

## Aging and Adrenal Aldosterone Production

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Aldosterone, the primary mineralocorticoid, is synthesized in the outer zone of the adrenal cortex called the zona glomerulosa (ZG). The production of aldosterone is tightly regulated by angiotensin II (Ang II) and circulating potassium levels.<sup>1</sup> Physiologically, aldosterone plays a key role in the maintenance of intravascular volume and blood pressure through sodium retention in the kidney. Excess aldosterone causes hypertension and induces cardiovascular complications.<sup>2-5</sup> The autonomous secretion of aldosterone, independent of Ang II and sodium status, is known as primary aldosteronism (PA). PA is the most common cause of endocrine-related hypertension with a prevalence of 5% to 10% in hypertensive population<sup>6-10</sup> and  $\approx 20\%$  in resistant hypertension.<sup>11-13</sup> Because of the increased risk for cardiovascular complications in patients with PA, early detection of the disease and targeted treatment is recommended.<sup>14</sup>

The molecular pathogenesis of PA was largely unknown until recently. The development of specific antibodies against aldosterone synthase (CYP11B2), which is required for the final steps of aldosterone production, has allowed the detection of aldosterone-producing cells in resected adrenals.<sup>15,16</sup> Using these antibodies, non-neoplastic foci of CYP11B2-expressing cells called aldosterone-producing cell clusters (APCC)<sup>15</sup> have been identified in adrenal tissues adjacent to aldosterone-producing adenomas (APA) and in normal human adrenal glands without tumor or hyperplasia.<sup>15-20</sup> Studies have shown that APCC are a common occurrence in normal human adrenals.<sup>21-23</sup> Because circulating renin and Ang II levels are suppressed in patients with PA, the observation of APCC adjacent to APA suggested that APCC may represent a source of autonomous and renin-independent aldosterone secretion, perhaps even a precursor to APA.<sup>24</sup>

Over the past 6 years, somatic and germline mutations that cause inappropriate aldosterone production have been identified in patients with PA. Most of the mutations cause increased intracellular calcium ( $\text{Ca}^{2+}$ ) levels, resulting in activated *CYP11B2* transcription and elevation of aldosterone production.<sup>24-26</sup> These somatic mutations have also been observed in APCC in normal adrenal glands.<sup>23,27</sup> Recent histopathologic studies demonstrated that older age is associated with greater adrenal APCC content<sup>21-23</sup> and a concomitant biochemical phenotype that supports progressive autonomous aldosteronism.<sup>21</sup> These findings provide a possible link between age-related

histological changes in adrenal CYP11B2 expression and age-related physiological changes in aldosterone secretion. In this review, we cover recent topics on physiology and dysregulation of aldosterone production and discuss potential age-related effects on the aldosterone and adrenal physiology.

### Human Adrenal Zonation and Steroid Production

Human adrenal glands are composed of an outer cortex and an inner medulla. The adrenal cortex is further divided into 3 functionally distinct zones; ZG, zona fasciculata, and zona reticularis (Figure 1A). In the ZG, the mineralocorticoid aldosterone is produced with the physiological role of maintaining fluid and electrolyte balance. The production of aldosterone is mainly regulated by Ang II, potassium ( $\text{K}^+$ ), and adrenocorticotropic hormone.<sup>1</sup> Glucocorticoids are synthesized in the zona fasciculata under the regulation of adrenocorticotropic hormone. The zona reticularis is also mainly regulated by adrenocorticotropic hormone and produces adrenal androgens, including dehydroepiandrosterone and dehydroepiandrosterone sulfate. There is considerable evidence that aging is associated with a decline in circulating levels of dehydroepiandrosterone and dehydroepiandrosterone sulfate.<sup>28</sup> The decrease in these adrenal steroids results from intra-adrenal changes that include a decrease in zona reticularis size and expression of steroidogenic enzymes required for dehydroepiandrosterone synthesis.<sup>29,30</sup> In contrast to dehydroepiandrosterone, little is known about the age-related adrenal changes related to aldosterone synthesis.

ZG synthesis of aldosterone occurs through the action of a series of enzymes, including cholesterol side-chain cleavage (CYP11A1), type 2  $3\beta$ -hydroxysteroid dehydrogenase (HSD3B2), 21-hydroxylase (CYP21A2), and CYP11B2<sup>1</sup> (Figure 1B). Of these enzymes, CYP11B2 is expressed specifically in the ZG, whereas expression of  $11\beta$ -hydroxylase (CYP11B1), which is required for the final step in the biosynthesis of cortisol, is limited to the zona fasciculata and zona reticularis.<sup>31</sup> This functional zonation acts to compartmentalize aldosterone production to the ZG.<sup>32,33</sup> On the basis of the critical nature of CYP11B2 in aldosterone production, we and others have studied the cell-signaling mechanisms regulating this enzyme in normal and pathological conditions within the adrenal.

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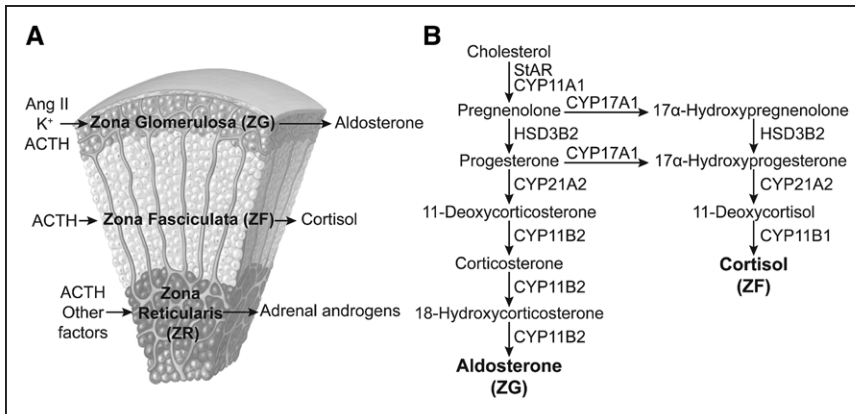
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(*Hypertension*. 2018;71:218-223. DOI: 10.1161/HYPERTENSIONAHA.117.10391.)

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*Hypertension* is available at <http://hyper.ahajournals.org>

DOI: 10.1161/HYPERTENSIONAHA.117.10391



**Figure 1.** Adrenal zonation and steroidogenic pathway. **A**, Adrenal cortex zonation with the primary regulators of steroid production. **B**, Steroidogenic pathway for the production of aldosterone and cortisol. Aldosterone and cortisol production occur in the zona glomerulosa (ZG) and zona fasciculata (ZF), respectively. ACTH indicates adrenocorticotropic hormone; and StAR, steroidogenic acute regulatory protein.

### Physiological and Pathological Regulation of Adrenal Cell Aldosterone Production

The primary physiological regulators of aldosterone production are Ang II,  $K^+$ , and adrenocorticotropic hormone. Other factors that have been proposed to modulate aldosterone production include serotonin,<sup>34,35</sup> estrogen,<sup>36</sup> parathyroid hormone,<sup>37,38</sup> vasopressin,<sup>39,40</sup> endothelin-1,<sup>41,42</sup> and very-low-density lipoprotein.<sup>43–45</sup> A recent study demonstrated that leptin induces *CYP11B2* expression and aldosterone production via  $Ca^{2+}$ -dependent mechanisms.<sup>46</sup> Several signaling pathways are involved in adrenal cell aldosterone production. The key intracellular pathways include  $Ca^{2+}$ ,<sup>47,48</sup> cAMP,<sup>49,50</sup> and phospholipase D signaling.<sup>51–53</sup> Of these, the  $Ca^{2+}$ -signaling pathway seems to be the most important in both physiological and pathological conditions. Both Ang II and  $K^+$  use  $Ca^{2+}$  for physiological activation, whereas inappropriate aldosterone production, as seen in PA, is primarily regulated by somatic and germline mutations that dysregulate and raise intracellular  $Ca^{2+}$ .

Mutations in *KCNJ5*,<sup>54</sup> *ATP1A1*,<sup>55</sup> *ATP2B3*,<sup>55</sup> *CACNAID*,<sup>56,57</sup> and *CACNAIH*<sup>58</sup> (aldosterone-driver genes) have been identified in APA and familial hyperaldosteronism. In vitro studies have shown that these mutations cause increased aldosterone production.<sup>59–64</sup> The *KCNJ5* gene encodes the inward-rectifying potassium channel GIRK4, and somatic mutations in this gene are considered to be the most prevalent genetic alteration in APA.<sup>65–67</sup> Mutations in *KCNJ5* gene cause a loss of ion selectivity, increased  $Na^+$  conductance, cell membrane depolarization, and voltage-gated calcium channel opening, leading to increased intracellular  $Ca^{2+}$  concentration.<sup>54,59,68</sup> The *ATP1A1* gene encodes the  $\alpha 1$ -subunit of the  $Na^+/K^+$  ATPase, which acts to maintain the resting cell membrane potential and electric excitability. *ATP1A1* mutations disrupt this function and cause cell membrane depolarization and cellular acidification because of a pathological  $H^+$  leak, resulting in aldosterone overproduction.<sup>55,61,69</sup> The *ATP2B3* gene encodes the plasma membrane  $Ca^{2+}$  ATPase isoform 3, which regulates intracellular  $Ca^{2+}$  homeostasis by transporting cytoplasmic  $Ca^{2+}$  out of the cells. Mutations in *ATP2B3* gene seem to increase intracellular  $Ca^{2+}$  levels through a reduced  $Ca^{2+}$  export as a result of a loss of pump function and nonphysiological  $Na^+$  and possible  $Ca^{2+}$  permeability to the cells.<sup>62</sup> The *CACNAID* gene encodes the voltage-dependent L-type calcium channel subunit  $\alpha$ -1D

( $Ca_v 1.3$ ). Mutations in *CACNAID* gene cause channel activation at less depolarized potentials, leading to increased  $Ca^{2+}$  influx and aldosterone overproduction.<sup>56,57,63,70</sup> The *CACNAIH* gene encodes the voltage-dependent T-type calcium channel subunit  $\alpha$ -1H ( $Ca_v 3.2$ ). In a recent study, a recurrent germline mutation in *CACNAIH* gene (p.Met1549Val) was identified in early-onset PA patients.<sup>58</sup> In vitro studies demonstrated that the *CACNAIH* mutation caused impaired channel inactivation and a shift of activation to less depolarized potentials, leading to increased  $Ca^{2+}$  entry and elevated aldosterone production.<sup>58,64</sup>

The cellular origins of APA with somatic mutations remain an area of active research. If the adrenal accumulation of somatic mutations follows other tissues, it would be expected that there would be an age dependence. We and others have approached this research by focusing on *CYP11B2* because it is required for aldosterone synthesis and its expression is highly cell specific within the adrenal. Development of specific antibodies against human *CYP11B2* and *CYP11B1* has provided tools to better study adrenal functional histopathology.<sup>15,16</sup> Nishimoto et al<sup>15</sup> reported that there are 2 patterns of *CYP11B2* expression within the adrenal: (1) conventional zonation with ZG cells expressing *CYP11B2* and (2) variegated zonation consisting of a subcapsular non-neoplastic cell foci expressing *CYP11B2* named APCC. The expression pattern of steroidogenic enzymes in APCC, that is, high expression of *HSD3B2* and *CYP11B2* and low expression of *17\alpha*-hydroxylase (*CYP17A1*) and *CYP11B1* (needed for cortisol biosynthesis), supports the ability of APCC to produce aldosterone.<sup>15</sup> APCC have also been identified in adrenal tissues adjacent to APA<sup>15,17–20</sup>; given that renin and Ang II are suppressed in PA caused by APA, these observations supported APCC as non-neoplastic sources of autonomous and renin-independent aldosterone secretion. Of note, before the development of *CYP11B2* antibodies, in situ hybridization methods allowed the identification of cell foci expressing *CYP11B2* mRNA but no *CYP11B1* or *CYP17* mRNA.<sup>71</sup>

### Adrenal ZG and Aldosterone-Producing Cell Clusters in Aging

Age-related histological changes in adrenal *CYP11B2* expression have been investigated. There is growing evidence for an age-dependent accumulation of APCC in adrenal glands.<sup>21–23</sup> Further, there is age-related loss of the classic continuous *CYP11B2* expression pattern within the ZG that

is frequently observed in young adrenal glands (Figure 2).<sup>21,72</sup> Collectively, older adrenal glands have less normal CYP11B2 expression in the ZG and greater amount of APCC. Although there is no direct evidence to explain why such age-related changes in CYP11B2 expression patterns might occur, potential considerations may include genomic, epigenetic, or other environmental factors. The histological findings suggest a transition from continuous CYP11B2 expression in the ZG to APCC predominance with advancing age<sup>21</sup> and raise the hypothesis that with aging, there may be a progressive transition from normal physiological aldosterone regulation to autonomous and renin-independent aldosterone secretion. If true, this clinical phenotype associated with these age-related histopathologic changes should involve greater autonomous aldosteronism and higher risk for aldosterone-mediated cardiovascular disease.

To better define the cellular characteristics of APCC, we performed transcriptome analysis and targeted next-generation sequencing analysis on APCC isolated from normal adrenal glands from kidney donors.<sup>27</sup> Transcriptome analysis demonstrated that APCC mRNA profile had similar characteristics to those of ZG but with higher *CYP11B2* expression in APCC, indicating increased capacity to produce aldosterone in APCC.<sup>27</sup> Mutation analysis revealed that 8 out of 23 APCC (35%) had known aldosterone-driver somatic mutations in genes including *CACNAID* and *ATPIA1*.<sup>27</sup> A recent study using a modified targeted next-generation sequencing protocol identified somatic mutations in 21 of 61 APCC (34%) in adrenals from normotensive Japanese autopsy cases.<sup>23</sup> No *KCNJ5* mutation, which is the most prevalent somatic mutation in APA, was identified in either American or Japanese cohorts.<sup>23,27</sup> In contrast to APA, both studies suggest that APCC are most likely to have aldosterone-driver mutations in the *CACNAID* gene.<sup>23,27</sup> These findings suggest that L-type calcium channel blockers should be considered as a targeted treatment option that could reduce inappropriate autonomous aldosterone production from APCC. Interestingly, in 1 early-onset PA patient with germline de novo *CACNAID* mutation (p.Gly403Asp), treatment with a calcium channel blocker, amlodipine, resulted in normalization of blood pressure and resolution of biventricular hypertrophy that had been present since birth.<sup>57</sup>

Although somatic mutations in APCC support the concept of renin-independent aldosterone production in APCC, genetic characteristics of majority of the APCC are still unknown.

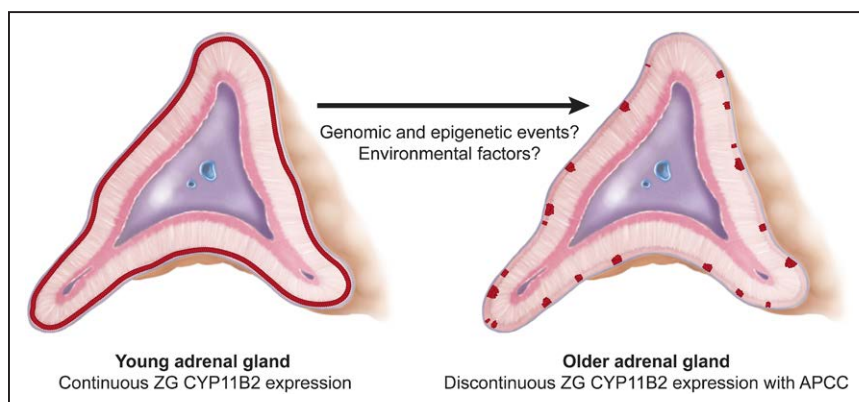
Further, because previous research on human APCC has been conducted from either postmortem adrenal specimen or adrenals containing APA, the biochemical phenotype of APCC has not been specifically quantified. Future clinical studies are needed to directly assess whether APCC autonomously secrete aldosterone.

### Renin–Angiotensin–Aldosterone System in Aging

With aging, there is a decline in normal function of several physiological systems. Consistent with the histological findings of age-related changes in adrenal CYP11B2 expression patterns, physiological changes in the renin–angiotensin–aldosterone system seem to occur with advancing age. There have been several studies with relatively small sample size suggesting that older individuals may secrete less aldosterone and have lower plasma renin activity.<sup>73–77</sup> Specifically, studies have shown that older age is associated with a blunted ability to secrete aldosterone despite stimulation by its major regulators: Ang II with sodium restriction<sup>74</sup> and potassium during intravenous infusion.<sup>78</sup> We recently demonstrated the dynamics of aldosterone physiology in a large cross-sectional study that included a broad age continuum.<sup>21</sup> The most notable observations were a strong and significant association between older age with higher aldosterone:renin ratio and a blunted ability to secrete aldosterone with sodium restriction.<sup>21</sup> Taken together, older age was associated with greater autonomous aldosterone secretion and less physiological aldosterone secretion. Given the aforementioned age-related histopathologic changes in CYP11B2 expression, we hypothesized that this clinical phenotype may have been the consequence of parallel age-related increases in adrenal APCC content and concomitant decreases in normal adrenal CYP11B2 expression.

### Clinical Perspectives

The concept of age-related autonomous aldosteronism may provide new avenues to investigate the pathogenesis and treatments for hypertension and cardiovascular diseases. Recent studies have demonstrated that autonomous aldosterone production and even overt PA can be detected in normotensive subjects<sup>79,80</sup>; in parallel with the finding that a subset of adrenal glands from normotensives have APCC-harboring somatic mutations in aldosterone-driver genes,<sup>23</sup> this may suggest that the spectrum of PA can range from mild and subclinical in normotension (possibly as



**Figure 2.** Age-related histological changes in human adrenal glands. Continuous CYP11B2 expression in the zona glomerulosa (ZG) is often seen in young adrenal glands, whereas older adrenals have less normal CYP11B2 expression and more aldosterone-producing cell clusters (APCC). Potential effects that might cause the age-associated increase in CYP11B2-positive APCC include genomic, epigenetic events or environmental factors.

a result of autonomous aldosterone production from APCC) to more severe in hypertension (possibly as a result of a combination of APCC and neoplastic and hyperplastic processes such as APA and bilateral adrenal hyperplasia).<sup>81</sup> Consistent with this, normotensive patients with evidence for subclinical PA have a significantly higher risk for developing incident hypertension.<sup>79,81,82</sup> A recent population-based cohort study demonstrated that older normotensives were more likely to have a suppressed renin phenotype and that higher aldosterone levels in the context of this renin suppression were associated with a substantially higher risk for developing hypertension.<sup>81</sup> Further, physiological investigations have also shown age-related declines in 11 $\beta$ -hydroxysteroid dehydrogenase type 2 activity that result in a phenotype of renin suppression and cortisol-mediated mineralocorticoid receptor activation.<sup>83</sup> Given the accumulating evidence suggesting age-related histopathologic and biochemical changes consistent with renin-independent aldosteronism and MR activation, future studies may investigate to what extent the essential hypertension of aging may be mediated by mineralocorticoids, such as aldosterone and cortisol.

### Acknowledgments

We would like to thank Diantha La Vine for assistance in preparing the medical illustrations.

### Sources of Funding

This work was supported by grants from the American Heart Association (17SDG33660447) to K. Nanba and the National Institutes of Diabetes and Digestive and Kidney Disease (DK106618) to W.E. Rainey. A. Vaidya was supported by the National Institutes of Diabetes and Digestive and Kidney Disease under award R01 DK107407, by the National Heart, Lung, and Blood Institute under award K23 HL111771, and by grant 2015085 from the Doris Duke Charitable Foundation.

### Disclosures

None.

### References

- Hattangady NG, Olala LO, Bollag WB, Rainey WE. Acute and chronic regulation of aldosterone production. *Mol Cell Endocrinol*. 2012;350:151–162. doi: 10.1016/j.mce.2011.07.034.
- Milliez P, Girerd X, Plouin PF, Blacher J, Safar ME, Mourad JJ. Evidence for an increased rate of cardiovascular events in patients with primary aldosteronism. *J Am Coll Cardiol*. 2005;45:1243–1248. doi: 10.1016/j.jacc.2005.01.015.
- Catena C, Colussi G, Nadalini E, Chiuch A, Baroselli S, Lapenna R, Sechi LA. Cardiovascular outcomes in patients with primary aldosteronism after treatment. *Arch Intern Med*. 2008;168:80–85. doi: 10.1001/archinternmed.2007.33.
- Mulatero P, Monticone S, Bertello C, Viola A, Tizzani D, Iannaccone A, Crudo V, Burrello J, Milan A, Rabbia F, Veglio F. Long-term cardio- and cerebrovascular events in patients with primary aldosteronism. *J Clin Endocrinol Metab*. 2013;98:4826–4833. doi: 10.1210/jc.2013-2805.
- Savard S, Amar L, Plouin PF, Steichen O. Cardiovascular complications associated with primary aldosteronism: a controlled cross-sectional study. *Hypertension*. 2013;62:331–336. doi: 10.1161/HYPERTENSIONAHA.113.01060.
- Rossi GP, Bernini G, Caliumi C, et al; PAPY Study Investigators. A prospective study of the prevalence of primary aldosteronism in 1,125 hypertensive patients. *J Am Coll Cardiol*. 2006;48:2293–2300. doi: 10.1016/j.jacc.2006.07.059.
- Fardella CE, Mosso L, Gómez-Sánchez C, Cortés P, Soto J, Gómez L, Pinto M, Huete A, Oestreicher E, Foradori A, Montero J. Primary hyperaldosteronism in essential hypertensives: prevalence, biochemical profile, and molecular biology. *J Clin Endocrinol Metab*. 2000;85:1863–1867. doi: 10.1210/jcem.85.5.6596.
- Loh KC, Koay ES, Khaw MC, Emmanuel SC, Young WF Jr. Prevalence of primary aldosteronism among Asian hypertensive patients in Singapore. *J Clin Endocrinol Metab*. 2000;85:2854–2859. doi: 10.1210/jcem.85.8.6752.
- Lim PO, Dow E, Brennan G, Jung RT, MacDonald TM. High prevalence of primary aldosteronism in the Tayside hypertension clinic population. *J Hum Hypertens*. 2000;14:311–315.
- Monticone S, Burrello J, Tizzani D, Bertello C, Viola A, Buffolo F, Gabetti L, Mengozzi G, Williams TA, Rabbia F, Veglio F, Mulatero P. Prevalence and clinical manifestations of primary aldosteronism encountered in primary care practice. *J Am Coll Cardiol*. 2017;69:1811–1820. doi: 10.1016/j.jacc.2017.01.052.
- Calhoun DA, Nishizaka MK, Zaman MA, Thakkar RB, Weissmann P. Hyperaldosteronism among black and white subjects with resistant hypertension. *Hypertension*. 2002;40:892–896.
- Strauch B, Zelinka T, Hampf M, Bernhardt R, Widimsky J Jr. Prevalence of primary hyperaldosteronism in moderate to severe hypertension in the Central Europe region. *J Hum Hypertens*. 2003;17:349–352. doi: 10.1038/sj.jhh.1001554.
- Gallay BJ, Ahmad S, Xu L, Toivola B, Davidson RC. Screening for primary aldosteronism without discontinuing hypertensive medications: plasma aldosterone-renin ratio. *Am J Kidney Dis*. 2001;37:699–705.
- Funder JW, Carey RM, Mantero F, Murad MH, Reincke M, Shibata H, Stowasser M, Young WF Jr. The management of primary aldosteronism: case detection, diagnosis, and treatment: an Endocrine Society Clinical Practice Guideline. *J Clin Endocrinol Metab*. 2016;101:1889–1916. doi: 10.1210/jc.2015-4061.
- Nishimoto K, Nakagawa K, Li D, Kosaka T, Oya M, Mikami S, Shibata H, Itoh H, Mitani F, Yamazaki T, Ogishima T, Suematsu M, Mukai K. Adrenocortical zonation in humans under normal and pathological conditions. *J Clin Endocrinol Metab*. 2010;95:2296–2305. doi: 10.1210/jc.2009-2010.
- Gomez-Sanchez CE, Qi X, Velarde-Miranda C, Plonczynski MW, Parker CR, Rainey W, Satoh F, Maekawa T, Nakamura Y, Sasano H, Gomez-Sanchez EP. Development of monoclonal antibodies against human CYP11B1 and CYP11B2. *Mol Cell Endocrinol*. 2014;383:111–117. doi: 10.1016/j.mce.2013.11.022.
- Nanba K, Tsuiji M, Sawai K, Mukai K, Nishimoto K, Usui T, Tagami T, Okuno H, Yamamoto T, Shimatsu A, Katabami T, Okumura A, Kawa G, Tanabe A, Naruse M. Histopathological diagnosis of primary aldosteronism using CYP11B2 immunohistochemistry. *J Clin Endocrinol Metab*. 2013;98:1567–1574. doi: 10.1210/jc.2012-3726.
- Volpe C, Höög A, Ogishima T, Mukai K, Lu M, Thorén M, Hamberger B. Immunohistochemistry improves histopathologic diagnosis in primary aldosteronism. *J Clin Pathol*. 2013;66:351–354. doi: 10.1136/jclinpath-2012-201287.
- Dekkers T, ter Meer M, Lenders JW, Hermus AR, Schultze Kool L, Langenhuijsen JF, Nishimoto K, Ogishima T, Mukai K, Azizan EA, Tops B, Deinum J, Küsters B. Adrenal nodularity and somatic mutations in primary aldosteronism: one node is the culprit? *J Clin Endocrinol Metab*. 2014;99:E1341–E1351. doi: 10.1210/jc.2013-4255.
- Monticone S, Castellano I, Versace K, Lucatello B, Veglio F, Gomez-Sanchez CE, Williams TA, Mulatero P. Immunohistochemical, genetic and clinical characterization of sporadic aldosterone-producing adenomas. *Mol Cell Endocrinol*. 2015;411:146–154. doi: 10.1016/j.mce.2015.04.022.
- Nanba K, Vaidya A, Williams GH, Zheng I, Else T, Rainey WE. Age-related autonomous aldosteronism. *Circulation*. 2017;136:347–355. doi: 10.1161/CIRCULATIONAHA.117.028201.
- Nishimoto K, Seki T, Hayashi Y, Mikami S, Al-Eyd G, Nakagawa K, Morita S, Kosaka T, Oya M, Mitani F, Suematsu M, Kabe Y, Mukai K. Human adrenocortical remodeling leading to aldosterone-producing cell cluster generation. *Int J Endocrinol*. 2016;2016:7834356. doi: 10.1155/2016/7834356.
- Omata K, Anand SK, Hovelson DH, Liu CJ, Yamazaki Y, Nakamura Y, Ito S, Satoh F, Sasano H, Rainey WE, Tomlins SA. Aldosterone-producing cell clusters frequently harbor somatic mutations and accumulate with age in normal adrenals. *J Endocr Soc*. 2017;1:787–799. doi: 10.1210/js.2017-00134.
- Zennaro MC, Boulkroun S, Fernandes-Rosa F. Genetic causes of functional adrenocortical adenomas. *Endocr Rev*. 2017. doi: 10.1210/er.2017-00189.
- Azizan EA, Brown MJ. Novel genetic determinants of adrenal aldosterone regulation. *Curr Opin Endocrinol Diabetes Obes*. 2016;23:209–217. doi: 10.1097/MED.0000000000000255.

26. Monticone S, Else T, Mulatero P, Williams TA, Rainey WE. Understanding primary aldosteronism: impact of next generation sequencing and expression profiling. *Mol Cell Endocrinol*. 2015;399:311–320. doi: 10.1016/j.mce.2014.09.015.
27. Nishimoto K, Tomlins SA, Kuick R, Cani AK, Giordano TJ, Hovelson DH, Liu CJ, Sanjanwala AR, Edwards MA, Gomez-Sanchez CE, Nanba K, Rainey WE. Aldosterone-stimulating somatic gene mutations are common in normal adrenal glands. *Proc Natl Acad Sci USA*. 2015;112:E4591–E4599. doi: 10.1073/pnas.1505529112.
28. Hornsby PJ. Aging of the human adrenal cortex. *Ageing Res Rev*. 2002;1:229–242.
29. Dharia S, Slane A, Jian M, Conner M, Conley AJ, Brissie RM, Parker CR Jr. Effects of aging on cytochrome b5 expression in the human adrenal gland. *J Clin Endocrinol Metab*. 2005;90:4357–4361. doi: 10.1210/jc.2005-0017.
30. Parker CR Jr, Mixon RL, Brissie RM, Grizzle WE. Aging alters zonation in the adrenal cortex of men. *J Clin Endocrinol Metab*. 1997;82:3898–3901. doi: 10.1210/jcem.82.11.4507.
31. Rainey WE. Adrenal zonation: clues from 11 $\beta$ -hydroxylase and aldosterone synthase. *Mol Cell Endocrinol*. 1999;151:151–160.
32. Domalik LJ, Chaplin DD, Kirkman MS, Wu RC, Liu WW, Howard TA, Seldin MF, Parker RL. Different isozymes of mouse 11 $\beta$ -hydroxylase produce mineralocorticoids and glucocorticoids. *Mol Endocrinol*. 1991;5:1853–1861. doi: 10.1210/mend-5-12-1853.
33. Ogishima T, Suzuki H, Hata J, Mitani F, Ishimura Y. Zone-specific expression of aldosterone synthase cytochrome P-450 and cytochrome P-45011 $\beta$  in rat adrenal cortex: histochemical basis for the functional zonation. *Endocrinology*. 1992;130:2971–2977. doi: 10.1210/endo.130.5.1572304.
34. Rocco S, Ambroz C, Aguilera G. Interaction between serotonin and other regulators of aldosterone secretion in rat adrenal glomerulosa cells. *Endocrinology*. 1990;127:3103–3110. doi: 10.1210/endo-127-6-3103.
35. Mantero F, Opocher G, Boscaro M, Armanini D. Effect of serotonin on plasma aldosterone in man. *J Endocrinol Invest*. 1982;5:97–99. doi: 10.1007/BF03350498.
36. Caroccia B, Seccia TM, Campos AG, Gioco F, Kuppusamy M, Ceolotto G, Guerzoni E, Simonato F, Maresio S, Lenzini L, Fassina A, Rossi GP. GPER-1 and estrogen receptor- $\beta$  ligands modulate aldosterone synthesis. *Endocrinology*. 2014;155:4296–4304. doi: 10.1210/en.2014-1416.
37. Isales CM, Barrett PQ, Brines M, Bollag W, Rasmussen H. Parathyroid hormone modulates angiotensin II-induced aldosterone secretion from the adrenal glomerulosa cell. *Endocrinology*. 1991;129:489–495. doi: 10.1210/endo-129-1-489.
38. Mazzocchi G, Aragona F, Malendowicz LK, Nussdorfer GG. PTH and PTH-related peptide enhance steroid secretion from human adrenocortical cells. *Am J Physiol Endocrinol Metab*. 2001;280:E209–E213.
39. Woodcock EA, Mcleod JK, Johnston CI. Vasopressin stimulates phosphatidylinositol turnover and aldosterone synthesis in rat adrenal glomerulosa cells: comparison with angiotensin II. *Endocrinology*. 1986;118:2432–2436. doi: 10.1210/endo-118-6-2432.
40. Guillon G, Grazzini E, Andrez M, Breton C, Trueba M, Serradeil-LeGal C, Boccara G, Derick S, Chouinard L, Gallo-Payet N. Vasopressin: a potent autocrine/paracrine regulator of mammal adrenal functions. *Endocr Res*. 1998;24:703–710.
41. Cozza EN, Gomez-Sanchez CE, Foecking MF, Chiou S. Endothelin binding to cultured calf adrenal zona glomerulosa cells and stimulation of aldosterone secretion. *J Clin Invest*. 1989;84:1032–1035. doi: 10.1172/JCI114226.
42. Rossi GP, Albertin G, Bova S, Belloni AS, Fallo F, Pagotto U, Trevisi L, Palù G, Pessina AC, Nussdorfer GG. Autocrine-paracrine role of endothelin-1 in the regulation of aldosterone synthase expression and intracellular Ca<sup>2+</sup> in human adrenocortical carcinoma NCI-H295 cells. *Endocrinology*. 1997;138:4421–4426. doi: 10.1210/endo.138.10.5267.
43. Xing Y, Rainey WE, Apolzan JW, Francone OL, Harris RB, Bollag WB. Adrenal cell aldosterone production is stimulated by very-low-density lipoprotein (VLDL). *Endocrinology*. 2012;153:721–731. doi: 10.1210/en.2011-1752.
44. Tsai YY, Rainey WE, Pan ZQ, Frohman MA, Choudhary V, Bollag WB. Phospholipase D activity underlies very-low-density lipoprotein (VLDL)-induced aldosterone production in adrenal glomerulosa cells. *Endocrinology*. 2014;155:3550–3560. doi: 10.1210/en.2014-1159.
45. Tsai YY, Rainey WE, Johnson MH, Bollag WB. VLDL-activated cell signaling pathways that stimulate adrenal cell aldosterone production. *Mol Cell Endocrinol*. 2016;433:138–146. doi: 10.1016/j.mce.2016.05.018.
46. Huby AC, Antonova G, Groenendyk J, Gomez-Sanchez CE, Bollag WB, Filosa JA, Belin de Chantemèle EJ. Adipocyte-derived hormone leptin is a direct regulator of aldosterone secretion, which promotes endothelial dysfunction and cardiac fibrosis. *Circulation*. 2015;132:2134–2145. doi: 10.1161/CIRCULATIONAHA.115.018226.
47. Barrett PQ, Bollag WB, Isales CM, McCarthy RT, Rasmussen H. Role of calcium in angiotensin II-mediated aldosterone secretion. *Endocr Rev*. 1989;10:496–518. doi: 10.1210/edrv-10-4-496.
48. Kojima I, Kojima K, Kreutter D, Rasmussen H. The temporal integration of the aldosterone secretory response to angiotensin occurs via two intracellular pathways. *J Biol Chem*. 1984;259:14448–14457.
49. Kojima I, Kojima K, Rasmussen H. Role of calcium and cAMP in the action of adrenocorticotropin on aldosterone secretion. *J Biol Chem*. 1985;260:4248–4256.
50. Gallo-Payet N, Grazzini E, Côté M, Chouinard L, Chorvátová A, Bilodeau L, Payet MD, Guillon G. Role of Ca<sup>2+</sup> in the action of adrenocorticotropin in cultured human adrenal glomerulosa cells. *J Clin Invest*. 1996;98:460–466. doi: 10.1172/JCI118812.
51. Bollag WB, Barrett PQ, Isales CM, Liscovitch M, Rasmussen H. A potential role for phospholipase-D in the angiotensin-II-induced stimulation of aldosterone secretion from bovine adrenal glomerulosa cells. *Endocrinology*. 1990;127:1436–1443. doi: 10.1210/endo-127-3-1436.
52. Bollag WB, Jung E, Calle RA. Mechanism of angiotensin II-induced phospholipase D activation in bovine adrenal glomerulosa cells. *Mol Cell Endocrinol*. 2002;192:7–16.
53. Zheng X, Bollag WB. AngII induces transient phospholipase D activity in the H295R glomerulosa cell model. *Mol Cell Endocrinol*. 2003;206:113–122.
54. Choi M, Scholl UI, Yue P, et al. K<sup>+</sup> channel mutations in adrenal aldosterone-producing adenomas and hereditary hypertension. *Science*. 2011;331:768–772. doi: 10.1126/science.1198785.
55. Beuschlein F, Boukroun S, Osswald A, et al. Somatic mutations in ATP1A1 and ATP2B3 lead to aldosterone-producing adenomas and secondary hypertension. *Nat Genet*. 2013;45:440–4, 444e1. doi: 10.1038/ng.2550.
56. Azizan EA, Poulsen H, Tuluc P, et al. Somatic mutations in ATP1A1 and CACNA1D underlie a common subtype of adrenal hypertension. *Nat Genet*. 2013;45:1055–1060. doi: 10.1038/ng.2716.
57. Scholl UI, Goh G, Stölting G, et al. Somatic and germline CACNA1D calcium channel mutations in aldosterone-producing adenomas and primary aldosteronism. *Nat Genet*. 2013;45:1050–1054. doi: 10.1038/ng.2695.
58. Scholl UI, Stölting G, Nelson-Williams C, et al. Recurrent gain of function mutation in calcium channel CACNA1H causes early-onset hypertension with primary aldosteronism. *Elife*. 2015;4:e06315. doi: 10.7554/eLife.06315.
59. Oki K, Plonczynski MW, Luis Lam M, Gomez-Sanchez EP, Gomez-Sanchez CE. Potassium channel mutant KCNJ5 T158A expression in HAC-15 cells increases aldosterone synthesis. *Endocrinology*. 2012;153:1774–1782. doi: 10.1210/en.2011-1733.
60. Hattangady NG, Karashima S, Yuan L, Ponce-Balbuena D, Jalife J, Gomez-Sanchez CE, Auchus RJ, Rainey WE, Else T. Mutated KCNJ5 activates the acute and chronic regulatory steps in aldosterone production. *J Mol Endocrinol*. 2016;57:1–11. doi: 10.1530/JME-15-0324.
61. Stindl J, Tauber P, Sterner C, Tegmeier I, Warth R, Bandulik S. Pathogenesis of adrenal aldosterone-producing adenomas carrying mutations of the Na(+)/K(+)-ATPase. *Endocrinology*. 2015;156:4582–4591. doi: 10.1210/en.2015-1466.
62. Tauber P, Aichinger B, Christ C, Stindl J, Rhayem Y, Beuschlein F, Warth R, Bandulik S. Cellular pathophysiology of an adrenal adenoma-associated mutant of the plasma membrane Ca(2+)-ATPase ATP2B3. *Endocrinology*. 2016;157:2489–2499. doi: 10.1210/en.2015-2029.
63. Xie CB, Shaikh LH, Garg S, Tanriver G, Teo AE, Zhou J, Maniero C, Zhao W, Kang S, Silverman RB, Azizan EA, Brown MJ. Regulation of aldosterone secretion by Cav1.3. *Sci Rep*. 2016;6:24697. doi: 10.1038/srep24697.
64. Reimer EN, Walenda G, Seidel E, Scholl UI. CACNA1H(M1549V) mutant calcium channel causes autonomous aldosterone production in HAC15 cells and is inhibited by mibefradil. *Endocrinology*. 2016;157:3016–3022. doi: 10.1210/en.2016-1170.
65. Fernandes-Rosa FL, Williams TA, Riester A, et al. Genetic spectrum and clinical correlates of somatic mutations in aldosterone-producing adenoma. *Hypertension*. 2014;64:354–361. doi: 10.1161/HYPERTENSIONAHA.114.03419.
66. Zheng FF, Zhu LM, Nie AF, Li XY, Lin JR, Zhang K, Chen J, Zhou WL, Shen ZJ, Zhu YC, Wang JG, Zhu DL, Gao PJ. Clinical characteristics of somatic mutations in Chinese patients with aldosterone-producing adenoma. *Hypertension*. 2015;65:622–628. doi: 10.1161/HYPERTENSIONAHA.114.03346.

67. Williams TA, Monticone S, Mulatero P. KCNJ5 mutations are the most frequent genetic alteration in primary aldosteronism. *Hypertension*. 2015;65:507–509. doi: 10.1161/HYPERTENSIONAHA.114.04636.
68. Tauber P, Penton D, Stindl J, Humberg E, Tegmeier I, Sterner C, Beuschlein F, Reincke M, Barhanin J, Bandulik S, Warth R. Pharmacology and pathophysiology of mutated KCNJ5 found in adrenal aldosterone-producing adenomas. *Endocrinology*. 2014;155:1353–1362. doi: 10.1210/en.2013-1944.
69. Williams TA, Monticone S, Schack VR, et al. Somatic ATP1A1, ATP2B3, and KCNJ5 mutations in aldosterone-producing adenomas. *Hypertension*. 2014;63:188–195. doi: 10.1161/HYPERTENSIONAHA.113.01733.
70. Tan GC, Negro G, Pinggera A, et al. Aldosterone-producing adenomas: histopathology-genotype correlation and identification of a novel CACNA1D mutation. *Hypertension*. 2017;70:129–136. doi: 10.1161/HYPERTENSIONAHA.117.09057.
71. Enberg U, Volpe C, Höög A, Wedell A, Farnebo LO, Thorén M, Hamberger B. Postoperative differentiation between unilateral adrenal adenoma and bilateral adrenal hyperplasia in primary aldosteronism by mRNA expression of the gene CYP11B2. *Eur J Endocrinol*. 2004;151:73–85.
72. Aiba M, Fujibayashi M. Alteration of subcapsular adrenocortical zonation in humans with aging: the progenitor zone predominates over the previously well-developed zona glomerulosa after 40 years of age. *J Histochem Cytochem*. 2011;59:557–564. doi: 10.1369/0022155411404071.
73. Weidmann P, De Myttenaere-Bursztejn S, Maxwell MH, de Lima J. Effect on aging on plasma renin and aldosterone in normal man. *Kidney Int*. 1975;8:325–333.
74. Crane MG, Harris JJ. Effect of aging on renin activity and aldosterone excretion. *J Lab Clin Med*. 1976;87:947–959.
75. Hegstad R, Brown RD, Jiang NS, Kao P, Weinshilboum RM, Strong C, Wisgerhof M. Aging and aldosterone. *Am J Med*. 1983;74:442–448.
76. Tsunoda K, Abe K, Goto T, Yasujima M, Sato M, Omata K, Seino M, Yoshinaga K. Effect of age on the renin-angiotensin-aldosterone system in normal subjects: simultaneous measurement of active and inactive renin, renin substrate, and aldosterone in plasma. *J Clin Endocrinol Metab*. 1986;62:384–389. doi: 10.1210/jcem-62-2-384.
77. Kerstens MN, Kobold AC, Volmer M, Koerts J, Sluiter WJ, Dullaart RP. Reference values for aldosterone-renin ratios in normotensive individuals and effect of changes in dietary sodium consumption. *Clin Chem*. 2011;57:1607–1611. doi: 10.1373/clinchem.2011.165662.
78. Mulkerrin E, Epstein FH, Clark BA. Aldosterone responses to hyperkalemia in healthy elderly humans. *J Am Soc Nephrol*. 1995;6:1459–1462.
79. Markou A, Pappa T, Kaltsas G, Gouli A, Mitsakis K, Tsounas P, Prevoli A, Tsiavos V, Papanastasiou L, Zografos G, Chrousos GP, Padiotis GP. Evidence of primary aldosteronism in a predominantly female cohort of normotensive individuals: a very high odds ratio for progression into arterial hypertension. *J Clin Endocrinol Metab*. 2013;98:1409–1416. doi: 10.1210/jc.2012-3353.
80. Baudrand R, Guarda FJ, Fardella C, Hundemer G, Brown J, Williams G, Vaidya A. Continuum of renin-independent aldosteronism in normotension. *Hypertension*. 2017;69:950–956. doi: 10.1161/HYPERTENSIONAHA.116.08952.
81. Brown JM, Robinson-Cohen C, Luque-Fernandez MA, Allison MA, Baudrand R, Ix JH, Kestenbaum B, de Boer IH, Vaidya A. The spectrum of subclinical primary aldosteronism and incident hypertension: a cohort study. *Ann Intern Med*. 2017;167:630–641. doi: 10.7326/M17-0882.
82. Vasan RS, Evans JC, Larson MG, Wilson PW, Meigs JB, Rifai N, Benjamin EJ, Levy D. Serum aldosterone and the incidence of hypertension in nonhypertensive persons. *N Engl J Med*. 2004;351:33–41. doi: 10.1056/NEJMoa033263.
83. Campino C, Martinez-Aguayo A, Baudrand R, Carvajal CA, Aglony M, Garcia H, Padilla O, Kalergis AM, Fardella CE. Age-related changes in 11 $\beta$ -hydroxysteroid dehydrogenase type 2 activity in normotensive subjects. *Am J Hypertens*. 2013;26:481–487. doi: 10.1093/ajh/hps080.

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*Hypertension*. 2018;71:218-223; originally published online December 11, 2017;  
doi: 10.1161/HYPERTENSIONAHA.117.10391

*Hypertension* is published by the American Heart Association, 7272 Greenville Avenue, Dallas, TX 75231  
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Print ISSN: 0194-911X. Online ISSN: 1524-4563

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