Angiotensin II Relaxations of Bovine Adrenal Cortical Arteries
Role of Angiotensin II Metabolites and Endothelial Nitric Oxide

Kathryn M. Gauthier, David X. Zhang, Lijie Cui, Kasem Nithipatikom, William B. Campbell

Abstract—Angiotensin (Ang) II regulates adrenal steroidogenesis and adrenal cortical arterial tone. Vascular metabolism could decrease Ang II concentrations and produce metabolites with vascular activity. Our goals were to study adrenal artery Ang II metabolism and to characterize metabolite vascular activity. Bovine adrenal cortical arteries were incubated with Ang II (100 nmol/L) for 10 and 30 minutes. Metabolites were analyzed by mass spectrometry. Ang (1-7), Ang III, and Ang IV concentrations were 146±21, 173±42 and 58±11 pg/mg at 10 minutes and 845±163, 70±14, and 31±3 pg/mg at 30 minutes, respectively. Concentration-related relaxations of U46619-preconstricted cortical arteries to Ang II (maximum relaxation=29±3%; EC50=3.4 pmol/L) were eliminated by endothelium removal and inhibited by the NO synthase inhibitor, nitro-L-arginine (30 μmol/L; maximum relaxation=14±7%). Ang II relaxations were enhanced by the angiotensin type-1 receptor antagonist losartan (1 μmol/L; maximum relaxation=41±3%; EC50=11 pmol/L). Losartan-enhanced Ang II relaxations were inhibited by nitro-L-arginine (maximum relaxation=18±5%) and the angiotensin type-2 receptor antagonist PD123319 (10 μmol/L; maximum relaxation=27±5%). Ang (1-7) and Ang III caused concentration-related relaxations with less potency (EC50=43 and 24 nmol/L, respectively) but similar efficacy (maximum relaxations=39±3% and 48±5%, respectively) as losartan-enhanced Ang II relaxations. Ang (1-7) relaxations were inhibited by nitro-L-arginine (maximum relaxation=16±4%) and the Ang (1-7) receptor antagonist 7-D-Ala-Ang (1-7) (1 μmol/L; maximum relaxation=10±3%) and eliminated by endothelium removal. Thus, Ang II metabolism by adrenal cortical arteries to metabolites with decreased vascular activity represents an inactivation pathway possibly decreasing Ang II presentation to adrenal steroidogenic cells and limits Ang II vascular effects. (Hypertension. 2008;52:1-6.)

Key Words: angiotensin (1-7) ■ angiotensin III ■ angiotensin IV ■ mass spectrometry

Angiotensin (Ang) II is a major regulator of vascular resistance, adrenal steroid hormone release, electrolyte and blood volume, and systemic blood pressure (see reviews in References 1–5). Ang II is produced by the conversion of Ang I to Ang II by Ang-converting enzyme (ACE). Numerous cardiovascular diseases, including hypertension and heart failure, have been linked to increased activity of the renin-Ang system.4,5 Within the past decade, increasing attention has focused on the role of the Ang heptapeptide metabolite Ang (1-7). Ang (1-7) is produced primarily through the action of the ACE homologue ACE2.5–9 ACE2 removes the carboxy-terminal phenylalanine of Ang II to produce Ang (1-7). Other enzymes may convert Ang II to Ang(1-7), including nephrilysin,9 but their roles in the physiological production in Ang (1-7) have not been clearly defined.

Ang (1-7) has numerous biological effects, including vasorelaxation at low concentrations, vasoconstriction at high concentrations, blockade of Ang II constriction, inhibition of ACE, alteration of cardiac output, ischemic cardiac protection, and antifibrotic and proliferative effects (see reviews in References 2, 5, 7, 10, and 11). Thus, Ang (1-7) displays a myriad of cardiovascular protective properties. The biological effects of the Ang II metabolites Ang III and Ang IV have not been as thoroughly examined. However, Ang III shares many biological activities as Ang II, including vasodilation.11,12 Similarly, Ang IV has been shown to cause vascular dilation11 and increases cell proliferation.13

Adrenal blood flow is closely coupled to steroidogenesis. Increases in adrenal flow alone can stimulate steroid hormone release.14 Increased vascular resistance of adrenal cortical arteries restricts blood flow and decreases adrenal hormone release. In isolated bovine small cortical arteries, Ang II causes a biphasic response on vascular diameter. At low concentrations, Ang II causes dilations that reverse to constriction at higher concentrations (>10 nmol/L).15 Changes in circulating Ang II concentrations or local metabolism of Ang

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II could alter cortical arterial tone and cortical blood flow to alter adrenal hormone release. In addition, cortical metabolism of Ang II could limit Ang II presentation to adrenal steroidogenic cells.

Previously, we demonstrated that adrenal zona glomerulosa cells and adrenal vascular endothelial cells metabolize Ang II to several smaller peptides, including Ang III, Ang IV, and Ang (1-7). Although Ang III is equipotent to Ang II in stimulating steroidogenesis, Ang IV and Ang (1-7) are inactive. Ang II metabolism by intact adrenal arteries has not been evaluated, and, thus, the consequence of this metabolism on adrenal arterial vascular tone has not been characterized.

Methods

Isometric Tension Recording
Fresh bovine adrenal glands were obtained from a local abattoir. Small cortical arteries closely attached to the adrenal surface (200 to 300 μm) were dissected in HEPES buffer (in mmol/L: HEPES 10, NaCl 150, KCl 5, CaCl2 2, MgCl21, and glucose 6 [pH 7.4]), cleaned of connective tissue, and mounted in a 4-chamber wire myograph (model 610M, Danish Myo Technology A/S), as described previously. Arteries were maintained at 37°C in physiological saline solution (in mmol/L: NaCl 119, KCl 4.7, CaCl2 2.5, MgSO4 1.17, NaHCO3 24, KH2PO4 1.18, EDTA 0.026, and glucose 5.5), gassed with 95% O2/5% CO2. Arteries were stretched to a resting tension of 1 millinewton and stimulated 2 to 3 times with KCl (60 mmol/L) plus the thromboxane mimetic U46619 (100 nmol/L) for 10 minutes at 10-minute intervals. Arteries were contracted with submaximal concentrations of U46619 (30 to 300 nmol/L) to 50% to 75% of their maximum KCl and U46619 challenge. Where indicated, the endothelium was removed by gently rubbing the arterial intimal surface with a human hair. The endothelium was considered intact if acetylcholine (1 μmol/L) caused >50% relaxation and was effectively removed if acetylcholine induced <10% relaxation. Cumulative concentration responses to Ang II (0.1 pmol/L to 10 nmol/L), Ang III (10 pmol/L to 10 μmol/L), and Ang (1-7) (10 pmol/L to 10 μmol/L) were performed. Responses were repeated in arterial segments pretreated with the Ang II type 1 (AT1) receptor antagonist losartan (1 μmol/L), the Ang II type 2 (AT2) receptor antagonist PD123319 (1 to 10 μmol/L), the Ang (1-7) receptor antagonist (13C5)-Ala7-Ang (1-7) (Ala7-Ang [1-7]; 1 μmol/L), or the NO synthase inhibitor nitro-L-arginine (L-NA; 30 μmol/L), either separately or with losartan. Results are expressed as the percentage of relaxation of the U46619-treated arteries, with 100% representing basal tension.

Ang Incubations and Extractions
Adrenal cortical arteries (35-53 mg tissue; wet weight/incubation) were incubated in 2 mL of HEPES buffer at 37°C with Ang II (100 nmol/L) for 10 and 30 minutes. The medium was removed, and an internal standard (15N1-N2-Ang IV, 2.0 ng) was added. Samples were extracted as described previously. Briefly, formic acid was added (final concentration of 0.5%), and the samples were applied to Waters C18 SPE cartridges that had been preconditioned with 5 mL of ethanol and 15 mL of deionized water, respectively. The cartridges were washed with deionized water (3 mL), and the columns were dried by vacuum. Ang peptides were eluted with 7 mL of methanol containing 5% formic acid and dried under a stream of nitrogen gas. For liquid chromatography-mass spectrometry (LC-MS) and liquid chromatography-tandem mass spectrometry (LC-MS/MS) analysis preparation, samples were redissolved in 30 μL of 16% acetonitrile in water containing 0.1% formic acid.

LC-MS and LC-MS/MS Analyses
Analyses were performed using a Waters-Micromass Quattro micro API electrospray triple quadrupole mass spectrometric system coupled with a Waters 2695 high-performance liquid chromatograph, as described previously. The mass spectrometer is equipped with a Z-spray dual orthogonal ionization source and is controlled by MassLynx 4.0 software (Waters Corporation). Samples were separated on a reverse-phase C18 column (Jupiter 2.0×250 mm, Phenomenex) using water-acetonitrile with 0.1% formic acid as a mobile phase at a flow rate of 0.2 mL/min at ambient room temperature. The mobile phase of 16% acetonitrile in water linearly increased to 38% acetonitrile over 10 minutes, then to 100% acetonitrile over the final 15 minutes. Positive ion electrospray ionization mass spectrometric conditions were as follows: capillary voltage, 3.5 kV; cone voltage, 15 to 18 V; desolvation temperature, 300°C; desolvation gas flow, 1000 L/h; and source temperature, 120°C. LC-MS analysis was performed in positive electrospray ionization mode in the single-ion recording mode. We used m/z of 300.7, 311.8, 350.0, 388.8, and 391.3 for Ang (1-7), Ang III, Ang II, Ang IV, and the internal standard, respectively. LC-MS/MS was performed with multiple reaction monitoring of transitions: m/z 300.6→109.6 (Ang [1-7]), m/z 311.3→228.4 (Ang III), m/z 349.6→255.2 (Ang II), m/z 388.8→263.4 (Ang IV), and m/z 391.3→269.2 (internal standard). Argon gas was used as collision gas, and the collision voltages were set at 27 to 30 eV. Standard calibration curves were constructed over the range of 5 to 625 pg for LC-MS and 50 to 1250 pg for LC-MS/MS per injection. Sample Ang peptide concentrations were determined by comparing ratios of the peak areas to the standard calibrations.

Drugs and Chemicals
Ang (1-7), L-NA, PD123319, and indomethacin were purchased from Sigma. U46619 was obtained from Cayman Chemical Company. Ang II, Ang III, Ang IV, and Ala7-Ang (1-7) were purchased from Bachem. Losartan was a kind gift from Merck. The 15N1-N2-Ang IV internal standard was synthesized by the Medical College of Wisconsin Protein Nucleic Acid Facility. All of the solvents were high-performance liquid chromatography grade and purchased from Burdick and Jackson.

Data Analysis
Data are presented as means±SEMs. Significant differences between mean values were evaluated by Student’s t test or ANOVA followed by the Student-Newman-Keuls multiple comparison test. EC50 values were calculated using GraphPad Prism analysis software (GraphPad Software). A value of P<0.05 was considered statistically significant.

Results

Role of the Endothelium in Ang II Relaxations
Small adrenal cortical arteries were contracted with the thromboxane mimetic U46619, and responses to increasing Ang II concentrations were determined (Figure 1). Ang II caused concentration-dependent relaxations (maximum relaxation=28±5% at 10 nmol/L). The relaxations were eliminated by endothelium removal. Thus, Ang II causes vascular relaxation through endothelium-dependent mechanisms. Relaxations to Ang II could occur through endothelial cell Ang II metabolism to vasoactive peptides and/or through direct stimulation of endothelial cell relaxing factor release.

Identification and Quantification of Ang II Metabolites by Adrenal Cortical Arteries
To clarify the role of metabolites in Ang II relaxations, we examined Ang II metabolism by freshly isolated adrenal cortical arteries. Cortical arteries were incubated with Ang II (100 nmol/L) for 30 minutes. The buffer was removed, extracted, and analyzed for Ang peptides by LC-MS/MS. The LC-MS/MS chromatogram in Figure 2 shows recovery of the internal standard and the presence of Ang IV, Ang II, Ang III,
and Ang (1-7). Abundances of the internal standard, Ang IV, Ang III, and Ang (1-7) were amplified 10 times for the demonstration of the relative production of the peptides. The predominant metabolite was Ang (1-7). LC-MS scan mode analysis of the Ang III and Ang (1-7) sample peptides showed strong (M + 3H)\(^+\) ions at \(m/z\) 300.7 and 350.0, respectively, and (M + 2H)\(^+\) ions at \(m/z\) 524.6 and 450.5, respectively (data not shown). These results are consistent with the mass spectra for Ang III and Ang (1-7).\(^17\)

Role of NO and AT\(_1\) and Ang (1-7) Receptors in Ang II Relaxations

Ang II relaxations were significantly enhanced by the AT\(_1\) receptor blocker losartan (1 \(\mu\)mol/L; maximum relaxation = 46±4\% at 10 \(\mu\)mol/L; Figure 4A). Ang II relaxations and the losartan-enhanced relaxations to Ang II were inhibited by the NO synthase inhibitor L-NA (30 \(\mu\)mol/L; maximum relaxation = 15±7\% and 18±5\%, respectively; Figure 4A and 4B). The Ang (1-7) receptor antagonist Ala\(7\)-Ang (1-7) (1 \(\mu\)mol/L) was without effect (Figure 4C). The AT2 receptor antagonist PD123319 inhibited the losartan-enhanced relaxations in a concentration-dependent manner (Figure 4D). Significant inhibition was observed at 10 \(\mu\)mol/L of PD123319. These results suggest that Ang II relaxations are mediated in part by AT\(_2\) receptor activation and are independent of AT\(_1\) and Ang (1-7) receptors. Conversely, AT\(_1\) receptor activation causes vascular constriction. In addition, Ang II relaxations are mediated by endothelial cell NO.

Ang III and Ang (1-7) Relaxations

Next, we evaluated the relaxation responses of the adrenal cortical arteries to Ang III and Ang (1-7). Ang III caused concentration-related relaxations with maximal relaxations of 48±5\% at 10 \(\mu\)mol/L (Figure 5). Similar relaxations were...
observed with Ang (1-7) (maximum relaxation of 44±4%). The averaged half-maximal effective concentrations (EC$_{50}$s) and averaged maximal relaxations of Ang II and the Ang metabolites are provided in the Table. Relaxations to Ang III and Ang (1-7) displayed similar efficacy as the losartan-enhanced Ang II relaxations but were less potent. This suggests that metabolism of Ang II to Ang III or Ang (1-7) represents an inactivation pathway to possibly limit Ang II relaxations.

Because the predominant Ang II metabolite was Ang (1-7), we further investigated the mechanisms of Ang (1-7) relaxations. Ang (1-7)-induced relaxations were eliminated by endothelial cell removal and inhibited by the NO synthase inhibitor L-NA (maximum relaxation = 16±4%; Figure 6A). Inhibition of AT$_1$ and AT$_2$ receptors with losartan or PD123319, respectively, did not alter the Ang (1-7) relaxations, whereas the Ang (1-7) receptor antagonist Ala7-Ang (1-7) caused significant inhibition (maximum relaxation=10±3%; Figure 6B). Thus, Ang (1-7) relaxations occur through the activation of Ang (1-7) receptors, and, similar to Ang II, the relaxations are mediated by endothelial cell NO.

**Discussion**

This is the first study to demonstrate Ang II metabolism in intact arteries. Importantly, these studies were performed in small resistance-sized adrenal cortical arteries, which are primary sites for the regulation of adrenal cortical blood flow. The physiological implications for Ang II metabolism at this site are 2-fold. First, because Ang II causes dynamic changes in vascular tone of these arteries (ie, dilations at low concentrations and constrictions at high concentrations), subtle changes in Ang II concentrations by vascular metabolism could alter vascular resistance and, therefore, adrenal cortical blood flow. Second, Ang II metabolism would decrease Ang II concentrations and increase metabolite concentrations presented to the steroidogenic cells. This alteration in Ang II:metabolite ratios could alter aldosterone release and, secondarily, blood volume and blood pressure.

In bovine adrenal cortical arteries, relaxations to Ang II were inhibited by the removal of the endothelium. However, it was not evident whether the relaxations occur through Ang II metabolism to vasoactive peptides that activate relaxing factor release or through the direct action of Ang II. Ang II

<table>
<thead>
<tr>
<th>Angiostatin Treatment</th>
<th>Ang II (Losartan Treated)</th>
<th>Ang III</th>
<th>Ang (1-7)</th>
</tr>
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<tbody>
<tr>
<td>Maximum relaxation</td>
<td>29±3%</td>
<td>41±3%</td>
<td>48±5%</td>
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<tr>
<td>relaxation</td>
<td>(10 nmol/L)</td>
<td>(100 nmol/L)</td>
<td>(10 μmol/L)</td>
</tr>
<tr>
<td>EC$_{50}$</td>
<td>3.4 pmol/L</td>
<td>11 pmol/L</td>
<td>24 nmol/L</td>
</tr>
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</table>

**Figure 4.** Ang II–induced relaxations of isolated bovine adrenal cortical arteries. A, Effect of the NO synthase inhibitor L-NA (30 μmol/L) and the AT$_1$ receptor antagonist losartan (1 μmol/L). Relaxations were performed under control conditions or after treatment with losartan or L-NA. B, Effect of L-NA on losartan-enhanced Ang II relaxations. Relaxations were performed on losartan-treated arteries before and after treatment with L-NA. C, Effect of the Ang (1-7) receptor antagonist Ala7-Ang (1-7) (1 μmol/L) on losartan-enhanced Ang II relaxations. Relaxations were performed on losartan-treated arteries before and after treatment with Ala7-Ang (1-7). D, Effect of the AT$_2$ receptor antagonist PD123319 (1 and 10 μmol/L) on losartan-enhanced Ang II relaxations. Relaxations were performed on losartan-treated arteries before and after treatment with PD123319. n=5 to 34; *P<0.05 vs control or losartan.

**Figure 5.** Concentration-dependent relaxations to Ang (1-7) and Ang III of isolated bovine adrenal cortical arteries. n=7 to 10.
metabolism to Ang III, Ang IV, and Ang (1-7) was demonstrated in endothelial cells from bovine adrenal arteries and human aorta and human umbilical veins. The contribution of the smooth muscle to the Ang II metabolism has not been clarified, although ACE2 transcriptional expression has been demonstrated in human coronary, cerebral, pulmonary, renal, and mesenteric arteries. In the intact adrenal cortical arteries from this study, metabolism of Ang II to Ang (1-7) does not appear to contribute to the vascular relaxations to Ang II. This was evident because the Ang (1-7) receptor antagonist did not alter the relaxations to Ang II. In contrast, in rat carotid arteries, Ang II relaxations are partially dependent on Ang II metabolism to Ang (1-7). Thus, it appears that the role of Ang metabolites in Ang II relaxation varies with the vascular lature and/or species.

Ang II relaxations were mediated by endothelial cell NO. This was demonstrated in the current study and a previous study of perfused bovine adrenal cortical arteries by the inhibition of the Ang II relaxations/dilations by L-NA. As reviewed by Toda et al., the role of the endothelium and NO production in Ang II and Ang (1-7) relaxations has also been demonstrated in numerous arteries from various species. Thus, our results fit the paradigm of NO as the primary effector of Ang II relaxations. Similarly, adrenal cortical arterial relaxations to Ang (1-7) were mediated by endothelial NO production. Because the relaxations induced by Ang (1-7) were blocked by the Ang (1-7) receptor antagonist, we conclude that this receptor activates endothelial cell signaling cascades that culminate with NO release.

Relaxations to Ang III and Ang (1-7) were >1000-fold less potent than Ang II relaxations. Therefore, Ang II metabolism to these metabolites should not significantly contribute to Ang II relaxations. However, Ang II metabolism and the impact on adrenal arterial vascular tone may be more important during times of high Ang II. Under this scenario, Ang II vascular constriction would overcome Ang II relaxation. Because Ang III and Ang (1-7) did not cause vascular constriction in the adrenal arteries, their relaxing properties could counterbalance Ang II constrictions. Alternatively, high concentrations of Ang (1-7) infusion in humans reduced forearm blood flow and in rats reduced blood flow to the kidney, mesentery, and skin.

The consequence of cortical artery Ang II metabolism on the concentrations of Ang II and Ang II metabolites that reach the adrenal steroidogenic cells depends on the steroidogenic activity of the metabolites. In this regard, Ang III was equipotent to Ang II in stimulating aldosterone and corticosterone production in conscious rats and aldosterone production from rat adrenal cortical cells. Alternatively, Ang IV and Ang (1-7) did not stimulate aldosterone production in bovine adrenal zona glomerulosa cells. Because the primary Ang II metabolite from the cortical arteries was Ang (1-7), we predict that vascular Ang II metabolism would reduce Ang II stimulation of steroid hormone release.

The role of Ang (1-7) as an antihypertensive peptide has not been demonstrated. In the kidney, the effects of Ang (1-7) on natriuresis and diuresis have been mixed. Therefore, other mechanisms must counteract the direct vasoconstrictory effects of Ang (1-7). Adrenal mechanisms, including an increase in blood flow and the consequent increase in aldosterone release, could compensate for the systemic vascular actions of Ang (1-7). Further studies will be required to address this possibility.

Perspectives

The regulation of adrenal blood flow is multifaceted and includes neural and humoral input. An additional level of regulation results from the interchange and cross-talk between the adrenal vasculature and steroidogenic cells. Relaxing factor(s) released by the cortical steroidogenic cells influence cortical arterial tone. Conversely, factors originating from the adrenal vasculature influence adrenal steroidogenesis. Endothelial cell NO inhibits aldosterone secretion, whereas an endothelium-derived peptide stimulates aldosterone production. The results from the current study suggest that vascular Ang II metabolism could influence the level of Ang II-dependent regulation of vascular tone and modulate the concentrations of Ang II available for the stimulation of steroidogenesis.

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Disclosures

None.

References


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