit sinoatrial node cell (Fig. 2) means that this effect is masked by the direct effect of heart rate on HRV.
In isolated denervated Langendorff-perfused hearts, HRV cannot of course be the result of fluctuations in autonomic nerve activity to the heart. Although there are parasympathetic ganglia in the heart and consequently intact postganglionic nerve fibres (as well as sympathetic nerve endings), it is well established that there is no residual nerve activity that could lead to significant HRV. Computer simulations show that a group of electrically-coupled sinoatrial node cells (each showing independent stochastic variation in beating rate) shows less HRV than the individual uncoupled sinoatrial node cells – see Fig. S6. This phenomenon of decreasing variability with increasing cell coupling has been recognized for some time, though not modelled as we have done here. In keeping with this, the HRV exhibited by the Langendorff-perfused rabbit heart is less than that demonstrated by the isolated rabbit sinoatrial node cell in a way that cannot be explained by the difference in CL (Fig. 1). The nature of $I_{per}$ in the isolated Langendorff-perfused heart is not known and warrants further study.

**Limitations of the study**
The simple model provides a general explanation of the dependence of HRV on heart rate. However, it depends on some simplifying assumptions. It depends on $I_{tot}$ (as well as $I_{per}$) being roughly constant throughout the pacemaker potential - $I_{tot}$ will vary in response to the $I_{per}$-induced change in $dV_m/dt$. However, the model is still valid if the membrane conductance during the pacemaker potential is roughly constant. To prevent the substantial decrease in $SDNN$ at high heart rates, there would have to be a substantially lower conductance during the pacemaker potential at high heart rates, and there is no reason for this – at high heart rates, although activation of some ionic currents will be less, residual activation of $I_{Kr}$ and $I_{Ks}$ will be greater. The simple model assumes action potential duration of a SANC to be constant at different rates. This is not an unreasonable assumption because the sinoatrial node does not show large rate-dependent changes in action potential duration unlike Purkinje fibres and ventricular muscle. In addition, in the biophysically detailed model the action potential duration is allowed to change and still HRV depends on heart rate in a similar manner to that predicted by the simple model.

In the biophysically detailed model, we modelled the effect of a randomly fluctuating perturbing current, $I_{per}$. In the future, it would be of interest to model fluctuating autonomic tone and analyse the power spectra.

**Detailed key for Fig. 3**
HRV data was taken from the following papers, in addition to our own: Gehrmann et al.; Uechi et al.; Laude et al.; Bissonette et al.; Wu et al.; Ecker et al.; Zaza et al.; Radaelli et al.; Beckers et al.; Ramaekers et al.; Seals et al.; Sacknoff et al.; Tsuji et al.; Casolo et al.; Claria et al.; Al-Ani et al.; Craft et al.; Aubert et al.; Abildstrom et al.; Jensen-Urstad et al.

**References**

2. Forecasting - duke university. 2005
**Fig. S1.** A-E: Power spectra under baseline conditions from all five species studied to show differences in power present at frequencies up to 0.5 Hz. Note differences in y-axis scales. A: conscious human; B: rabbit Langendorff heart; C: Rabbit SANC; D: rat Langendorff heart; E: conscious rat. F: summary data to demonstrate differences in mean total power between preparations. * = statistically significant difference on 1-way ANOVA. Letters atop bar reflect which preparation(s) are statistically significant from the starred bar.
**Fig. S2. Relationship between HRV (SDNN) and heart rate in different preparations.** Data shown for different species. Notice that the same exponential decay-like relationship is observed in every case.
Fig. S3. Effect of the amplitude of the perturbing current ($I_{\text{pert}}$) on the relationship between HRV (SDNN) and heart rate as predicted by the simple model.
Fig. S4. Correction of SDNN for heart rate by dividing the SDNN by $e^{\frac{\text{heart rate}}{58.8}}$ (equation 8). Top, relationship between uncorrected HRV (SDNN) and heart rate. Same data as shown in Fig. 3. There is an increase in HRV as heart rate decreases. Bottom, relationship between corrected HRV and heart rate. There is a flat relationship between HRV and heart rate.
Fig. S5. Figure to show the effect of simple normalisation (dividing the HRV parameter, in this case SDNN, by the heart rate at which it was measured). Top panel shows raw data taken from our simple model in red. Bottom panel shows the effect of simple normalisation on this data – there is persistence of a large effect of heart rate on HRV, especially at heart rates <100.
Fig. S6. Results of the computer modelling experiments investigating the differences in variability between cardiac preparations of differing complexity. A, pertains to modelled results from a single isolated SANC; B, pertains to results from two SANC joined together; C, pertains to results from a nine SANC matrix. A,i - a 30-second tachogram recorded from a spontaneously-beating single isolated SANC in vitro was used as the 'model variability', and this pattern of variable beating was instilled into the modelled cells. A,ii-iv - a one-minute recording of raw action potential data (A,ii), the resultant beat-to-beat tachogram (A,iii) and the ensuing Poincare plot.
(A,iv) in a computer modelled cell based on the Kurata 35 model. Initially (for the first 30 seconds), the modelled cell is allowed to beat at its own intrinsic rate. It can be seen that during this time, there is no beat-to-beat variability (the flat line in the first part of the tachogram most efficiently illustrates this - A,iii). However, at 30 seconds, a variable current (0.1 pA) based on the variability that had previously been recorded in vitro (A,i) was ‘injected’ into the model cell. This caused the immediate onset of similar beat-to-beat variability in the modelled cell (A,iii). This is reflected in the Poincaré plot, which illustrates the presence of both short- and long-term variability (A,iv). In a, v-vii, the process is repeated, except that this time the magnitude of the injected current is doubled (to 0.2 pA). It can be seen that this affects action potential amplitude (A,v), along with beat-to-beat variability, illustrated as an increase in amplitude of the tachogram (A,vi) and a greater spread of the points in the Poincaré (A,vii), in terms of both short- and long-term variability. B,i - the 30-second tachogram recorded from the same spontaneously-beating single SANC in vitro as used in the single cell modelling experiments was used to instil variability into two modelled SANC joined together. The timing of the injected variability was staggered in each of the two cells, in order that they might be considered as expressing variability independent of each other. When the modelled cells were not electrically coupled together (B,ii; coupling conductance \((g_j) = 0 \text{nS})\), the relative levels of variability expressed by each were high (B,iii). However, if the modelled cells were electrically coupled (B,iv; \(g_j = 25 \text{nS}\)), then the levels of variability as subjectively assessed from the resultant tachogram decreased (B,v). C,i - the same 30-second tachogram as recorded from the spontaneously-beating single SANC in vitro that was used in the single- and dual-modelled cell experiments was used to instil variability into each of nine cells arranged in a 3 x 3 matrix. When the cells were not electrically coupled together (C,ii; \(g_j = 0 \text{nS}\)), levels of expressed variability could be seen to be relatively high, both in terms of the amplitude of the resulting tachogram (C,iii) and the dispersion of the points on the Poincaré plot (C,iv). However, when the matrix was electrically coupled (C,v; \(g_j = 25 \text{nS}\)) the level of expressed variability decreased substantially in terms of both the amplitude of the resultant tachogram (C,vi) and the Poincaré plot (C,vii).