Spironolactone Prevents Endothelial Nitric Oxide Synthase Uncoupling and Vascular Dysfunction Induced by β-Adrenergic Overstimulation
Role of Perivascular Adipose Tissue

Jamaira A. Victorio, Stefano P. Clerici, Roberto Palacios, María J. Alonso, Dalton V. Vassallo, Iris Z. Jaffe, Luciana V. Rossoni, Ana P. Davel

Abstract—Sustained stimulation of β-adrenoceptors (β-ARs) and activation of renin–angiotensin–aldosterone system are common features of cardiovascular diseases with rising sympathetic activation, including essential hypertension, myocardial infarction, and heart failure. In this study, we investigated the role of AT1 receptor and mineralocorticoid receptor (MR) in the vascular alterations caused by β-AR overstimulation. β-AR overstimulation with associated cardiac hypertrophy and increased vasoconstrictor response to phenylephrine in aorta were modeled in rats by 7-day isoproterenol treatment. The increased vasoconstrictor response to phenylephrine in this model was blunted by the MR antagonist spironolactone, but not by the AT1 receptor antagonist losartan, despite the blunting of cardiac hypertrophy with both drugs. Spironolactone, but not losartan, restored NO bioavailability in association with lower endothelial nitric oxide synthase–derived superoxide production, increased endothelial nitric oxide synthase dimerization, and aortic HSP90 upregulation. MR genomic and nongenomic functions were activated in aortas from isoproterenol-treated rats. Isoproterenol did not modify plasma levels of MR ligands aldosterone and corticosterone but rather increased perivascular adipose tissue–derived corticosterone in association with increased expression of 11β-hydroxysteroid dehydrogenase type 1. The anticontractive effect of aortic perivascular adipose tissue was impaired by β-AR overstimulation and restored by MR blockade. These results suggest that activation of vascular MR signaling contributes to the vascular dysfunction induced by β-AR overstimulation associated with endothelial nitric oxide synthase uncoupling. These findings reveal an additional explanation for the protective effects of MR antagonists in cardiovascular disorders with sympathetic activation. (Hypertension. 2016;68:726-735. DOI: 10.1161/HYPERTENSIONAHA.116.07911.) • Online Data Supplement

Key Words: adipose tissue ■ aorta ■ beta-adrenergic receptors ■ endothelial nitric oxide synthase ■ losartan ■ mineralocorticoid receptor

Sympathetic overactivity with rising catecholamines levels and adrenergic receptors stimulation is a common feature of many cardiovascular disorders, including hypertension, myocardial infarction (MI), congestive heart failure, and acute cerebrovascular events. In these conditions, the hyperadrenergic state has a major and independent prognostic impact. Although the importance of β-adrenoceptor (β-AR) overstimulation in the pathogenesis of left ventricular dysfunction has been widely studied, less is known about its effects on vascular function. Following in vivo β-AR overstimulation, we and others have demonstrated abnormal vasoconstrictor response to agonists in aorta, coronary artery, and cerebral artery. In thoracic aorta, increased vasoconstrictor response induced by isoproterenol treatment was associated with increased reactive oxygen species generation and uncoupling of endothelial nitric oxide synthase (eNOS). These studies demonstrate that β-AR overstimulation induces vascular dysfunction but the molecular mechanisms remain to be elucidated.

Activation of the renin–angiotensin–aldosterone system is also involved in the pathogenesis of cardiovascular and metabolic diseases, including hypertension, MI, heart failure, and obesity. It is known that β-AR signaling in juxtaglomerular cells stimulates renin release, thereby stimulating renin–angiotensin–aldosterone system. In addition, β-AR agonist...
isoproterenol increases cardiac expression of angiotensin-converting enzyme and antagonism or deficiency of AT1 receptor (AT1R) attenuate isoproterenol-induced cardiac remodeling in mice. Elevated circulating aldosterone is also associated with isoproterenol-induced heart failure, and blockade of aldosterone-binding mineralocorticoid receptor (MR) is protective from cardiac hypertrophy and diastolic dysfunction induced by chronic isoproterenol treatment in rats. Together, these studies have suggested beneficial cardiac effects of AT1R and MR blockade in preventing isoproterenol-induced cardiac remodeling and dysfunction. In the vasculature, either AT1R or MR activation induces proinflammatory, profibrotic, cardiac remodeling and dysfunction. In the vasculature, either AT1R or MR activation induces proinflammatory, profibrotic, and pro-oxidative vascular signaling pathways. However, whether AT1R and MR contribute to the vascular abnormalities caused by β-AR overstimulation has not been explored.

Vascular function is also known to be modified by perivascular adipose tissue (PVAT). Angiotensin II, via AT1R promotes aldosterone secretion from adipocytes, acting in a paracrine manner to regulate vascular function and contributing to endothelial dysfunction in obesity. PVAT of the thoracic aorta also releases adipocyte-derived relaxing factors that exhibit anticontractive effects. Acute β-AR activation stimulates the release of adipocyte-derived relaxing factors from PVAT of mesenteric artery. However, it is not known whether β-AR overstimulation could regulate the anticontractile effects of PVAT.

Therefore, in this study, we investigated a possible role of AT1R, MR, and PVAT in the vascular dysfunction induced by in vivo administration of isoproterenol as measured by enhanced vasoconstriction to phenylephrine. We hypothesized that MR activation induces uncoupling of eNOS, oxidative stress, and reduces anticontractile role of PVAT after β-AR overstimulation.

Methods

Animals

This study was approved by the Ethics Committee on Animal Use of the University of Campinas (protocol no. 2609-1) and carried out in accordance with the ethical principles for animal experimentation adopted by the Brazilian Society of Laboratory Animal Science (SBCAL/COBEA).

Male Wistar rats (12-week old) were obtained from the Multidisciplinary Center for Biological Research of the University of Campinas (Campinas, Brazil). Animals were housed at a constant room temperature (22°C), 12:12 hour light:dark cycle, and with normal chow and water provided ad libitum. Isoproterenol (0.3 mg/kg per day, sc) or the vehicle were administrated once daily for 7 days, concomitantly with the treatment or not via oral gavage with the AT1R antagonist losartan (40 mg/kg per day), or with the MR antagonist spironolactone (200 mg/kg per day).

Vascular Reactivity, Blood Pressure, and Biochemical and Molecular parameters

Detailed methods are available in the only-online Data Supplement.

Statistical Analysis

Data are presented as mean±SEM. Data were analyzed by the Student t test or two-way ANOVA followed by the Bonferroni post-test by using GraphPad Prism 5.0 software (GraphPad Software Corp, San Diego, CA). Values of P<0.05 were considered significantly different.

Results

Losartan and Spironolactone Treatments Similarly Reduce Isoproterenol-Induced Cardiac Hypertrophy

To confirm the efficacy of isoproterenol treatment in inducing long-term β-AR stimulation, the ventricular weight:body weight ratio was measured as an index of myocardial hypertrophy. Isoproterenol treatment increased ventricular weight:body weight ratio without affecting body weight that was similarly reduced by losartan and spironolactone (Table S1). No effect of either isoproterenol or spironolactone on blood pressure or heart rate was observed, but losartan decreased diastolic blood pressure in both control and isoproterenol-treated rats (Table S1).

Spironolactone, but Not Losartan, Prevented the Increased Aortic Reactivity to Phenylephrine in Isoproterenol-Treated Rats

Aortic rings from isoproterenol-treated rats showed an increased contractile response to phenylephrine compared with the control group (Figure 1A). This high contractility was not altered by cotreatment with losartan (Figure 1B), whereas it was fully prevented by spironolactone cotreatment (Figure 1C). These data support a role for MR, but not the AT1R, in the increased aortic contractile response induced by β-AR overstimulation. The relaxation to either acetylcholine

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**Figure 1.** Spironolactone, but not losartan, prevented the enhanced contraction to phenylephrine induced by β-adrenergic overstimulation. Concentration–response curves to phenylephrine obtained in aortic rings from rats treated with vehicle (CT) or isoproterenol (ISO; A) combined with losartan (LOS; B) or with spironolactone (SPI; C). Data are expressed as mean±SEM; number of animals is indicated in parenthesis. Two-way ANOVA: *P<0.001 vs CT or LOS.
or sodium nitroprusside was not modified by the treatments (Figure S1).

MR Antagonist Restored NO Bioavailability, NOS Dimerization, and HSP90 Protein Levels in Aortas of Isoproterenol-Treated Rats

Incubation with a nonspecific NOS inhibitor, No-nitro-L-arginine methyl ester, enhanced the contractile response to phenylephrine in the aorta of all the groups (Figure 2A). The NOS-dependent anticontractile component of phenylephrine response (as measured by difference of the area under the curve before and after No-nitro-L-arginine methyl ester) was impaired in the isoproterenol group, whereas spironolactone reversed this effect (Figure 2B). Pretreatment of aortas with superoxide dismutase (SOD, superoxide scavenger) or tetrahydrobiopterin (BH4, eNOS cofactor) reduced vascular contraction in the isoproterenol group but not in the control and spironolactone-treated groups (Figure 2C and 2E). There was an increased difference of the area under the curve to phenylephrine in the presence of SOD and BH4 in isoproterenol-treated rats, which was normalized by spironolactone (Figure 2D and 2F). These data support a role for MR in increasing superoxide and reducing NO production and bioavailability after β-AR overstimulation. Indeed, aortic NO levels (evaluated by the fluorescence to diaminofluorescein) were decreased in rats exposed to long-term isoproterenol and spironolactone treatments enhanced NO to levels similar to the control group (Figure 3A). By contrast, losartan treatment did not prevent the impairment in NO bioavailability induced by isoproterenol (Figure S2).

Although aortic expression of total eNOS protein was increased in isoproterenol-treated rats (Figure 3B), the abundance of its dimeric form was reduced (Figure 3C), as well as its phosphorylation in Ser1177 (Figure S3). Spironolactone did not affect the isoproterenol-induced increase in total eNOS protein levels, but it normalized eNOS dimerization (Figure 3B and 3C). eNOS phosphorylation were not affected by spironolactone or losartan treatment (Figure S3). Because HSP90 is an eNOS chaperone that augments NO production and inhibits superoxide formation,16 we investigated HSP90 expression. Isoproterenol treatment significantly reduced HSP90 protein expression, which was restored by spironolactone (Figure 3D).

Figure 2. Effect of No-nitro-L-arginine methyl ester (L-NAME) (A, square symbols), superoxide dismutase (superoxide dismutase (SOD, superoxide scavenger); C, triangle symbols), and tetrahydrobiopterin (BH4, eNOS cofactor) (E, diamond symbol) on the concentration–response curves to phenylephrine of aortic rings from control (CT) and isoproterenol (ISO) groups without or with spironolactone (SPI) treatment. B, D, and F. The difference of the area under the curve (dAUC) to phenylephrine in the presence and absence of L-NAME, SOD, or BH4, respectively. Data are expressed as mean±SEM (n=4–15 in each group). Two-way ANOVA: P<0.05 +vs respective group without incubation; *vs CT; #vs ISO.
eNOS-Derived Superoxide Anion Production After β-AR Overstimulation Is Prevented by Spironolactone

We measured reactive oxygen species production by quantification of the fluorescence formed after exposing aortic slices to hidroethidine. The fluorescent signal was almost undetectable after incubation with the SOD mimetic MnTMPyP, indicating superoxide as the main reactive oxygen species detectable by hidroethidine in aortas (Figure 3E). Isoproterenol induced an increase in vascular superoxide production that was inhibited by Nω-nitro-L-arginine methyl ester incubation, indicating enhanced eNOS-derived superoxide production (Figure 3E). Importantly, spironolactone, but not losartan, blocked this increase (Figure 3E; Figure S4). The data indicate that β-AR overstimulation induces vascular oxidative stress by eNOS, dependent on MR activation.

β-Adrenergic Overstimulation Activates Aortic MR Genomic and Nongenomic Activity

MR functions by translocating to the nucleus to regulate gene transcription (genomic mechanisms) and also by activating cytoplasmic signaling pathways (nongenomic mechanisms).
Isoproterenol treatment increased the nuclear:cytoplasmic MR ratio in the aorta (Figure 4A) and increased gene expression of the smooth muscle cell MR target gene, osteopontin (Figure 4B) and the endothelial MR target protein, the γ-subunit of the epithelial sodium channel (γENaC; C), Src (D), and ERK1/2 (E) phosphorylation in thoracic aorta of control (CT) and isoproterenol (ISO) groups without or with spironolactone (SPI) treatment. Data represent mean±SEM; number of animals used in each group is indicated into the bars. Two-way ANOVA: $P<0.05$ *vs CT; #vs ISO.

Enhanced Corticosterone Levels and Increased Expression of 11β-HSD1 in PVAT After Isoproterenol Treatment

MR can be activated by the mineralocorticoid aldosterone and also by some corticosteroids. To investigate if isoproterenol activates the MR by modulating levels of endogenous ligands,
we measured plasma and PVAT levels of aldosterone and corticosterone (the main glucocorticoid in rodents). Plasma and PVAT levels of aldosterone were increased by spironolactone treatment, consistent with a feedback mechanism associated with effective MR blockade (Figure 5A and 5B). Neither isoproterenol nor spironolactone treatment affected plasma corticosterone levels (Figure 5C). However, corticosterone content was enhanced in PVAT after isoproterenol treatment, whereas spironolactone did not alter this effect (Figure 5D). Treatments did not affect aortic PVAT weight (data not shown). These data suggest that enhanced PVAT-derived corticosterone could be a mechanism leading to paracrine activation of vascular MR after β-adrenergic overstimulation.

PVAT protein expression of aldosterone synthase (CYP11B1), a final enzyme required for glucocorticoid synthesis, was not affected by isoproterenol treatment (Figure S5A); whereas, isoproterenol increased the PVAT expression of 11β-hydroxysteroid dehydrogenase type-1 (11β-HSD1), the enzyme that generates active glucocorticoids from their inactive 11-keto derivatives (Figure 5E), with no effect in 11β-hydroxysteroid dehydrogenase type-2 (11β-HSD2) expression (Figure 5F). Spironolactone decreased the expression of 11β-HSD1 and 11β-HSD2 (Figure 5E and 5F). Glucocorticoid synthase (CYP11B2) protein expression in PVAT did not differ from the control and isoproterenol groups (Figure S5B).

**Spironolactone Restores the Anticontractile Effect of Perivascular Tissue, Which Is Impaired by Isoproterenol**

Next, we investigated the role of PVAT on vascular contraction to phenylephrine. As expected, the presence of PVAT reduced the contraction to phenylephrine in aortas from control rats (Figure 6A). However, this anticontractile effect of PVAT was impaired by isoproterenol treatment (Figure 6B and 6E).

**Figure 5. β-AR overstimulation enhanced corticosterone content and protein expression of 11β-HSD1 in perivascular adipose tissue (PVAT) of aorta.** Aldosterone (A and B) and corticosterone (C and D) content measured in plasma and aortic PVAT from control (CT) and isoproterenol (ISO) groups without or with spironolactone (SPI) treatment. Protein expression of 11β-HSD1 (E) and 11β-HSD2 (F) were evaluated in PVAT from CT, ISO, SPI, and ISO+SPI groups. Data represent mean±SEM; number of animals is indicated in the bars. Two-way ANOVA: *P<0.05 vs CT; +vs SPI; #vs ISO.
Spironolactone did not affect the basal role of PVAT on the contractile response to phenylephrine (Figure 6C and 6E) but rather, in the presence of MR antagonist, the anticontractile effect of PVAT was restored in rats cotreated with isoproterenol and spironolactone (Figure 6D and 6E).

Recently, it was demonstrated that PVAT from thoracic aorta expresses eNOS that produces NO as a PVAT-derived relaxing factor, whereas uncoupled eNOS in this tissue might be a mechanism involved in vascular dysfunction. Therefore, we explored eNOS uncoupling in PVAT after β-adrenergic overstimulation. PVAT-derived NO production was impaired by isoproterenol treatment and restored by spironolactone treatment or BH4 incubation (Figure 6F).

**Discussion**

This study demonstrated that the MR antagonist spironolactone, but not the AT1R blocker losartan, prevented the increased vasoconstrictor response to phenylephrine induced by β-AR overstimulation. The protective vascular effect of spironolactone was associated with (1) increased eNOS dimerization, HSP90 expression and NO production, (2) reduced eNOS-derived superoxide production, (3) inhibition of genomic and nongenomic vascular MR pathways, and (4) restoration of the anticontractile role of aortic PVAT. Furthermore, we found elevated corticosterone content in aortic PVAT after β-adrenergic stimulation. These findings support a model in which chronic β-adrenergic stimulation promotes vascular MR activation, which results in eNOS uncoupling and oxidative stress. This model provides a novel mechanism by which MR antagonists can be protective in patients with cardiovascular disease by preventing vascular dysfunction associated with hyperadrenergic state, such as in heart failure, MI, and essential hypertension.

Renin–angiotensin–aldosterone system inhibitors, including angiotensin-converting enzyme inhibitors and AT1R antagonists, and more recently MR antagonists improve survival in patients with left ventricular dysfunction. The diuretic effect of the MR antagonist was not sufficient
to explain its cardioprotective effects in patients with heart failure. Although both losartan and spironolactone reduced ventricular hypertrophy in a similar magnitude, we observed that only MR antagonism was effective in the prevention of increased vasoconstrictor response to phenylephrine, NO bioavailability, and eNOS-dependent oxidative stress. This result demonstrates for the first time a role for MR, but not AT1R, in mediating the major vascular effects of β-adrenergic overstimulation. In addition, losartan, but not spironolactone, reduced diastolic blood pressure measured invasively in anesthetized rats. However, we cannot exclude the possibility of group differences in blood pressure because we did not monitor this parameter 24 hours per day in awake, unrestrained animals.

Abnormal vasoconstrictor responses to agonists including phenylephrine have been demonstrated in several vascular beds in the isoproterenol-induced left ventricular hypertrophy model. These studies put forward a key role for β-AR overstimulation in inducing vascular dysfunction. We previously demonstrated a role for uncoupled eNOS in the altered vascular contractility induced by β-AR stimulation. Because aldosterone-induced MR activation can impair eNOS-derived NO production associated with a reduction in eNOS dimerization in endothelial cells, we hypothesized that MR blockade could improve this endothelial pathway. In accordance, improved NO bioavailability, enhanced eNOS dimer expression, and reduced eNOS-derived superoxide were observed in aortas from isoproterenol+spironolactone–treated rats. Although eNOS uncoupling was associated with exacerbated contractile response to phenylephrine, the endothelium-dependent relaxation to acetylcholine was not significantly changed by isoproterenol treatment. This apparently contradictory finding might be explained by the greater sensitivity of basal NO to destruction by superoxide when compared with agonist-stimulated NO production.

Dimerization of eNOS regulates its catalytic activity and NO production. Association of eNOS with HSP90 has been demonstrated to be an important mechanism regulating eNOS dimerization, rather than eNOS phosphorylation. Here, we show that β-AR overstimulation significantly reduced aortic expression of this chaperone, which was prevented by spironolactone. Interestingly, impaired interaction of HSP90 with eNOS results in decreased NO production and superoxide generation. Therefore, reduced vascular expression of HSP90 is a potential link between β-AR overstimulation and eNOS dysfunction that can be prevented by MR blockade.

Increased nuclear:cytoplasmic MR ratio was found in aorta from isoproterenol-treated rats. Given that MR dissociation from HSP90 induces MR cytoplasmic-to-nucleus trafficking, reduction in HSP90 expression by isoproterenol could also contribute to enhance MR transcriptional activity. Spironolactone treatment enhanced HSP90 expression and attenuated nuclear localization of MR, osteopontin mRNA, and ENaC protein levels in isoproterenol-treated rats. Osteopontin is a multifunctional glycosphosphoprotein that can be secreted by endothelium and vascular smooth muscle cells; its gene expression is induced by MR and contributes to proinflammatory and profibrotic effects of aldosterone. In addition, recent data suggest that MR activation induces binding to the ENaC promoter, increasing expression of ENaC mRNA in endothelial cells with associated endothelial dysfunction. We observed an increased aortic expression of the regulatory γ-subunit of ENaC in aorta of isoproterenol-treated rats that was prevented by cotreatment with spironolactone, suggesting that β-adrenergic stimulation might upregulate ENaC through MR activation. To our knowledge, this is the first demonstration that osteopontin and ENaC can be upregulated in vascular tissue in response to β-adrenergic stimulation and could be an additional mechanism involved in the vascular pathology associated with sympathetic hyperactivity.

In addition to genomic activity of MR, a nongenomic MR pathway in the vasculature has been demonstrated to contribute to oxidative stress, inflammation, and vascular dysfunction. Aldosterone rapidly activates several kinases, including Src and mitogen-activated protein kinases in vascular smooth muscle cells. Here, we reported that MR blockade attenuated phosphorylation of Src and ERK1/2 in aortas from isoproterenol-treated rats, which indicates convergence of the β-adrenergic and MR-signaling pathways. However, the observed association between the spironolactone-induced changes in the biochemical and molecular parameters evaluated and the vascular protection in the contractile function may not necessarily reflect cause and effect relationship, representing a limitation of this study.

Both aldosterone and glucocorticoids bind to the MR to activate its genomic and nongenomic functions. In this study, β-AR overstimulation did not change either aldosterone or corticosterone plasma levels; however, corticosterone, but not aldosterone, was enhanced in aortic PVAT from isoproterenol-treated rats. Spironolactone did not attenuate the enhanced levels of corticosterone induced by isoproterenol in PVAT, suggesting that the beneficial vascular effects of spironolactone may be downstream and could be because of blockade of glucocorticoid-induced MR activation. Mature adipocytes express CYP11B2 and CYP11B1 and can produce aldosterone and corticosterone. The expression of these enzymes was not modified by isoproterenol treatment. Glucocorticoid content is also regulated by 11β-HSD type 1 and 2. 11β-HSD2 converts glucocorticoids into inactive metabolites, which favors aldosterone-MR interaction. However, 11β-HSD1 in the presence of NADPH promotes glucocorticoids regeneration from inert 11-keto metabolites. Therefore, an upregulation of 11β-HSD1 in PVAT after chronic β-adrenergic stimulation might be a mechanism associated with high perivascular corticosterone content, thereby leading to paracrine activation of vascular MR. However, as β-AR signal through the small GTPase Rac1, which has been shown to activate MR without ligand, we should also consider ligand-independent MR activation as a potential mechanism for isoproterenol-induced MR activation.

Spironolactone treatment prevented the isoproterenol-induced 11β-HSD1 expression in PVAT. Earlier studies demonstrated that suppression of 11β-HSD1 abolishes the inhibitory effect of glucocorticoids on eNOS expression and prevents heart failure development after MI. Therefore, reduction in 11β-HSD1 expression in PVAT is a potential cardiovascular protective mechanism of spironolactone. Despite this effect, corticosterone levels were still high in PVAT from rats cotreated with isoproterenol and spironolactone, which
could be related to the minor expression of 11β-HSD2 induced by spironolactone. However, the cellular origin of corticosterone secretion induced by isoproterenol in PVAT and the signaling pathway involved in this effect remains an open question. In addition, as endothelium, vascular smooth muscle cells, inflammatory cells and adipocytes express functional MR further investigation using cell-type-specific knock-out mice should address which cell-type-specific MR signaling is mediating the vascular effects induced by β-adrenergic stimulation.

In conclusion, our findings revealed a novel mechanism of regulation of vascular dysfunction mediated by β-AR overstimulation inducing PVAT-derived corticosterone production, associated with impaired PVAT anticontractile effect and vascular MR activation. MR blockade with spironolactone protected from increased vasoconstrictor response to phenylephrine, upregulation of ENaC, and downregulation of HSP90 and eNOS uncoupling. The present findings uncover a role for MR blockade in sympathoexcitatory cardiovascular diseases and provide an additional novel vascular mechanism for the protective effects of MR antagonism.

**Perspectives**

We demonstrated that MR antagonist spironolactone prevents the vascular alterations induced by long-term β-AR stimulation, including enhanced vasoconstriction, uncoupling of eNOS, reduced NO bioavailability, and oxidative stress. The beneficial vascular effects of spironolactone were independent of changes in systemic levels of MR ligand. Instead, we observed increased levels of PVAT-derived corticosterone in response to β-adrenergic stimulation associated with enhanced protein levels of 11β-HSD1 that regenerates glucocorticoids. MR genomic and nongenomic signaling were observed in arterioles of isoproterenol-treated rats. Therefore, this study suggests a novel link between β-AR signaling and vascular MR activation in causing vascular dysfunction. Moreover, the results indicate an additional mechanism for the protective vascular effects of MR antagonists in cardiovascular diseases associated with increased sympathetic activity, such as essential hypertension, MI, and heart failure.

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**Disclosures**

None.

**References**


**Novelty and Significance**

**What Is New?**

- Spironolactone prevents the increased vasoconstrictor response to phenylephrine and uncoupling of endothelial nitric oxide synthase in aorta of a rat model of chronic β-adrenergic stimulation induced by isoproterenol.
- β-AR overstimulation impairs the anticontractile function of perivascular adipose tissue and induces perivascular adipose tissue–derived glucocorticoid production.

**What Is Relevant?**

- Mineralocorticoid receptor blockade with spironolactone prevented the vascular dysfunction induced by β-AR overstimulation, independent of changes in systemic levels of mineralocorticoid receptor ligands, suggesting a novel mechanism for the protective vascular effects of mineralocorticoid receptor antagonists in cardiovascular diseases associated with increased sympathetic activity, such as essential hypertension and heart failure.

**Summary**

Mineralocorticoid receptor activation is crucial for the vascular alterations induced by long-term β-AR stimulation, including increased vasoconstriction, uncoupling of endothelial nitric oxide synthase, and impaired anticontractile function of PVAT.
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SUPPLEMENTAL MATERIAL

Spironolactone prevents eNOS uncoupling and vascular dysfunction induced by β-adrenergic overstimulation: role of perivascular adipose tissue

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Supplemental methods

Blood pressure, heart rate and myocardial hypertrophic index

Under general anesthesia (urethane, 1.2 g.Kg\(^{-1}\), i.p.), a fluid-filled polyethylene catheter (PE-50) connected to a pressure transducer (MTL844 AdInstruments, Sydney-NSW, Australia) was placed on the right common carotid artery. Systolic blood pressure (SBP), diastolic blood pressure (DBP), and heart rate were monitored during 30 min (LabChart 7, AdInstruments). Next, the rats were euthanized, the heart was removed, and the ratio of ventricle (right and left) weight to body weight was calculated.

Vascular reactivity

The thoracic aorta was harvested and immediately placed in cold Krebs–Henseleit solution (in mM: 118 NaCl, 4.7 KCl, 25 NaHCO\(_3\), 2.5 CaCl\(_2\)-2H\(_2\)O, 1.2 KH\(_2\)PO\(_4\), 1.2 MgSO\(_4\)-7H\(_2\)O, 11 glicose and 0.01 EDTA; pH=7.4). Transverse aortic segments (3 mm long) with or without PVAT were obtained and placed in an organ chamber bath (Panlab Harvard Apparatus, Cornellà-Barcelona, Spain) maintained at 37°C with a resting tension of 1 g, as previously described\(^1\). Isometric tension was recorded using a force transducer (MLT0420, AdInstruments) connected to an acquisition system (PowerLab 8/30 system, AdInstruments).

After 30 min of equilibration period, aortic rings were exposed to 75 mM KCl to assess the maximum tension. Following wash, rings were pre-contracted with a submaximal concentration of phenylephrine (1 µM) to obtain a relaxation response curve to acetylcholine (0.001-10 µM) and to sodium nitroprusside (0.001-1 µM). The constrictor response to phenylephrine (0.001-10 µM) was assessed in the absence or presence of the nonselective NOS inhibitor, N\(^\omega\)-nitro-L-arginine methyl ester (L-NAME, 100 µM), superoxide dismutase (SOD, 150 U/mL) or tetrahydrobiopterin (BH\(_4\), 100 µM) added 30 min before concentration-response curves to phenylephrine were performed.
Reactive oxygen species (ROS) and NO detection

Frozen transverse sections (10 µm) from thoracic aorta and PVAT embedded in freezing medium were obtained. Slices were placed into a light-protected humidified chamber at 37°C and the following protocols were performed: 1) for NO measurement, slices from aorta and PVAT were incubated for 30 min with phosphate buffer (PB 0.1 mM, pH= 7.4) containing 2 mM CaCl$_2$ and 4,5-diaminofluorescein diacetate (DAF-2, 8 µM). BH$_4$ (100 µM) was applied in some PVAT slices in addition to DAF-2 in the latest 15 min; 2) for ROS analysis, slices were incubated with Krebs+HEPES buffer (in mM: 130 NaCl, 5.6 KCl, 2 CaCl$_2$, 0.24 MgCl$_2$, 8.3 HEPES and 11 glicose, pH= 7.4) plus dihydroethidium (DHE, 2 µM). The cell-permeable SOD mimetic MnTMPyP (25 µM) or L-NAME (1 mM) was topically applied in some aortic sections in addition to DHE.

Digital images were obtained with an epifluorescence microscope (Eclipse Ti-S, Nikon, Tokyo, Japan), using a fluorescein and a rhodamine filters, for DAF-2 and DHE analysis, respectively. The captured images under 20X objective were analyzed using Image J 1.46p software (National Institutes of Health, Bethesda- MD, USA) by measurement of the mean optical density of the fluorescence. Fluorescence was normalized by vessel or PVAT area (µm$^2$).

Western blot analysis

Total and subcellular protein fraction

Thoracic aorta and respective PVAT were homogenized in cold RIPA lysis buffer and added with 1 mM phenylmethylsulphonyl fluoride (PMSF), 1 mM Na$_3$VO$_4$ and 2 µL/mL protease inhibitor cocktail (PIC) to obtain total protein extract.

For nuclear and cytosol fractions, pulverized aortas were homogenized in cold HEPES lysis buffer [10 mM HEPES (pH=7.9), 1.5 mM MgCl$_2$, 10 mM KCl, 1 mM DTT, 0.5 mM PMSF and 2 µL/mL PIC] and then centrifuged for 10 min at 850 g. The supernatant was discarded and the pellet was re-suspended in lysis buffer containing 0.1% Triton-X. After 10 min, samples were centrifuged (10 min, 850 g) and the resulted supernatants were the cytosol protein fraction.
The resulted pellets were washed in ice-cold buffer [20 mM HEPES (pH= 7.9), 25% glycerol, 1.5 mM MgCl₂, 420 mM NaCl, 0.2 mM EDTA, 1.0 mM dithiothreitol, 0.5 mM PMSF and 2 μL/mL PIC] and then centrifuged (15 min at 16,000 g, 4°C) to obtain the supernatants containing nuclear proteins.

Protein extracts (aorta: 50 µg for total, 15 µg for nuclear and cytosolic fractions; PVAT: 100 µg for total) were electrophoretically separated on 7.5 or 10% acrylamide SDS–PAGE at room temperature and then transferred to PVDF membranes using a Mini Trans-Blot Cell system (Bio-Rad, Hercules- CA, USA). After blocked with 5% non-fat milk in TBST for 2 h at room temperature, membranes containing total homogenates were incubated with primary antibodies against eNOS, Ser¹¹⁷⁷ and Thr⁴⁹⁵ phospho-eNOS, HSP90, ERK1/2, anti-Thr²⁰²/²⁰⁴ phospho-ERK1/2, anti-Src and anti-Tyr⁴¹⁸ phospho-Src; anti-CYP11B1 (glucocorticoid synthase), anti-CYP11B2 (aldosterone synthase), 11β-hydroxysteroid dehydrogenase (11β-HSD) type 1 and type 2; anti-gamma-ENaC; membranes containing nuclear/cytosolic fractions were incubated with anti-MR antibody. After washing, membranes were incubated for 90 min with the horseradish peroxidase-conjugated IgG antibody according to each primary antibody used. Blots were detected with Pierce ECL Western Blotting Substrate (Thermo Scientific, Rockford-IL, USA) and visualized using ImageQuant LAS 4000 (GE Healthcare, Little Chalfont — Buckinghamshire, UK). The intensity was quantified by using the Image J 1.46p software (National Institutes of Health). Protein expression of α-actin (aorta), β-actin (PVAT), or Ponceau staining (nuclear/cytosolic fraction) were used to normalize expression of the evaluated proteins in each sample.

eNOS dimerization

eNOS dimerization was investigated as previously described²–³. Briefly, thoracic aortas were longitudinally opened and incubated for 1 h at 4°C in lysis buffer (50 mM Tris-HCl, pH= 8.0; 0.2% Nonidet P-40; 180 mM NaCl; 0.5 mM EDTA; 25 mM PMSF; 0.1 mM dithiothreitol; 2 μL/mL PIC). Non-boiled samples (75 µg) were separated by 6% SDS-PAGE at 4°C and transferred to PVDF membranes. After blockade (5% albumin in TBST), membranes were
incubated overnight at 4°C with primary antibody anti-eNOS (1:750), and then incubated for 90 min with the anti-mouse horseradish peroxidase-conjugated secondary antibody (1:10,000). eNOS dimer (260 kDa) and monomer (130 kDa) blots were detected as described in western blot analysis section and then the dimer to monomer ratio was calculated.

**RT-PCR analysis**

Frozen thoracic aortas were homogenized in TRizol and RNA (1 µg) were used to obtain the cDNA (TaqMan Reverse Transcription Reagents, Applied Biosystems, Foster City- CA, USA). PCR reactions were made in duplicate in a thermocycler (ABI PRISM 7000 Sequence Detection System, Applied Biosystems) using in the reaction for each sample 1 µL cDNA, 1 x TaqMan Universal PCR Master Mix (Applied Biosystems), and 20 x TaqMan Gene Expression Assay (Osteopontin – SPP1: Rn00681031_m1, Applied Biosystem) in a final volume of 20 µL. The amplification of GAPDH (Rn01775763_g1, Applied Biosystem) was used as the internal normalization. Analysis of relative gene expression data was performed using the \(2^{-\Delta\Delta CT}\) method.

**Determination of plasma and perivascular aldosterone and corticosterone levels**

Blood samples were collected via abdominal aorta and then centrifuged to obtain plasma samples. PVAT surrounding thoracic aorta was weighted and 100 mg of the tissue was homogenized in cold PBS 0.1 M (pH= 7.4). Protein was quantified using Bradford reagent. The enzymatic immunoassay was performed for aldosterone (Aldosterone EIA kit – Monoclonal No 10004377, Cayman Chemical, Ann Arbor- MI, USA) and corticosterone (Corticosterone EIA kit No 500651, Cayman Chemical) following the manufacturer’s instructions. Results were expressed as pg/mL for plasma samples and as pg/mL/mg protein for PVAT.

**Reagents**
Spironolactone and all reagents used for vascular reactivity, isoproterenol, Na₃VO₄, HEPES, Tris, dithiothreitol, PIC, BH₄ and DAF-2 were purchased from Sigma-Aldrich CO (Saint Louis-MO, USA). Losartan was purchased from Medley (Campinas- São Paulo, BR). DHE was purchased from Invitrogen (Grand Island, NY, USA), MnTMPyP from Calbiochem (San Diego, CA, USA) and RIPA lysis buffer from Merck Millipore (Billerica- MA, USA).

Primary antibodies anti-phospho-eNOS^{Ser1177}, anti-ERK1/2 and anti-phospho-ERK1/2 were purchased from Cell Signaling (Danvers- MA, USA); anti-eNOS from BD Transduction (Franklin Lakes- NJ, USA); anti-Src, anti-phospho-eNOS^{Thr495}, anti-CYP11B1 and anti-CYP11B2 from Merck Millipore; anti-HSP90, anti-11β-HSD1, anti-11β-HSD2 and anti-β-actin from Santa Cruz (Dallas- TX, USA); anti-MR and anti-α-actin from Abcam (Cambridge, UK); anti-gamma-ENaC from StressMarq (Victoria-BC, Canada) and anti-phospho-Src from Invitrogen. Secundary antibodies were obtained from Abcam and Bio Rad.
Supplemental references


### Supplemental table

**Table S1:** Morphometric parameters, blood pressure, and heart rate in experimental groups.

<table>
<thead>
<tr>
<th>Parameters</th>
<th>CT</th>
<th>LOS</th>
<th>SPI</th>
<th>ISO</th>
<th>ISO+ LOS</th>
<th>ISO+ SPI</th>
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<tr>
<td>BW (g)</td>
<td>374 ± 14</td>
<td>397 ± 8</td>
<td>369 ± 8</td>
<td>365 ± 8</td>
<td>380 ± 7</td>
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<tr>
<td>VW/ BW (mg g⁻¹)</td>
<td>2.64 ± 0.06</td>
<td>2.52 ± 0.03</td>
<td>2.48 ± 0.03</td>
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<td>3.14 ± 0.10*</td>
<td>2.98 ± 0.03*</td>
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<td>SBP (mmHg)</td>
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<td>116 ± 3.6</td>
<td>126 ± 3.3</td>
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<td>122 ± 2.5</td>
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<td>DBP (mmHg)</td>
<td>74 ± 2.1</td>
<td>58 ± 3.9*</td>
<td>81 ± 4.0</td>
<td>81 ± 2.7</td>
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<tr>
<td>HR (bpm)</td>
<td>408 ± 8</td>
<td>412 ± 14</td>
<td>385 ±11</td>
<td>386 ± 6</td>
<td>400 ± 14</td>
<td>382 ± 9</td>
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</table>

Values are expressed as mean ± SEM (N=8-12). CT: Control; ISO: isoproterenol; LOS: losartan; SPI: spironolactone; SBP: systolic blood pressure; DBP: diastolic blood pressure; HR: heart rate; BW: body weight; VW: ventricle weight. Two-way ANOVA: *p<0.05 vs. CT; #p<0.05 vs. ISO.
Supplemental figures

Figure S1: Isoproterenol (ISO) alone or combined with losartan (LOS) or with spironolactone (SPI) did not modify the vasorelaxation to acetylcholine (A) or to sodium nitroprusside (B) compared with control group (CT). Two-way ANOVA (n=4-12 in each experimental group).
Figure S2: Losartan (LOS) treatment did not ameliorate the impaired NO bioavailability induced by 7-day isoproterenol treatment (ISO). Aorta from ISO-treated rats showed reduced diaminofluorescein (DAF-2) fluorescence indicating reduced NO levels that were not modified by co-treatment with LOS. Representative fluorescent images of DAF-2 are shown in left side of bar graph (20x, white bar=100 µm). Bar graphs represents mean ± SEM; number of animals used in each group is indicated in the bars. Two-way ANOVA: p<0.05 * vs. CT; + vs. LOS.
Figure S3: Ser1177 (A, B) and Thr495 (C, D) eNOS phosphorylation in thoracic aorta from control (CT) and isoproterenol (ISO) groups without or with treatment with spironolactone (SPI) or (LOS). Phospho-eNOS was quantified as ratio of total eNOS expression. *P<0.05 vs. CT; ^p<0.05 vs. SPI or LOS: Two-way ANOVA. Number of animals used in each group is showed into the bars.
Figure S4: Losartan (LOS) treatment did not ameliorate vascular oxidative stress induced by 7-day isoproterenol treatment (ISO). Oxidative fluorescent signal to hidroethidine (DHE) was obtained under basal condition or incubated with L-NAME (1 mM) in thoracic aortic sections from CT, ISO, LOS and ISO+LOS groups. Representative fluorescent images of DHE are shown in left side of bar graphs (20x, white bar=100 µm). Bar graphs represent mean ± SEM; number of animals used in each group is indicated in the bars. Two-way ANOVA: p<0.05 * vs. CT; + vs. LOS; # vs. basal condition.
Figure S5: Protein expression of CYP11B1 (A) and CYP11B2 (B) in aortic PVAT from control (CT) and isoproterenol (ISO) groups. Upper panels show representative blots. Data represent mean ± SEM; number of animals is indicated into the bars. Student’s t-test: p>0.05.