The kidney plays an important role in the long-term control of blood pressure and is the major organ involved in the regulation of sodium homeostasis. The inappropriate sodium retention in hypertension results from enhanced renal sodium transport per se or a failure to respond appropriately to signals that decrease renal sodium transport in the face of increased sodium intake. Humans with polygenic essential hypertension have increased renal sodium transport that is not properly regulated by natriuretic and antinatriuretic hormones and humoral factors, including dopamine and angiotensin II (Ang II). Dopamine and Ang II exert their effects via G protein-coupled receptors (GPCRs).

GPCRs constitute by far the largest receptor family in mammals, which are encoded by >800 genes in the human genome and play a vital role in the regulation of most cellular and physiological functions in the body. On ligand binding, GPCRs regulate and modulate a variety of cell functions by coupling to heterotrimeric G proteins and regulating downstream effectors, such as adenylyl cyclases, phospholipases, protein kinases, and ion channels. Activation of renal GPCRs, including dopamine and Ang II receptors, leads to either natriuresis (sodium excretion) or antinatriuresis (sodium retention), keeping a normal sodium balance, resulting in the maintenance of a normal blood pressure.

GPCR kinases (GRKs) constitute a family of 7 serine/threonine protein kinases characterized by their ability to specifically recognize and phosphorylate agonist-activated GPCRs. GRK-mediated receptor phosphorylation is one of the well-characterized mechanisms for GPCR desensitization. In the process of GPCR desensitization, GRKs phosphorylate agonist-bound receptors, leading to the translocation and binding of arrestins to the receptors and inhibition of subsequent receptor activation by blocking GPCR-G protein coupling. In particular, GRK4 seems to play a vital role in regulating dopamine-mediated natriuresis and renin–angiotensin system (RAS)–mediated antinatriuresis. Increasing number of studies show that GRK4 is associated with hypertension and blood pressure response to antihypertensive medicines and adverse cardiovascular outcomes of antihypertensive treatment.

In this report, we review our evolving understanding of the role of GRK4 in the regulation of dopamine and Ang II receptor function, which advances our understanding of the role of GRK4 in the control of blood pressure and highlights potential and novel strategies for the prevention and treatment of hypertension.

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phosphorylation and desensitization of the D₃R. However, the GRKs have received, by far, the most attention in the regulation of renal dopamine and Ang II receptors in hypertension.

**GPCR Kinase Family**

**Classification of GRKs**

There are over 800 known GPCRs in the human genome, but only 7 GRKs have been identified. All GRKs have a similar general structure: a highly conserved central protein kinase domain, inserted in the regulator of G protein signaling homology domain that keeps the ability of the kinase domain to phosphorylate activated GPCRs. The first 20 or so amino acids of GRKs are highly conserved, whereas the carboxy tail region is GRK subtype-specific: prenylated in the GRK1 subfamily; binds to Gpγ and contains a pleckstrin homology domain in the GRK2 subfamily; and has a C-terminal helix/palmitylation site in the GRK4 subfamily. The GRK1 subfamily (opsin kinase family) consists of GRK1 and GRK7; GRK2-like subfamily (β-adrenergic receptor kinase family) consists of GRK2 and GRK3; and the GRK4-like subfamily consists of GRK4, GRK5, and GRK6. GRK1 and GRK7 are found almost exclusively in the retina and modulate opsins. GRK2, GRK3, GRK5, and GRK6 are ubiquitously expressed, whereas GRK4 is expressed in only a few organs (vide infra).

**GRKs and Hypertension**

GRKs have multiple physiological effects on the regulation of blood pressure. Vascular smooth muscle overexpression of GRK2 in transgenic mice attenuates β-adrenergic receptor–induced vasodilation and increases resting blood pressure. GRK2 expression and GRK activity are increased in both the lymphocytes and vascular smooth muscles of patients with essential hypertension and in SHR. GRK2 hemizygous knockout mice have increased nitric oxide bioavailability that protects against Ang II–induced hypertension. The transgenic mice with vascular smooth muscle–specific GRK5 overexpression are hypertensive. By contrast, GRK3 expression in human lymphocytes has been reported to be inversely correlated with blood pressure, suggesting a protective role for GRK3 in the regulation of blood pressure that is supported by the findings in transgenic mice. Overexpression of GRK2, GRK3, and GRK5 in human embryonic kidney (HEK293) cells desensitizes the D₃R. Inhibition of GRK6 prevents intestinal D₃R desensitization. However, renal GRK6 levels are lower in hypertensive subjects and SHR than their normotensive controls.

**Role of GRK4 in Hypertension**

**GRK4 Isoforms**

GRK4 has some inherent characteristics. For example, GRK4 has constitutive activity under basal conditions, which may, in part, be because of its ability to bind to inactive Gα and Gtα subunits. It is the only GRK subtype that is capable of phosphorylating unstimulated GPCRs. GRK4 is expressed in a limited number of tissues, for example, artery, bone, cerebellum, heart, kidney, myometrium, small intestines, and testes, unlike GRK2, GRK3, GRK5, and GRK6, which are ubiquitously expressed. Moreover, 4 splice variants (GRK4α, β, γ, and δ) of GRK4 have been identified in humans. Alternative splicing generates 4 isoforms of human GRK4 mRNA that differ in the presence or absence of exon 2 and exon 15: GRK4α is the longest isoform and contains all of the 16 exons; GRK4β, which lacks exon 2, has a 32-codon-deletion that encompasses the phosphatidylinositol bisphosphate–binding domain near the amino terminus; GRK4γ, which lacks exon 15, has a 46-codon-deletion near the carboxyl terminus; and GRK4δ lacks both exons 2 and 15. The human GRK4 gene locus at 4p16.3 is linked to hypertension. Numerous studies show that abnormal GRK4 function has the potential to affect GPCR (such as D₁R, D₃R, and AT₁R)-regulated biological responses in many physiological and pathological conditions, such as hypertension, which makes GRK4 as an attractive candidate for a genetic determinant for essential hypertension.

**Distribution of GRK4 in Kidney and Artery**

GRK4 is expressed in the rat renal cortex. In both WKY and SHR, GRK4 is expressed in the subapical membranes of the RPT, thick ascending limb of Henle, and renal artery, with much less expression in the glomerulus. Renal cortical GRK4 expression is increased in SHR compared with WKY rats, whereas cardiac GRK4 expression is similar in the 2 rat strains, indicating that the increased GRK4 expression in hypertension has organ specificity. In mice, the renal expression of GRK4 is strain-dependent and influenced by salt intake, for example, lower on normal but higher on high-salt diet in C57BL/6J than in S/JL/J mice. C57BL/6 mice are salt-sensitive and have an impaired ability to excrete a NaCl load that is associated with an increase in blood pressure, whereas S/JL/J mice are salt-resistant. All 4 GRK4 isoforms are expressed abundantly in human RPT cells; GRK4 is localized at the RPT cell surface membrane and cytoplasm and internalized after stimulation of dopamine receptors.

GRK4 activity is increased in kidneys of hypertensive humans. Antisense GRK4 oligonucleotides completely blocked the constitutive serine phosphorylation of the D₁R and restored the ability of the D₁-like receptor agonist, fenoldopam, to stimulate cAMP accumulation in RPT cells from hypertensive subjects, which suggests that the major GRK involved in the phosphorylation of the D₁R in hypertension is GRK4 and not the other GRKs. However, there are no differences in the expression of the GRK4 isoforms (α/β, γ/δ) in kidneys or cultured RPT cells between hypertensive and normotensive subjects. Therefore, we assume that the increased activity of GRK4 in the kidney of hypertensive subjects is not caused by increased renal GRK4 protein expression but rather by constitutively active variants of GRK4.

In Sprague–Dawley rats and C57BL/6J mice, GRK4 is well-expressed in large and small arterial vessels, including the carotid arteries, thoracic aorta, superior mesenteric artery, and renal artery. In the aorta, GRK4 is expressed in the tunica media and adventitia. However, removal of the adventitia does not affect the Ang II–mediated vasoconstriction, suggesting that GRK4 in the adventitia does not participate in Ang II–mediated vasoconstriction.
Regulation of GRK4

As a regulator of GPCRs, GRK4 per se is regulated by transcription factors and signaling molecules. The GRK4 promoter region, containing 1851 bp of the 5′-flanking region and 275 bp of the 5′-untranslated region, is reported to be highly active. The GRK4 core promoter resides in the first 1851 bp upstream of its transcription start site, suggesting that the complex DNA–protein and protein–protein interaction patterns at this portion may affect the transcriptional and expression capacities of GRK4. The transcription factor c-Myc, binding to the promoter of GRK4, positively regulates GRK4 protein expression in human RPT cells, which connects aberrant Ang II activation to D1R–adenylyl cyclases uncoupling. The GRK4 subfamily, including GRK4, is potently inhibited by ubiquitous calcium-binding protein calmodulin, which has little or no effect on members of other GRK subfamilies.

Sorting nexins are involved in receptor endocytosis and trafficking through the endosomes. Sorting nexin 5 directly interacts with GRK4 and prevents GRK4 from targeting the trafficking through the endosomes. Sorting nexin 5 selectively depletes GRK4 in renal cortical membranes of SHRs than WKY rats. Silencing of renal cortical GRK4 decreases serine-phosphorylated D3R levels are much higher in renal cortical membranes of SHRs than WKY rats. Silencing of renal cortical GRK4 decreases serine-phosphorylated D3R to a greater extent in SHRs than WKY rats. Depletion of renal cortical GRK4 also increases urine flow and sodium excretion and attenuates the increased blood pressure in SHRs, but does not affect the blood pressure in WKY rats.

In vitro studies also support the regulation of D3R by GRK4. GRK4 constitutively phosphorylates the D3R in the absence of agonist activation, whereas depletion of GRK4 blunts the D3R desensitization. Moreover, increased activity of GRK4 because of constitutively active GRK4 variants, not GRK4 protein abundance, causes the decrease in D3R function in RPT cells from hypertensive subjects, which is restored by GRK4 depletion. All 4 GRK4 isoforms are expressed in RPT cells. GRK4α and not the other GRK4 isoforms can phosphorylate D3R in certain HEK293 (eg, HEK293T) cells, whereas GRK4γ can phosphorylate D3R in Chinese hamster ovary cells and HEK293 that do not stably express the SV40 large T antigen (J.J. Gildea, P.A. Jose, R.A. Felder, unpublished data, 2015). The phosphorylation of D3R is regulated by both GRK4α and GRK4γ (GRK4γ>GRK4α) isoforms in human RPT cells. These results indicate that the regulatory activity of GRK4 isoforms on the desensitization of dopamine receptors is cell-specific. It should also be noted that the effect of human GRK4α variant on blood pressure in mice has not been studied, whereas the effect of human GRK4γ wild-type and variants on blood pressure has been reported in transgenic mice.

Similar to D1Rs, the AT1R is also regulated by GRK4. Our previous studies have shown that AT1R expression and actions are enhanced in both kidney and artery in transgenic mice expressing GRK4 variants. The increased renal GRK4 expression in old rats is also associated with increased renal AT1R expression and function. Interestingly, the renal-selective depletion of AT1R in SHRs does not decrease blood pressure because the interruption of the renin–angiotensin negative feedback loop results in increased circulating renin and Ang II. However, combined renal-selective silencing of both GRK4 and AT1R decreases blood pressure, as well as plasma renin activity and Ang II levels in both rat strains, which can explain, in part, the crucial role of GRK4 in the regulation of sodium excretion and blood pressure.

Regulation of Blood Pressure by GRK4 Variants

Three missense single nucleotide polymorphisms in the coding region of GRK4γ impair D3R function (Table 1). They are nucleotide 448, CGT to CTT (amino acid 65R>L, rs2960306); nucleotide 679, GCC to GTC (amino acid 142A>V, rs1024323); and nucleotide 1711, GCG to GTG (amino acid 275 V>M, rs1801058). GRK4γ single nucleotide polymorphisms (65R>L, 142A>V, 486A>V) markedly impair D3R-mediated cAMP accumulation that is not caused by differences in the quantity of the expression of either D3R or GRK4γ. Human GRK4γ (hGRK4γ) 142V transgenic mice have higher blood pressures and greater heart weights than hGRK4γ wild-type transgenic mice. Infusion of fenoldopam, a D1-like receptor agonist, increases urine flow and sodium excretion in hGRK4γ wild-type mice, but not in the hypertensive hGRK4γ 142V transgenic mice. Additional studies showed that the higher blood pressure in hGRK4γ 142V transgenic mice is not because of the transgene integration sites, flanking genes, or copy numbers, but due mainly to the effect of hGRK4γ 142V transgene acting via D1R. This is confirmed by in vitro studies: in the transfected Chinese hamster ovary cells, GRK4γ 142V increases GRK activity and causes D3R phosphorylation, which may explain, in part, the decreased responsiveness of the D3R in hypertensive hGRK4γ 142V transgenic mice. There is, however, cell specificity of the ability of GRK4γ to regulate D3R function; GRK4γ regulates D3R function in human RPT, Chinese hamster ovary cells, and HEK293 cells (vide supra), whereas it is GRK4α in HEK293T cells. We also found that the function of D3R is also impaired in the hGRK4γ 142V-transfected human RPT cells (J. Yang, V.A.M. Villar, and P.A. Jose, unpublished data, 2015).

The hypertension in hGRK4γ 142V transgenic mice is also associated with increased AT1R expression and function in both kidney and artery. GRK4γ 142V transgenic mice have increased renal AT1R expression and function, for example, increased blood pressure response to Ang II infusion and AT1R blockade. By contrast, GRK4γ 142V transgenic mice that are deficient of AT1R have normal blood pressure. We have reported that both AT1R expression and AT1R-mediated


Table 1. GRK4 Variant Transgenic Mice and In Vitro Studies

<table>
<thead>
<tr>
<th>GRK4 Variants</th>
<th>Mouse Phenotype</th>
<th>Functional Deficit(s)</th>
<th>Receptor Defect(s)</th>
<th>Variants Transduced Cell Studies</th>
</tr>
</thead>
<tbody>
<tr>
<td>R65L</td>
<td>Salt-sensitive hypertension (high-salt diet)</td>
<td>Not determined</td>
<td>Not determined</td>
<td>CHO</td>
</tr>
<tr>
<td>A142V</td>
<td>Hypertension (normal-salt diet)</td>
<td>Decreased urine flow and sodium excretion</td>
<td>Decreased renal expression and responsiveness of D₁R</td>
<td>CHO</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td>Increased AT₁R expression and responsiveness</td>
<td>Human RPT cells</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td>Increased AT₁R expression and responsiveness</td>
<td>Vascular smooth muscle cells</td>
</tr>
<tr>
<td>A486V</td>
<td>Salt-sensitive hypertension (high-salt diet)</td>
<td>Impaired pressure-natriuresis plot</td>
<td>Increased AT₁R expression</td>
<td>CHO</td>
</tr>
</tbody>
</table>

AT₁R indicates angiotensin type 1 receptor; CHO, Chinese hamster ovary; D₁R, D₁-like receptors; GRK4, GPCR kinase 4; and RPT, renal proximal tubules.

Vasoconstriction are higher in the aorta of hGRK4γ 142V transgenic mice. Moreover, infusion of Ang II causes a greater increase in blood pressure, whereas infusion of the AT₁R antagonist candesartan causes a greater decrease in blood pressure in hGRK4γ 142V transgenic mice than GRK4 wild-type transgenic mice. AT₁R mRNA and protein expression and function are higher in hGRK4γ142V than in GRK4γ wild-type cells, but the opposite is true for AT₁R phosphorylation and degradation, indicating that the regulation of AT₁R expression by hGRK4γ occurs at both transcriptional and post-translational levels.

Depending on the genetic background and sodium intake, hGRK4γ 486V transgenic mice may develop increased blood pressure. hGRK4γ 486V transgenic mice have increased renal AT₁R expression and develop hypertension only after an increase in sodium intake, in contrast to GRK4γ 142V transgenic mice, which have increased AT₁R expression and develop hypertension even on a normal-salt diet. Depending on the genetic background of the mouse, hGRK4γ wild-type prevents salt-sensitive hypertension, whereas hGRK4γ 486V converts a salt-resistant phenotype to a salt-sensitive phenotype. hGRK4γ 65L transgenic mice, similar to hGRK4γ 486V transgenic mice, have normal blood pressure on a normal-salt diet, but have increased blood pressure on high-salt diet (L.D. Asico and P.A. Jose, unpublished data, 2015). In vitro studies showed that in single (65L or 486V) or double variant GRK4 (65L/486V)-transfected Chinese hamster ovary cells, there is an increase in basal D₁R phosphorylation and impairment of D₁R-mediated cAMP production. The mechanism for the increase in blood pressure in GRK4γ 65L or 486V transgenic mice only when salt intake is increased remains unclear.

GRK4 Polymorphisms and Hypertension

The GRK4 gene polymorphisms have different allele frequencies among different populations. GRK4 486V is more frequent in Asians and less frequent in blacks than in other populations (Hispanics and whites). The GRK4 locus on human chromosome 4p16.3 is linked to essential hypertension and salt sensitivity. The first report in 2002 by Bengra et al found a significant association between GRK4 A486V variant and an Italian population of mildly hypertensive patients. Subsequent studies showed that GRK4 gene variants R65L, A142V, and A486V are each associated with essential hypertension in several ethnic groups (Table 2). In Euro-Australians, GRK4 486V is associated with essential hypertension, whereas the 65L and 142V variants track with elevation in diastolic blood pressure only in male hypertensives. In a Chinese Han population, GRK4 A486V is also associated with hypertension in additive, dominant, and recessive model, whereas GRK4 142V is associated with hypertension in an additive model only. However, there are reports that do not show the association between GRK4 variants and hypertension. In a population of blacks 18 to 49 years of age, GRK4 A486V variant was found to be negatively associated with hypertension. Although the reasons leading to the differences among studies are not known, the negative studies could be the consequence of not taking into account salt sensitivity (particularly for GRK4 R65L and A486V) or assessing the role of GRK4 in conjunction with other single nucleotide polymorphisms of GRK4 and other genes.

Salt-sensitive hypertension is associated with GRK4 gene variants. The Italian patients whose hypertension is associated with GRK4 A486V are actually salt-sensitive. In a Japanese population, the GRK4 variants (R65L, A142V, and A486V) are more frequent in salt-sensitive than salt-resistant Japanese hypertensive patients. A genetic model of GRK4 R65L, A142V, and A486V is 94.4% predictive of salt sensitivity. By contrast, the single-locus model with only GRK4 A142V is 78.4% predictive, whereas a 2-locus model of GRK4 A142V and A486V is 94.4% predictive of salt sensitivity.
Hypertension and aldosterone synthase CYP11B2 is 77.8% predictive of low-renin hypertension.57 The ability to excrete a salt load is inversely related to the number of GRK4 variant alleles (R65L, A142V, and A486V) in hypertensive Japanese.57 GRK4 variants are also associated with salt sensitivity in normotensive subjects.55–57 In black normotensive adolescents, the GRK4 65L allele is associated with a reduced urinary sodium excretion in response to stress.56 In young normotensive twins, GRK4 65L was associated with the steepest increase in blood pressure; the GRK4 65L-142V-A486V haplotype had a 1.05 mm Hg steeper increase in systolic blood pressure per year increase in age, relative to those with GRK4 R65, A142, and A486 haplotype.54 Therefore, genetic variations of GRK4 may contribute to variations of blood pressure in normotensive individuals, potentially influencing the development of hypertension.

As indicated earlier, GRK4 variants interact with other genes in the pathogenesis of hypertension. Multilocus analyses have shown association between GRK4 variants and other gene variants with high blood pressure. In an African population from Ghana, the combination of angiotensin-converting enzyme and GRK4 R65L is the best genetic model to predict hypertension (70.5% prediction of hypertension).53 Among Japanese subjects, the best combination that is predictive of hypertension, not classified according to salt sensitivity, is GRK4, angiotensin-converting enzyme, and CYP11B2, with an estimated prediction success of 63%; however, for low-renin hypertension in Japanese, the single best genetic model includes only GRK4 A142V and CYP11B2, with an estimated prediction success of 77.8%.57,66 Normotensive Japanese with GRK4 polymorphisms were reported to have increased N-terminal pro-B-type natriuretic peptide levels.65 A recent study among African-derived Brazilian populations reported that an interaction between GRK4 A486V and endothelial nitric oxide synthase is associated with increased diastolic blood pressure.62

There are currently 2 meta-analyses on the associations of GRK4 polymorphisms with hypertension risk. Our previous meta-analysis showed that GRK4 486V increases the risk for essential hypertension with an odds ratio of 1.5 (95% confidence interval, 1.2–1.9).7 A more recent meta-analysis showed that GRK4 486V is inversely associated with hypertension among East Asians (odds ratio =0.39, 95% confidence interval, 0.28–0.55), but positively associated with hypertension among Europeans (odds ratio =2.38, 95% confidence interval, 1.38–4.10); GRK4 65L was also associated with hypertension among Europeans.11

**Table 2. Association Studies of GRK4 Variants in Hypertensive or Normotensive Subjects**

<table>
<thead>
<tr>
<th>GRK4 Variants</th>
<th>Ethnic Group (Year)</th>
<th>Single or Multilocus Analyses</th>
<th>Hypertension Phenotype</th>
<th>Ethnic Group (Year)</th>
<th>Single or Multilocus Analyses</th>
<th>Blood Pressure or Sodium Excretion</th>
</tr>
</thead>
</table>

ACE indicates angiotensin-converting enzyme; GRK4, GPCR kinase 4; NOS, nitric oxide synthase; SBP, systolic blood pressure; and UNaV, urinary sodium excretion.

ACE indicates angiotensin-converting enzyme; GRK4, GPCR kinase 4; NOS, nitric oxide synthase; SBP, systolic blood pressure; and UNaV, urinary sodium excretion.
is the absence of some genes in the chips. For example, GRK4 was not linked to hypertension in the GWAS probably because the GRK4 polymorphisms are not consistently present in the Affymetrix and Illumina chips; GRK4 65L, 142V, and 486V are present only in the Illumina Human 1 mol/L beadchip, and GRK4 142V is not present in any of the Affymetrix Chips. These limitations may also be found in GWAS for pharmacogenomics of essential hypertension.\(^{69}\)

There are several classes of antihypertensive medicines used in clinic and include calcium channel blockers, drugs that target the RAS (angiotensin-converting enzyme inhibitors, angiotensin receptor blockers, direct renin inhibitor), diuretics (thiazides and thiazide-like diuretics, loop diuretics, potassium sparing diuretics), and drugs that target the sympathetic nervous system (α-adrenergic receptor blockers, α-adrenergic receptor blockers, direct vasodilators, central α-adrenergic receptor agonists, and adrenergic depleters [eg, reserpine, probably not used any longer]). The treatment of hypertension is currently not based on pharmacogenetics, but rather based on consensus guidelines.\(^{76,71}\)

Current evidence shows that the presence or absence of GRK4 gene variants may be important in guiding therapeutic antihypertensive strategies. In the African American Study of Kidney Disease and Hypertension Study, male (but not female) blacks with GRK4 A142 were found to have a faster decrease in blood pressure response to the β₁-adrenoceptor blocker metoprolol, but were less responsive to metoprolol in the presence of GRK4 65L.\(^{73}\) The Pharmacogenomic Evaluation of Antihypertensive Responses study involving hypertensive black and Euro-Americans showed that the presence of GRK4 65L, 142V, and 486V variant alleles in Euro-Americans was associated with reduced response to the β₁-adrenergic receptor blocker, atenolol.\(^{12}\) Our studies in hypertensive Japanese subjects show that GRK4 142V is associated with greater decrease in both systolic blood pressure and diastolic blood pressure in response to angiotensin receptor blockers (H. Sanada and P.A. Jose, unpublished data, 2015), which also normalized the increased blood pressure of GRK4γ 142V transgenic mice.\(^{46}\) The lowering of blood pressure with a decrease in dietary salt intake among South African blacks was observed in those with no GRK4 variants or one variant GRK4 65L or GRK4 142V allele; the effect of GRK4 486V was not evaluated.\(^{8}\) However, in a small cohort of Japanese hypertensive subjects, those with GRK4 486V had a good antihypertensive response to low-salt diet or diuretics.\(^{72}\) Patients homozygous to 65L and 142V have been reported to need more antihypertensive treatment, especially diuretics.\(^{73}\) These results suggest that the presence of GRK4 variant alleles may be important determinants in the blood pressure response to antihypertensive drugs or dietary intervention and risk for adverse cardiovascular events.

**Conclusions and Perspectives**

In summary, increasing evidence shows that constitutively active GRK4 variants, specifically the human GRK4γ 65L, 142V, and 486V variants, play a crucial role in regulating the function of the dopamine receptors and AT₁R and are, therefore, involved in the pathogenesis of hypertension (Figure). Genetic association has been found between GRK4 gene variants and hypertension. Antihypertensive medicines have different responses in patients with individual GRK4 variants, indicating that GRK4 variants may be important in choosing the initial antihypertensive medication. Increased understanding of GRK4 in the genetics and pharmacogenetics of hypertension may provide therapeutic targets for hypertension in the future.

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**Disclosures**

Dr Jose, who is the Scientific Director of Hypogen, Inc, owns US Patent Number 6,660,474 for G protein–related kinase mutants in essential hypertension. The other authors report no conflicts.

**References**


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