

## Angiotensin-II, the Brain, and Hypertension An Update

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Despite enormous therapeutic advancements, the incidence of hypertension continues to rise<sup>1</sup> and remains a medical burden of global proportions.<sup>2</sup> This is likely due, in part, to the complex pathogenesis of hypertension.<sup>1</sup> In addition to pathological processes in peripheral cardiovascular organs, the central nervous system (CNS) has emerged as a major culprit. The renin-angiotensin system (RAS) plays a crucial role in cardiovascular control, and both systemic angiotensin-II (Ang-II) and centrally generated Ang-II signal through complex neural networks to contribute to the development of hypertension (Figure). Although the effects of Ang-II within the brain are multifaceted, several key hypertensive players have emerged, including oxidative stress, endoplasmic reticulum stress, inflammation, and transcription factor activation. The purpose of this review is to provide a brief update on recent work related to Ang-II, the brain, and hypertension with a specific focus on *Hypertension* articles when appropriate.

### Circulating Ang-II, Centrally Generated Ang-II, and Hypertension

Dysregulation of the RAS is common in human hypertension,<sup>3</sup> and several hypertensive animal models have proven particularly useful to understand brain Ang-II, such as genetic, systemic infusion of Ang-II, deoxycorticosterone acetate (DOCA)-dependent salt, and renal injury. Within the CNS, Ang-II promotes a hypertensive state by enhancing sympathetic neural outflow, altering the release of hormones involved in volume, salt, vascular, renal, and cardiac regulation, and modulating inflammatory processes. Emerging evidence also suggests that brain Ang-II may alter bone marrow-derived hematopoietic stem and progenitor cells and thus exacerbate hypertensive vascular pathologies (Figure).<sup>4-6</sup> Circulating Ang-II peptides are too large to cross the blood-brain barrier (BBB) and act at circumventricular organs—specialized CNS regions lacking a well-formed BBB. These unique neural areas act as relay centers through which peripheral Ang-II can influence cardiovascular control via efferent projections to downstream autonomic/neurosecretory centers, including hypothalamic and brain stem regions.<sup>7</sup> However, BBB disruption is common in hypertension<sup>8</sup> and thus provides a means

by which circulating Ang-II could also gain access to neural regions from which it is typically excluded. In support of this, Biancardi et al<sup>9</sup> recently used novel intravascular fluorescent dyes and found increased BBB permeability in key cardio-regulatory regions, including the paraventricular nucleus of the hypothalamus (PVN), nucleus tractus solitarii (nTS), and rostral ventral lateral medulla (RVLM), of spontaneously hypertensive rats (SHR), relative to normotensive controls. Administration of fluorescently labeled Ang-II also demonstrated that circulating Ang-II could gain access to neurons and glial cells within hypothalamic and brain stem regions of hypertensive animals. Interestingly, chronic treatment with the angiotensin type 1 receptor (AT<sub>1</sub>R) antagonist losartan, but not a peripheral vasodilator, prevented the BBB disruptions. These findings raise the possibility that, in addition to acting at circumventricular regions, circulating Ang-II initiates a positive feedback loop through AT<sub>1</sub>R to enhance BBB permeability and subsequently is able to access BBB-protected cardioregulatory neural areas.

In addition to circulating Ang-II, all components of the RAS system are present within the CNS, and local production of brain Ang-II is now well accepted to contribute to hypertension development. The mechanisms by which the CNS generates Ang-II are reviewed in detail elsewhere.<sup>10</sup> Of importance, a key question has focused on how local production of Ang-II can occur when levels of renin, the rate-limiting enzyme, are extremely low. Attention has been drawn to the (pro)renin receptor, a transmembrane receptor that binds renin and its precursor prorenin at nanomolar concentrations. Thus, the (pro)renin receptor could facilitate extracellular and intracellular Ang-II production, likely through nonproteolytic activation of prorenin.<sup>11,12</sup> In support of this, the (pro)renin receptor is widely expressed in neural cardioregulatory regions<sup>13,14</sup> and is significantly upregulated in the brain of hypertensive animals.<sup>13</sup> More recently, Li et al<sup>15</sup> demonstrated in renin-angiotensinogen double-transgenic hypertensive mice that selective knockdown of the (pro)renin receptor in the subfornical organ (SFO), a circumventricular organ crucial for cardiovascular regulation, blunted hypertension, and cardiac/vasomotor sympathetic tone, improved baroreflex sensitivity and reduced vasopressin levels.

Received May 13, 2015; first decision May 31, 2015; revision accepted August 6, 2015.

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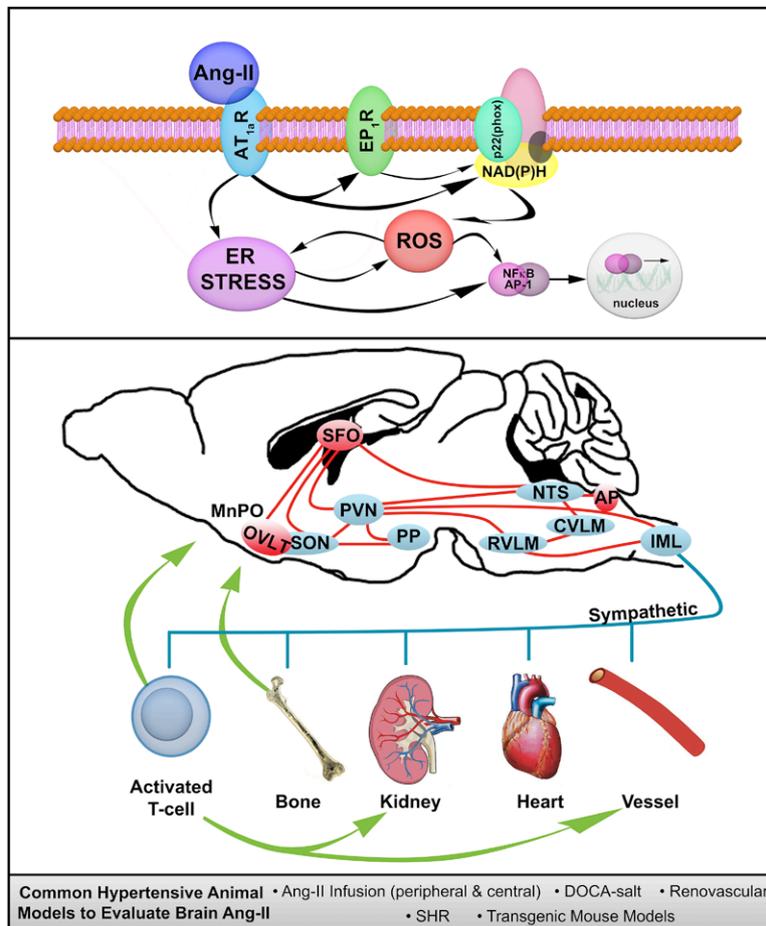
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(*Hypertension*. 2015;66:920-926. DOI: 10.1161/HYPERTENSIONAHA.115.03624.)

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*Hypertension* is available at <http://hyper.ahajournals.org>

DOI: 10.1161/HYPERTENSIONAHA.115.03624



**Figure.** Simplified schematic illustrating the signaling pathways (**top** box), as well as neural networks and sympathetic nervous system influenced physiological outputs (**middle** box) involved in the development of hypertension due to peripherally or locally generated angiotensin-II (Ang-II) action in the brain. Common animal models used to evaluate Ang-II, the brain, and hypertension are also highlighted (**bottom** box). See text for additional details. AP indicates area postrema; AP-1, activator protein-1; AT<sub>1a</sub>R, angiotensin type 1a receptor; CVLM, caudal ventral lateral medulla; DOCA, deoxycorticosterone acetate; ER, endoplasmic reticulum; EP<sub>1</sub>R, prostaglandin E receptor 1; IML, intermediolateral nucleus; MnPO, median preoptic nucleus; NAD(P)H, nicotinamide adenine dinucleotide phosphate; NTS, nucleus tractus solitarius; NF- $\kappa$ B, nuclear factor- $\kappa$ B; OVL, organum vasculosum lamina terminalis; PP, posterior pituitary; PVN, paraventricular nucleus of the hypothalamus; ROS, reactive oxygen species; RVLM, rostral ventral lateral medulla; SFO, subfornical organ; SHR, spontaneously hypertensive rat; and SON, supraoptic nucleus.

Using DOCA salt, a model that induces elevations in brain RAS activity and Ang-II signaling, the same group has also shown that neuron-specific knockout of the (pro)renin receptor attenuates DOCA salt-induced hypertension and sympathetic activation, concomitant with a reduction in brain Ang-II production.<sup>11</sup> Furthermore, a recently developed (pro)renin antagonist has been shown to attenuate DOCA salt-mediated increases in blood pressure and brain Ang-II, highlighting the therapeutic potential in targeting brain pro(renin)-mediated Ang-II production in hypertensive conditions.<sup>16</sup>

Whether centrally or peripherally generated, the hypertensive actions of brain Ang-II occur primarily via AT<sub>1</sub>R. Indeed, Cre-LoxP-mediated removal of AT<sub>1</sub>R from catecholaminergic cells blunts Ang-II-dependent hypertension and sympathetic modulation of blood pressure.<sup>17</sup> These data suggest that AT<sub>1</sub>R in this cell population, presumably within sympathetic premotor neurons, the adrenal medulla, and sympathetic ganglia, is involved in the hypertensive actions of Ang-II, although a role of AT<sub>1</sub>R in peripheral cell populations, such as T lymphocytes, cannot be excluded from these findings. Additional

support for brain AT<sub>1</sub>R signaling has also been demonstrated in a murine DOCA salt model. Hilzendeger et al<sup>18</sup> showed that selective knockdown of AT<sub>1</sub>R in the SFO blunted elevations in arterial blood pressure, reduced sympathetic cardiac modulation and urine vasopressin markers (copeptin), and positively affected fluid and salt homeostasis. Collectively, these recent findings build on existing evidence and provide additional insight into specific CNS nuclei and cell populations involved in the development of hypertension due to brain Ang-II-AT<sub>1</sub>R signaling.

Although the primary actions of AT<sub>1</sub>R are prohypertensive, as highlighted by the aforementioned studies, it is important to consider the neural region in which Ang-II-induced signaling occurs. The nTS, located in the dorsomedial aspect of the brain stem, is a key example. Stimulation of excitatory nTS projections activate inhibitory neurons in the caudal ventrolateral medulla, in turn inhibiting sympathoexcitatory neurons in the RVLM, resulting in decreases in arterial blood pressure. Indeed, recent observations indicate that AT<sub>1</sub>R expression is elevated in the nTS of SHR and small hairpin RNA knockdown

of AT<sub>1</sub>R in the nTS of hypertensive animals resulted in an approximate 30 mmHg increase in mean arterial pressure.<sup>19</sup> These findings suggest that nTS Ang-II–AT<sub>1</sub>R signaling exerts counterhypertensive actions. While outside of the context of this brief review, the compensatory actions of AT<sub>2</sub>R<sup>20</sup> and alternative RAS pathways (angiotensin I–converting enzyme 2/Ang-1–7/Mas)<sup>21</sup> should also be considered.

### Ang-II, Brain Oxidative Stress, and Hypertension

Over the past 20 years, a wealth of evidence has emerged implicating Ang-II–induced reactive oxygen species (ROS) generation in the pathogenesis of hypertension. Although several sources for ROS exist, NAD(P)H (nicotinamide adenine dinucleotide phosphate) oxidase is currently viewed as the predominant source of Ang-II–derived ROS production in the brain.<sup>22</sup> Several recent reports extend our understanding of ROS in the CNS in mediating the hypertensive actions of Ang-II. Building on previous findings demonstrating an obligatory role of ROS in the SFO in mediating Ang-II–dependent hypertension,<sup>23–25</sup> Lob et al<sup>26</sup> used a Cre-LoxP approach to ablate the NAD(P)H oxidase docking subunit p22(phox) from the SFO in a murine model. In response to low-dose infusion of Ang-II, removal of SFO p22(phox) prevented ROS production in the SFO and eliminated the hypertensive response; further demonstrating that NAD(P)H oxidase–derived ROS in the CNS promote Ang-II–dependent hypertension.<sup>24,25,27</sup> In line with these findings, overexpression of the (pro)renin receptor in neuronal cultures and elevations in Ang-II production promotes an increase in ROS that is dependent on NAD(P)H oxidase activity, although Ang-II–independent mechanisms also contribute.<sup>28</sup>

Parallel investigations have focused on additional mechanisms mediating Ang-II–induced oxidant production in the CNS. Cyclooxygenase (COX)-derived prostanoids have been implicated in the development of hypertension, although the source and site of action remain incompletely defined. In this regard, we recently demonstrated crosstalk between brain prostaglandin E<sub>2</sub>, its receptor EP<sub>1</sub> and ROS signaling in the development of Ang-II–induced hypertension. In response to systemic Ang-II infusion, hypertension and SFO-ROS production were abolished in mice with null mutations in the EP<sub>1</sub> receptor and COX-1 (the enzymatic source of prostaglandin E<sub>2</sub>), but not COX-2.<sup>29</sup> Genetic reconstitution of the EP<sub>1</sub> receptor selectively in the SFO was sufficient to restore these responses demonstrating that COX-1–derived prostaglandin E<sub>2</sub> ROS generation in the forebrain SFO is required for Ang-II–dependent hypertension.<sup>29</sup> Consistent with this, treatment with a selective COX-1 inhibitor (but not COX-2 inhibitor) has been shown to attenuate hypertension development and increases in whole body norepinephrine spillover in response to Ang-II infusion in rats on a high salt diet.<sup>30</sup> At the cellular level, recent *in vitro* findings in SFO cells suggest a complex signaling cascade in which Ang-II, via AT<sub>1</sub>R, promotes phospholipase A<sub>2</sub>–induced production of arachidonic acid.<sup>31</sup> Arachidonic acid is then metabolized to prostaglandin E<sub>2</sub> by COX-1 and acts on the EP<sub>1</sub> receptor (most likely in a paracrine fashion) to cause intracellular Ca<sup>2+</sup> release. Subsequent elevations in intracellular Ca<sup>2+</sup> promote NAD(P)H oxidase–derived

ROS production and activation of voltage-gated Ca<sup>2+</sup> channels, resulting in increases in neuronal firing.<sup>31</sup> Together, these recent findings, from the cellular to whole animal level, provide in depth insight into the mechanisms by which brain Ang-II–induced oxidative stress promotes hypertension development.

### Ang-II, Brain Endoplasmic Reticulum Stress, and Hypertension

The endoplasmic reticulum (ER) is a reticulated organelle specialized in the synthesis, folding, assembly, and modification of proteins. The ER also plays an important role in various cellular signaling processes, and ER homeostasis is strictly maintained in the face of a broad spectrum of potential stressors. However, prolonged periods of cell stress, such as those induced by Ang-II (eg, alterations in cellular redox status and Ca<sup>2+</sup> levels), can lead to an accumulation of un/misfolded proteins in the ER, triggering an adaptive signaling cascade, and the unfolded protein response (UPR). Although initially a compensatory attempt to maintain cellular homeostasis, sustained activation of the UPR is now thought to be the basis of several chronic diseases.<sup>32</sup> In this regard, we have recently shown that brain ER stress mediates the development of Ang-II–dependent hypertension.<sup>33</sup> In response to low-dose Ang-II infusion, robust UPR upregulation and ER morphological alterations were found in the CNS, particularly within the SFO. CNS delivery of a chemical ER stress inhibitor (tauroursodeoxycholic acid) or genetic inhibition of ER stress selectively in the SFO abolished the development of Ang-II–induced hypertension. Interestingly, inhibition of SFO-ER stress also prevented Ang-II–mediated elevations in ROS, suggesting that Ang-II–induced ER stress is a source of oxidative stress.<sup>33</sup>

In line with these findings, Chao et al<sup>34</sup> found upregulation of the UPR in the RVLM of SHR or in response to intracisternal infusion (which globally targets medullary regions) of Ang-II in control animals. Intracisternal infusion of salubrinal, a pharmacological agent that reduces ER stress, resulted in a reduction in arterial blood pressure in SHR. Interestingly, RVLM-targeted administration of the ROS scavenger tempol lessened Ang-II–induced and SHR-induced UPR activation, suggesting that ROS may also act upstream of ER stress.<sup>34</sup> Several recent observations have also suggested a role of ER stress in mediating cardiac damage, endothelial dysfunction, and aortic stiffening during Ang-II–induced hypertension.<sup>35,36</sup> It is important to note that although these investigations were not focused on the CNS, the results are based around systemic administration of ER stress–reducing agents that will act not only within peripheral regions but also within the brain, particularly at BBB-deficient regions (eg, SFO). Although yet to be determined, given the importance of the CNS in these pathological processes,<sup>37</sup> a role of brain ER stress cannot be excluded. Collectively, these findings highlight Ang-II–induced ER stress in the brain as a novel paradigm underlying the development of hypertension.<sup>38</sup> However, it is important to consider the model of hypertension that is investigated, as Jo et al<sup>39</sup> recently reported that relief of brain ER stress (with chemical chaperones) attenuated DOCA salt–induced

elevations in saline intake and heart rate, with no influence on the pressor response. Thus, it is plausible that brain ER stress may play a role in certain forms of hypertension (eg, systemic Ang-II and genetic models), but not others (eg, DOCA salt-dependent). This possibility will require further comparative investigation in various models of hypertension.

### Ang-II, Brain Inflammation, and Hypertension

In addition to modulation of the autonomic nervous system and volume homeostasis, the CNS actions of Ang-II promote activation of the immune system. This area of research is particularly intriguing, given the emerging role of the immune system and inflammation in the pathogenesis of hypertension.<sup>40,41</sup> During low-dose infusion of Ang-II in mice, lesioning of the anteroventral third cerebral ventricle region concomitantly prevents peripheral vascular infiltration of activated T cells and hypertension development,<sup>42</sup> supporting the concept of brain Ang-II signaling in the regulation of peripheral inflammation. A key link between CNS Ang-II and immune system activation seems to be oxidative stress, particularly within the SFO. Indeed, selective SFO removal of extracellular superoxide dismutase, to induce elevations in ROS, resulted in exaggerated elevations in circulating activated T cells and vascular infiltration of inflammatory cells during Ang-II-induced hypertension.<sup>43</sup> In line with this, ablation of the NAD(PH) oxidase subunit p22(phox) from the SFO prevents Ang-II-induced inflammatory responses in the peripheral vasculature.<sup>26</sup>

Although the precise mechanisms by which brain Ang-II affects the peripheral immune system remain incompletely defined, as detailed elsewhere,<sup>40,41,44–46</sup> enhanced sympathetic outflow or removal of suppressive parasympathetic influence lymphoid organs, direct adrenergic and cholinergic modulation of immune cell populations, or hypertension itself may all contribute. Building on previous observations of elevations in splenic sympathetic nerve activity and inflammatory cytokine expression in response to central Ang-II administration,<sup>47</sup> a series of recent experiments from Raizada's group indicates a link between CNS Ang-II, sympathetic input to bone marrow, and inflammatory responses. Central scavenging of mitochondrial ROS, with a superoxide dismutase mimetic, prevented Ang-II-induced increases and decreases in circulating bone marrow-derived inflammatory and endothelial progenitor cells (vascular reparative cells crucial for repair and maintenance of the endothelium), respectively. In addition, retrograde neuronal labeling demonstrated a brain to bone marrow connection, with efferent inputs arising from several cardioregulatory regions, including the PVN, RVLM, nTS, and SFO.<sup>6</sup> More recently, parallel elevations in sympathetic nerve activity to bone and an imbalance in inflammatory and endothelial progenitor cells were found in SHR compared with normotensive control rats,<sup>5</sup> although the role of Ang-II in these processes remains yet unknown. Collectively, these novel findings suggest that Ang-II-mediated increases in ROS within the CNS may contribute, in part, to peripheral inflammatory responses, while inhibiting vascular reparative processes, through efferent signaling to bone marrow.

It is important to consider that along with a CNS-immune system connection, immune system-CNS communication

also plays a role in Ang-II-induced hypertension. For example, acute injection of the proinflammatory cytokine, tumor necrosis factor- $\alpha$ , into the PVN induces elevations in arterial blood pressure, as well as lumbar and splanchnic sympathetic nerve activity.<sup>48</sup> Furthermore, chronic infusion of Ang-II promotes microglia activation and proinflammatory cytokine production in the PVN; responses that are prevented with central administration of an anti-inflammatory antibiotic, PVN-selective overexpression of the anti-inflammatory cytokine interleukin-10, or scavenging of mitochondrial ROS.<sup>6,49</sup> Interestingly, inhibition of central Ang-II-induced inflammatory responses has also been shown to correct the imbalance in circulating inflammatory and endothelial progenitor cells.<sup>6</sup> In addition to inflammatory activation within cardioregulatory nuclei, Waki et al<sup>50</sup> have theorized that activation of AT<sub>1</sub>R on cerebrovascular endothelial cells may promote vascular inflammation in the brain stem, thus contributing to the development of hypertension through altered vascular-neuronal signaling. Although the precise interplay remains unclear, the overall evidence to date suggests that a positive feedback loop likely exists in which Ang-II promotes inflammatory cascades with the brain, which subsequently promote activation of the peripheral immune system. In turn, activated circulating inflammatory cells may then feedback to the brain further perpetuating the process. In support of this, proinflammatory bone marrow-derived cells have been shown to migrate into the PVN of SHR,<sup>51</sup> whereas Ang-II-induced hypertension is associated with T-cell infiltration in the SFO.<sup>52</sup> In addition, recent intriguing findings indicate that acute microinjection of proinflammatory cytokines (tumor necrosis factor- $\alpha$  and interleukin-1 $\beta$ ) into the SFO induced elevations in renal sympathetic nerve activity and arterial blood pressure—responses that were prevented by blocking the RAS or COX-2. Moreover, SFO-targeted proinflammatory cytokine microinjections were associated with increases in RAS and COX-2 activity in the SFO and PVN, suggesting that brain inflammation may facilitate the hypertensive actions of brain Ang-II.<sup>53</sup>

### Ang-II, Brain Transcription Factors, and Hypertension

In order for CNS Ang-II signaling to cause sustained elevations in arterial blood pressure, transcription of genes that alter neuronal structural and functional properties is necessary.<sup>54</sup> The inducible transcription factors, nuclear factor- $\kappa$ B (NF- $\kappa$ B) and activator protein 1 (AP-1), have been shown to be strongly associated with ROS, ER stress, and inflammatory responses. Although *in vitro* examinations have clearly shown a role for these 2 transcription factors in neuronal signaling,<sup>54</sup> translation of these findings to the *in vivo* hypertension setting has been limited.

After 2-week<sup>55</sup> or 4-week<sup>56</sup> systemic infusion of Ang-II in rats, upregulation of NF- $\kappa$ B subunits, as well as increased activity of the p65 subunit, was found in the PVN; responses that were prevented by CNS antagonism of AT<sub>1</sub>R or scavenging of superoxide with intracerebroventricular administration of losartan or tempol, respectively.<sup>56</sup> Pharmacological<sup>56</sup> and PVN-targeted genetic inhibition of NF- $\kappa$ B<sup>55</sup> attenuated the

development of Ang-II–induced hypertension, suggesting a functional role of this transcription factor, although further studies are warranted to determine the extent of NF- $\kappa$ B activation in the PVN prior to the development of hypertension. In this context, we recently used the novel technique of in vivo bioluminescence imaging to track temporal activation of NF- $\kappa$ B in the brain during the development of hypertension.<sup>57</sup> In brief, this technique capitalizes on luciferase-dependent light emission and allows for in vivo determination of transcription factor activation in the same animal over time.<sup>58,59</sup> Using this approach, an AT<sub>1a</sub>R-, ROS-, and ER stress–mediated prehypertensive surge in NF- $\kappa$ B activation was found in the SFO during the development of Ang-II–induced hypertension.<sup>57</sup> Although yet to be completely understood, collectively these data indicate that a complex NF- $\kappa$ B neural network is involved in Ang-II–mediated hypertensive actions, with initial activation occurring in circumventricular organs that are subsequently followed by downstream activation in other cardiovascular regulatory regions.

In addition to NF- $\kappa$ B, Burmeister et al<sup>58</sup> examined a role for the transcription factor AP-1 in the PVN in a murine 2-kidney-1-clip model of hypertension, which is associated with activation of the brain RAS.<sup>60</sup> Temporal in vivo bioluminescence imaging evaluation revealed a progressive redox-dependent rise in PVN AP-1 activity that surged within 5 days and remained elevated for up to 2 weeks after renal artery clipping, whereas dominant-negative inhibition of AP-1 transcriptional activation in the PVN prevented renovascular hypertension. Collectively, these studies highlight the importance of CNS transcription factor activation in models of hypertension dependent on brain Ang-II, although future studies are needed to dissect out additional neural regions and pathways involved.

### Summary

Great strides have been made in recent years to advance our understanding of Ang-II actions in the brain in mediating neurogenic hypertension. It is now evident that peripherally and locally generated Ang-II promote a hypertensive state through influences on redox, ER, inflammatory, and transcription factor pathways in the brain (Figure). However, an integrated and complete understanding of these pathological processes is still lacking. Moving forward, novel approaches and techniques will be necessary to further our understanding of Ang-II, the brain, and hypertension, such as intricate neural network analyses, temporal evaluations of signaling pathways, and integration of CNS molecular processes with physiological end points. Moreover, although much has been learned from experimental animal models of hypertension, we must continue to ask whether Ang-II–mediated brain mechanisms act in the same manner in human hypertension.

### Sources of Funding

The authors are supported by National Institutes of Health grants R00HL116776 (C.N. Young), HL063887 (R.L. Davisson), and HL084207 (R.L. Davisson).

### Disclosures

None.

### References

- Chobanian AV. Shattuck Lecture. The hypertension paradox—more uncontrolled disease despite improved therapy. *N Engl J Med*. 2009;361:878–887. doi: 10.1056/NEJMsa0903829.
- Kearney PM, Whelton M, Reynolds K, Muntner P, Whelton PK, He J. Global burden of hypertension: analysis of worldwide data. *Lancet*. 2005;365:217–223. doi: 10.1016/S0140-6736(05)17741-1.
- Laragh JH. Biochemical profiling and the natural history of hypertensive diseases: low-renin essential hypertension, a benign condition. *Circulation*. 1971;44:971–974.
- Zubcevic J, Santisteban MM, Pitts T, Baekey DM, Perez PD, Bolser DC, Febo M, Raizada MK. Functional neural-bone marrow pathways: implications in hypertension and cardiovascular disease. *Hypertension*. 2014;63:e129–e139. doi: 10.1161/HYPERTENSIONAHA.114.02440.
- Zubcevic J, Jun JY, Kim S, Perez PD, Afzal A, Shan Z, Li W, Santisteban MM, Yuan W, Febo M, Mocco J, Feng Y, Scott E, Baekey DM, Raizada MK. Altered inflammatory response is associated with an impaired autonomic input to the bone marrow in the spontaneously hypertensive rat. *Hypertension*. 2014;63:542–550. doi: 10.1161/HYPERTENSIONAHA.113.02722.
- Jun JY, Zubcevic J, Qi Y, Afzal A, Carvajal JM, Thinschmidt JS, Grant MB, Mocco J, Raizada MK. Brain-mediated dysregulation of the bone marrow activity in angiotensin II-induced hypertension. *Hypertension*. 2012;60:1316–1323. doi: 10.1161/HYPERTENSIONAHA.112.199547.
- Ferguson AV. Angiotensinergic regulation of autonomic and neuroendocrine outputs: critical roles for the subfornical organ and paraventricular nucleus. *Neuroendocrinology*. 2009;89:370–376. doi: 10.1159/000211202.
- Faraco G, Iadecola C. Hypertension: a harbinger of stroke and dementia. *Hypertension*. 2013;62:810–817. doi: 10.1161/HYPERTENSIONAHA.113.01063.
- Biancardi VC, Son SJ, Ahmadi S, Filosa JA, Stern JE. Circulating angiotensin II gains access to the hypothalamus and brain stem during hypertension via breakdown of the blood-brain barrier. *Hypertension*. 2014;63:572–579. doi: 10.1161/HYPERTENSIONAHA.113.01743.
- Grobe JL, Xu D, Sigmund CD. An intracellular renin-angiotensin system in neurons: fact, hypothesis, or fantasy. *Physiology (Bethesda)*. 2008;23:187–193. doi: 10.1152/physiol.00002.2008.
- Li W, Peng H, Mehaffey EP, Kimball CD, Grobe JL, van Gool JM, Sullivan MN, Earley S, Danser AH, Ichihara A, Feng Y. Neuron-specific (pro)renin receptor knockout prevents the development of salt-sensitive hypertension. *Hypertension*. 2014;63:316–323. doi: 10.1161/HYPERTENSIONAHA.113.02041.
- Nguyen G, Delarue F, Burcklé C, Bouzahir L, Giller T, Sraer JD. Pivotal role of the renin/prorenin receptor in angiotensin II production and cellular responses to renin. *J Clin Invest*. 2002;109:1417–1427. doi: 10.1172/JCI14276.
- Shan Z, Shi P, Cuadra AE, Dong Y, Lamont GJ, Li Q, Seth DM, Navar LG, Katovich MJ, Summers C, Raizada MK. Involvement of the brain (pro)renin receptor in cardiovascular homeostasis. *Circ Res*. 2010;107:934–938. doi: 10.1161/CIRCRESAHA.110.226977.
- Contrepas A, Walker J, Koulakoff A, Franek KJ, Qadri F, Giaume C, Corvol P, Schwartz CE, Nguyen G. A role of the (pro)renin receptor in neuronal cell differentiation. *Am J Physiol Regul Integr Comp Physiol*. 2009;297:R250–R257. doi: 10.1152/ajpregu.90832.2008.
- Li W, Peng H, Cao T, Sato R, McDaniels SJ, Kobori H, Navar LG, Feng Y. Brain-targeted (pro)renin receptor knockdown attenuates angiotensin II-dependent hypertension. *Hypertension*. 2012;59:1188–1194. doi: 10.1161/HYPERTENSIONAHA.111.190108.
- Li W, Sullivan MN, Zhang S, Worker CJ, Xiong Z, Speth RC, Feng Y. Intracerebroventricular infusion of the (Pro)renin receptor antagonist PRO20 attenuates deoxycorticosterone acetate-salt-induced hypertension. *Hypertension*. 2015;65:352–361. doi: 10.1161/HYPERTENSIONAHA.114.04458.
- Jancovski N, Bassi JK, Carter DA, Choong YT, Connelly A, Nguyen TP, Chen D, Lukoshkova EV, Menuet C, Head GA, Allen AM. Stimulation of angiotensin type 1A receptors on catecholaminergic cells contributes to angiotensin-dependent hypertension. *Hypertension*. 2013;62:866–871. doi: 10.1161/HYPERTENSIONAHA.113.01474.
- Hilzendeger AM, Cassell MD, Davis DR, Stauss HM, Mark AL, Grobe JL, Sigmund CD. Angiotensin type 1a receptors in the subfornical organ are required for deoxycorticosterone acetate-salt hypertension. *Hypertension*. 2013;61:716–722. doi: 10.1161/HYPERTENSIONAHA.111.00356.
- Shan Z, Zubcevic J, Shi P, Jun JY, Dong Y, Murça TM, Lamont GJ, Cuadra A, Yuan W, Qi Y, Li Q, Paton JF, Katovich MJ, Summers C, Raizada MK.

- Chronic knockdown of the nucleus of the solitary tract AT1 receptors increases blood inflammatory-endothelial progenitor cell ratio and exacerbates hypertension in the spontaneously hypertensive rat. *Hypertension*. 2013;61:1328–1333. doi: 10.1161/HYPERTENSIONAHA.111.00156.
20. Blanch GT, Freiria-Oliveira AH, Speretta GF, Carrera EJ, Li H, Speth RC, Colombari E, Summers C, Colombari DS. Increased expression of angiotensin II type 2 receptors in the solitary-vagal complex blunts renovascular hypertension. *Hypertension*. 2014;64:777–783. doi: 10.1161/HYPERTENSIONAHA.114.03188.
  21. Xu P, Sriramula S, Lazartigues E. ACE2/ANG-(1-7)/Mas pathway in the brain: the axis of good. *Am J Physiol Regul Integr Comp Physiol*. 2011;300:R804–R817. doi: 10.1152/ajpregu.00222.2010.
  22. Peterson JR, Sharma RV, Davisson RL. Reactive oxygen species in the neuropathogenesis of hypertension. *Curr Hypertens Rep*. 2006;8:232–241.
  23. Zimmerman MC, Lazartigues E, Lang JA, Sinnayah P, Ahmad IM, Spitz DR, Davisson RL. Superoxide mediates the actions of angiotensin II in the central nervous system. *Circ Res*. 2002;91:1038–1045.
  24. Zimmerman MC, Lazartigues E, Sharma RV, Davisson RL. Hypertension caused by angiotensin II infusion involves increased superoxide production in the central nervous system. *Circ Res*. 2004;95:210–216. doi: 10.1161/01.RES.0000135483.12297.e4.
  25. Zimmerman MC, Dunlay RP, Lazartigues E, Zhang Y, Sharma RV, Engelhardt JF, Davisson RL. Requirement for Rac1-dependent NADPH oxidase in the cardiovascular and dipsogenic actions of angiotensin II in the brain. *Circ Res*. 2004;95:532–539. doi: 10.1161/01.RES.0000139957.22530.b9.
  26. Lob HE, Schultz D, Marvar PJ, Davisson RL, Harrison DG. Role of the NADPH oxidases in the subfornical organ in angiotensin II-induced hypertension. *Hypertension*. 2013;61:382–387. doi: 10.1161/HYPERTENSIONAHA.111.00546.
  27. Peterson JR, Burmeister MA, Tian X, Zhou Y, Guraju MR, Stupinski JA, Sharma RV, Davisson RL. Genetic silencing of Nox2 and Nox4 reveals differential roles of these NADPH oxidase homologues in the vaso-pressor and dipsogenic effects of brain angiotensin II. *Hypertension*. 2009;54:1106–1114. doi: 10.1161/HYPERTENSIONAHA.109.140087.
  28. Peng H, Li W, Seth DM, Nair AR, Francis J, Feng Y. (Pro)renin receptor mediates both angiotensin II-dependent and -independent oxidative stress in neuronal cells. *PLoS One*. 2013;8:e58339. doi: 10.1371/journal.pone.0058339.
  29. Cao X, Peterson JR, Wang G, Anrather J, Young CN, Guraju MR, Burmeister MA, Iadecola C, Davisson RL. Angiotensin II-dependent hypertension requires cyclooxygenase 1-derived prostaglandin E2 and EP1 receptor signaling in the subfornical organ of the brain. *Hypertension*. 2012;59:869–876. doi: 10.1161/HYPERTENSIONAHA.111.182071.
  30. Asirvatham-Jeyaraj N, King AJ, Northcott CA, Madan S, Fink GD. Cyclooxygenase-1 inhibition attenuates angiotensin II-salt hypertension and neurogenic pressor activity in the rat. *Am J Physiol Heart Circ Physiol*. 2013;305:H1462–H1470. doi: 10.1152/ajpheart.00245.2013.
  31. Wang G, Sarkar P, Peterson JR, Anrather J, Pierce JP, Moore JM, Feng J, Zhou P, Milner TA, Pickel VM, Iadecola C, Davisson RL. COX-1-derived PGE2 and PGE2 type 1 receptors are vital for angiotensin II-induced formation of reactive oxygen species and Ca(2+) influx in the subfornical organ. *Am J Physiol Heart Circ Physiol*. 2013;305:H1451–H1461. doi: 10.1152/ajpheart.00238.2013.
  32. Schröder M, Kaufman RJ. The mammalian unfolded protein response. *Annu Rev Biochem*. 2005;74:739–789. doi: 10.1146/annurev.biochem.73.011303.074134.
  33. Young CN, Cao X, Guraju MR, Pierce JP, Morgan DA, Wang G, Iadecola C, Mark AL, Davisson RL. ER stress in the brain subfornical organ mediates angiotensin-dependent hypertension. *J Clin Invest*. 2012;122:3960–3964. doi: 10.1172/JCI64583.
  34. Chao YM, Lai MD, Chan JY. Redox-sensitive endoplasmic reticulum stress and autophagy at rostral ventrolateral medulla contribute to hypertension in spontaneously hypertensive rats. *Hypertension*. 2013;61:1270–1280. doi: 10.1161/HYPERTENSIONAHA.111.00469.
  35. Spitzer KM, Webb RC. Endoplasmic reticulum stress contributes to aortic stiffening via proapoptotic and fibrotic signaling mechanisms. *Hypertension*. 2014;63:e40–e45. doi: 10.1161/HYPERTENSIONAHA.113.02558.
  36. Kassam M, Galán M, Partyka M, Saifudeen Z, Henrion D, Trebak M, Matrougui K. Endoplasmic reticulum stress is involved in cardiac damage and vascular endothelial dysfunction in hypertensive mice. *Arterioscler Thromb Vasc Biol*. 2012;32:1652–1661. doi: 10.1161/ATVBAHA.112.249318.
  37. Fisher JP, Young CN, Fadel PJ. Central sympathetic overactivity: maladies and mechanisms. *Auton Neurosci*. 2009;148:5–15. doi: 10.1016/j.autneu.2009.02.003.
  38. Hasty AH, Harrison DG. Endoplasmic reticulum stress and hypertension - a new paradigm? *J Clin Invest*. 2012;122:3859–3861. doi: 10.1172/JCI65173.
  39. Jo F, Jo H, Hilzendeger AM, Thompson AP, Cassell MD, Rutkowski DT, Davissan RL, Grobe JL, Sigmund CD. Brain endoplasmic reticulum stress mechanistically distinguishes the saline-intake and hypertensive response to deoxycorticosterone acetate-salt. *Hypertension*. 2015;65:1341–1348. doi: 10.1161/HYPERTENSIONAHA.115.05377.
  40. Zubcevic J, Waki H, Raizada MK, Paton JF. Autonomic-immune-vascular interaction: an emerging concept for neurogenic hypertension. *Hypertension*. 2011;57:1026–1033. doi: 10.1161/HYPERTENSIONAHA.111.169748.
  41. Harrison DG, Guzik TJ, Lob HE, Madhur MS, Marvar PJ, Thabet SR, Vinh A, Weyand CM. Inflammation, immunity, and hypertension. *Hypertension*. 2011;57:132–140. doi: 10.1161/HYPERTENSIONAHA.110.163576.
  42. Marvar PJ, Thabet SR, Guzik TJ, Lob HE, McCann LA, Weyand C, Gordon FJ, Harrison DG. Central and peripheral mechanisms of T-lymphocyte activation and vascular inflammation produced by angiotensin II-induced hypertension. *Circ Res*. 2010;107:263–270. doi: 10.1161/CIRCRESAHA.110.217299.
  43. Lob HE, Marvar PJ, Guzik TJ, Sharma S, McCann LA, Weyand C, Gordon FJ, Harrison DG. Induction of hypertension and peripheral inflammation by reduction of extracellular superoxide dismutase in the central nervous system. *Hypertension*. 2010;55:277–283, 6p following 283. doi: 10.1161/HYPERTENSIONAHA.109.142646.
  44. Abboud FM, Harwani SC, Chapleau MW. Autonomic neural regulation of the immune system: implications for hypertension and cardiovascular disease. *Hypertension*. 2012;59:755–762. doi: 10.1161/HYPERTENSIONAHA.111.186833.
  45. Harwani SC, Chapleau MW, Legge KL, Ballas ZK, Abboud FM. Neurohormonal modulation of the innate immune system is proinflammatory in the prehypertensive spontaneously hypertensive rat, a genetic model of essential hypertension. *Circ Res*. 2012;111:1190–1197. doi: 10.1161/CIRCRESAHA.112.277475.
  46. Li DJ, Evans RG, Yang ZW, Song SW, Wang P, Ma XJ, Liu C, Xi T, Su DF, Shen FM. Dysfunction of the cholinergic anti-inflammatory pathway mediates organ damage in hypertension. *Hypertension*. 2011;57:298–307. doi: 10.1161/HYPERTENSIONAHA.110.160077.
  47. Ganta CK, Lu N, Helwig BG, Blecha F, Ganta RR, Zheng L, Ross CR, Musch TI, Fels RJ, Kenney MJ. Central angiotensin II-enhanced splenic cytokine gene expression is mediated by the sympathetic nervous system. *Am J Physiol Heart Circ Physiol*. 2005;289:H1683–H1691. doi: 10.1152/ajpheart.00125.2005.
  48. Bardgett ME, Holbein WW, Herrera-Rosales M, Toney GM. Ang II-salt hypertension depends on neuronal activity in the hypothalamic paraventricular nucleus but not on local actions of tumor necrosis factor- $\alpha$ . *Hypertension*. 2014;63:527–534. doi: 10.1161/HYPERTENSIONAHA.113.02429.
  49. Shi P, Diez-Freire C, Jun JY, Qi Y, Katovich MJ, Li Q, Sriramula S, Francis J, Summers C, Raizada MK. Brain microglial cytokines in neurogenic hypertension. *Hypertension*. 2010;56:297–303. doi: 10.1161/HYPERTENSIONAHA.110.150409.
  50. Waki H, Gouraud SS, Maeda M, Raizada MK, Paton JF. Contributions of vascular inflammation in the brainstem for neurogenic hypertension. *Respir Physiol Neurobiol*. 2011;178:422–428. doi: 10.1016/j.resp.2011.05.004.
  51. Santisteban MM, Ahmari N, Carvajal JM, Zingler MB, Qi Y, Kim S, Joseph J, Garcia-Pereira F, Johnson RD, Shenoy V, Raizada MK, Zubcevic J. Involvement of bone marrow cells and neuroinflammation in hypertension. *Circ Res*. 2015;117:178–191. doi: 10.1161/CIRCRESAHA.117.305853.
  52. Pollow DP, Uhrlaub J, Romero-Aleshire MJ, Sandberg K, Nikolich-Zugich J, Brooks HL, Hay M. Sex differences in T-lymphocyte tissue infiltration and development of angiotensin II hypertension. *Hypertension*. 2014;64:384–390. doi: 10.1161/HYPERTENSIONAHA.114.03581.
  53. Wei SG, Yu Y, Zhang ZH, Felder RB. Proinflammatory cytokines upregulate sympathoexcitatory mechanisms in the subfornical organ of the rat. *Hypertension*. 2015;65:1126–1133. doi: 10.1161/HYPERTENSIONAHA.114.05112.
  54. Summers C, Fleegal MA, Zhu M. Angiotensin AT1 receptor signalling pathways in neurons. *Clin Exp Pharmacol Physiol*. 2002;29:483–490.
  55. Cardinale JP, Sriramula S, Mariappan N, Agarwal D, Francis J. Angiotensin II-induced hypertension is modulated by nuclear factor- $\kappa$ B in the paraventricular nucleus. *Hypertension*. 2012;59:113–121. doi: 10.1161/HYPERTENSIONAHA.111.182154.

56. Kang YM, Ma Y, Zheng JP, Elks C, Sriramula S, Yang ZM, Francis J. Brain nuclear factor-kappa B activation contributes to neurohumoral excitation in angiotensin II-induced hypertension. *Cardiovasc Res*. 2009;82:503–512. doi: 10.1093/cvr/cvp073.
57. Young CN, Li A, Dong FN, Horwath JA, Clark CG, Davisson RL. Endoplasmic reticulum and oxidant stress mediate activation of nuclear factor- $\kappa$ -B in the subfornical organ during angiotensin-II hypertension. *AJP-Cell Physiology*. 2015;308:C803–C812.
58. Burmeister MA, Young CN, Braga VA, Butler SD, Sharma RV, Davisson RL. In vivo bioluminescence imaging reveals redox-regulated activator protein-1 activation in paraventricular nucleus of mice with renovascular hypertension. *Hypertension*. 2011;57:289–297. doi: 10.1161/HYPERTENSIONAHA.110.160564.
59. Peterson JR, Infanger DW, Braga VA, Zhang Y, Sharma RV, Engelhardt JF, Davisson RL. Longitudinal noninvasive monitoring of transcription factor activation in cardiovascular regulatory nuclei using bioluminescence imaging. *Physiol Genomics*. 2008;33:292–299. doi: 10.1152/physiolgenomics.00296.2007.
60. Lazartigues E, Lawrence AJ, Lamb FS, Davisson RL. Renovascular hypertension in mice with brain-selective overexpression of AT1a receptors is buffered by increased nitric oxide production in the periphery. *Circ Res*. 2004;95:523–531. doi: 10.1161/01.RES.0000140892.86313.c2.

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*Hypertension*. 2015;66:920-926; originally published online August 31, 2015;

doi: 10.1161/HYPERTENSIONAHA.115.03624

*Hypertension* is published by the American Heart Association, 7272 Greenville Avenue, Dallas, TX 75231

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Print ISSN: 0194-911X. Online ISSN: 1524-4563

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